www.blackwellmunksgaard.com/jopm

REVIEW ARTICLE

Molecular pathology of odontogenic tumors

H. Kumamoto

Division of Oral Pathology, Department of Oral Medicine and Surgery, Tohoku University Graduate School of Dentistry, Sendai, Japan

Odontogenic tumors are lesions derived from the elements of the tooth-forming apparatus and are found exclusively within the jawbones. This review represents a contemporary outline of our current understanding of the molecular and genetic alterations associated with the development and progression of odontogenic tumors, including oncogenes, tumor-suppressor genes, oncoviruses, growth factors, telomerase, cell cycle regulators, apoptosis-related factors, regulators of tooth development, hard tissue-related proteins, cell adhesion molecules, matrix-degrading proteinases, angiogenic factors, and osteolytic cytokines. It is hoped that better understanding of related molecular mechanisms will help to predict the course of odontogenic tumors and lead to the development of new therapeutic concepts for their management.

J Oral Pathol Med (2006) 35: 65-74

Keywords: molecular alteration; odontogenic tumor

Introduction

Odontogenic tumors are lesions derived from the epithelial and/or mesenchymal elements of the tooth-forming apparatus and are therefore found exclusively within the jawbones (1–3). These development-associated tumors (i) often occur in children or young adults and exhibit considerable histologic variation (Table 1), (ii) are usually intraosseous tumors that contain various amounts of epithelial components and interact with their specific microenvironment, and (iii) are generally benign tumors, but several odontogenic tumors show locally invasive behavior with a high risk of recurrence (1-6).

A series of genetic and molecular alterations appear to promote the development and progression of tumors via multiple steps (7–10). Although the etiology and pathogenesis of odontogenic tumors remain unknown, recent studies have identified various molecular alterations responsible for their development and progression (Table 2). This review provides a contemporary outline of our understanding of the molecular and genetic events associated with odontogenic tumors.

Molecules involved in tumorigenesis and/or cell differentiation of odontogenic tumors

Oncogenes

Oncogenes are normal cellular genes that contribute to neoplastic transformation by functions activated by gene amplification, translocation, or mutation (11, 12). Many oncogenes have been identified, and their gene products function as growth factors (platelet-derived growth factor (PDGF), fibroblast growth factor (FGF)), growth factor receptors [epidermal growth factor receptor (EGFR), HER-2, Ret], non-receptor tyrosine kinases (Src, Abl), serine/threonine kinases (Mos), signal transducers (Ras), and transcription factors (Myc, Fos). These factors participate in cellular functions related to proliferation and differentiation (11–16).

Ras oncogene was originally characterized on homology studies with rat-transforming gene, and three *Ras* genes, H-*Ras*, K-*Ras*, and N-*Ras*, have been identified in the mammalian genome (15). Products encoded by *Ras* genes, $p21^{Ras}$, are involved in the transduction of external stimuli most likely induced by growth factors (15–17). In ameloblastomas, ameloblastic fibromas, and odontogenic myxomas, $p21^{Ras}$ is expressed preferentially in odontogenic epithelial cells; overexpression of $p21^{Ras}$ is found in odontogenic tumors when compared with normal developing teeth (18). Point mutations of *Ras* genes have been identified in various tumors, including pancreatic, colorectal, bladder, and lung carcinomas (7, 11, 15). A K-*Ras* mutation at codon 12 has been detected in one of 23 ameloblastomas (19).

c-Myc oncogene encodes a nuclear phosphoprotein that regulates cellular proliferation (11, 13). Gene amplification and/or protein overexpression of this gene have been identified in many types of tumors (13, 20). In ameloblastomas, c-Myc oncoprotein is expressed predominantly in neoplastic cells neighboring the basement membrane (21). Transgenic mice carrying Myc and/or H-*Ras* gene have a high incidence of various odontogenic tumors (22–24).

Correspondence: Hiroyuki Kumamoto, Division of Oral Pathology, Department of Oral Medicine and Surgery, Tohoku University Graduate School of Dentistry, 4-1 Seiryo-machi, Aoba-ku, Sendai 980-8575, Japan. Tel: +81-22-717-8303. Fax: +81-22-717-8304. E-mail: kumamoto@mail.tains.tohoku.ac.jp Accepted for publication August 25, 2005

 Table 1
 Histologic classification of odontogenic tumors

Benign	
Epithelial tumors	
Ameloblastoma	
Squamous odontogenic tumor	
Calcifying epithelial odontogenic tumor (Pindborg tumor)	
Adenomatoid odontogenic tumor	
Calcifying odontogenic cyst/Odontogenic ghost cell tumor	
Mixed tumors	
Ameloblastic fibroma/Ameloblastic fibro-odontoma	
Odontoameloblastoma	
Odontoma	
Mesenchymal tumors	
Odontogenic fibroma	
Odontogenic myxoma	
Cementoblastoma	
Malignant	
Odontogenic carcinomas	
Malignant ameloblastoma	
Metastasizing ameloblastoma	
Ameloblastic carcinoma	
Clear cell odontogenic carcinoma	
Odontogenic ghost cell carcinoma	
Primary intraosseous carcinoma	
de novo	
cystogenic	
Odontogenic sarcomas	
Ameloblastic fibrosarcoma	
Ameloblastic fibro-odontosarcoma	
Odontogenic carcinosarcoma	
Related lesions	
Cemento-ossifying fibroma	
Cemento-osseous dysplasia	

Heikinheimo et al. have reported overexpression of *Fos* oncogene, which encodes a transcription factor participating in the control of cell proliferation and differentiation, in ameloblastomas on cDNA microarray and subsequent real-time reverse transcriptase polymerase chain reaction (RT-PCR; 25). These findings suggest that these oncogenes play a role in the pathogenesis of odontogenic tumors via dysregulation of cell proliferation.

Tumor-suppressor genes

Tumor-suppressor genes normally act as regulators of cell growth, and inactivation of these genes by mutations and/or loss of heterozygosity (LOH) in both alleles results in tumor development (11, 26–28). Several tumor-suppressor genes have been identified; *retinoblastoma* (*RB*), *p53*, *adenomatous polyposis coli* (*APC*), *WT-1*, and *patched* (*PTC*) genes are well known (27, 28).

p53 gene is one of the most frequently altered genes in tumors, and its gene product plays an important role in response to genomic damage by inducing cell cycle arrest or apoptosis (7, 29, 30). Increased immunohistochemical reactivity for p53 has been detected in ameloblastomas, malignant ameloblastomas, primary intraosseous carcinomas, and ameloblastic fibrosarcomas (31-36), although several studies have shown that p53 mutations are infrequent in ameloblastomas (34, 36–38). Regulators of p53, MDM2, and p14^{ARF}, are also expressed in ameloblastomas, adenomatoid odontogenic tumors, malignant ameloblastomas, and clear cell odontogenic carcinoma; overexpression of MDM2 and p14^{ARF} has been detected in ameloblastomas and malignant odontogenic tumors (34, 36, 39). Recently, p53 homologs, p63 and p73, have been analyzed by immunohistochemistry and RT-PCR, suggesting that these homologs function differently from p53 in odontogenic tissues (40).

APC gene was discovered on genetic analysis of families with familial adenomatous polyposis (FAP), and subsequent studies revealed that its gene product downregulates Wnt signaling pathway, inhibiting cell proliferation (27, 41, 42). While germline *APC* mutations are responsible for FAP, somatic mutations lead to the development of sporadic colorectal tumors (7, 42). Immunohistochemical reactivity for *APC* is lower in benign and malignant ameloblastomas than in tooth germs (43).

Retinoblastoma gene, the first tumor-suppressor gene to be isolated, codes for a nuclear protein controlling

Molecules possibly associated with tumorigenesis and/or tumor cell differentiation Oncogenes	Ras, Myc, Fos
e	
Tumor-suppressor genes	p53, APC
Oncoviruses	HPV, EBV
Growth factors	TGF-α, -β, FGF-1, -2, HGF
Telomerase	Telomerase
Cell cycle regulators	Cyclin D1, p16 ^{INK4a} , p21 ^{WAF1/Cip1} , p27 ^{Kip1}
Apoptosis-related factors	Bcl-2 family, IAP family, Fas, TNF-α, p53
Regulators of tooth development	SHH pathway, Wnt pathway
Hard tissue-related proteins	Amelogenin, Enamelin, Ameloblastin, Enamelysin,
	BSP, Osteonectin, Osteocalcin, Osteopontin, BMP
Molecules possibly associated with tumor progression	-
Cell adhesion molecules	E-selectin, ICAM-1, VCAM-1, E-cadherin, Integrins, CD44
Matrix-degrading proteinases	MMP-1, -2, -9/TIMP-1, -2, Heparanase
Angiogenic factors	VEGF
Osteolytic cytokines	IL-1, -6, TNF-a, PTHrP, RANKL/OPG

HPV, human papilloma virus; EBV, Epstein–Barr virus; TGF, transforming growth factor; FGF, fibroblast growth factor; HGF, hepatocyte growth factor; IAP, inhibitor of apoptosis protein; TNF, tumor necrosis factor; SHH, Sonic hedgehog; BSP, bone sialoprotein; BMP, bone morphogenetic protein; ICAM, intercellular adhesion molecule; VCAM, vascular cell adhesion molecule; MMP, matrix metalloproteinase; TIMP, tissue inhibitor of metalloproteinases; VEGF, vascular endothelial growth factor; IL, interleukin; PTHrP, parathyroid hormone-related protein; RANKL, receptor activator of nuclear factor-κB ligand; OPG, osteoprotegerin.

cell cycle progression together with a transcription factor, E2F (27). Although RB has not been investigated in odontogenic tumors, transgenic mice expressing E2F-1gene under the control of a keratin 5 promoter have been shown to develop skin tumors and ameloblastomas (44). These features suggest that these tumor-suppressor genes are involved in the development of odontogenic tumors via aberrant control of cell proliferation.

DNA-repair genes

Errors during DNA replication or repair are maintained by DNA-repair genes, such as *MSH2*, *MLH1*, and *PMS1* (45, 46). Aberrations in these genes cause genetic instability, occurring not only in hereditary non-polyposis colorectal cancer (HNPCC) but also in sporadic tumors of the colon, rectum, stomach, pancreas, and endometrium (45–47). In ameloblastomas, representative DNA-repair gene products, MSH2 and MLH1, are immunohistochemically expressed chiefly in peripheral neoplastic cells, suggesting that the development or progression of the odontogenic tumors does not depend on a defect of the DNA maintenance system (48).

Oncoviruses

Many DNA and RNA viruses are oncogenic in a wide variety of animals, and increasing evidence suggests that certain types of human tumors are caused by viruses, such as human papilloma virus (HPV), Epstein-Barr virus (EBV), and human T-cell leukemia virus type 1 (HTLV-1; 49). Approximately 70 genetically distinct types of HPV have been identified, and some types cause squamous papilloma of the skin and squamous cell carcinoma of the cervix and upper respiratory and digestive tracts (49, 50). EBV has been implicated in the pathogenesis of Burkitt lymphoma, Hodgkin's lymphoma, nasopharyngeal carcinoma, and gastric medullary carcinoma (49, 51, 52). Several investigators have reported HPV and EBV infections in ameloblastomas; however, the etiologic roles of these viruses in odontogenic tumors remain controversial (53-58).

Growth factors

Growth factors are hormone-like polypeptides that play key roles in the control of cell proliferation and differentiation, and many growth factors have been identified, including epidermal (EGF), platelet-derived (PDGF), insulin-like (IGF), hepatocyte (HGF), fibroblastic (FGF), vascular endothelial (VEGF), and transforming (TGF) growth factors (59). Growth factors transmit signals by binding to specific high-affinity cell surface receptors and involving subsequent signaling molecules (17, 60). Dysfunction of growth factors, growth factor receptors, or signaling components results in pathologic conditions, including neoplasia (17, 59, 60).

EGF and TGF- α regulate cell proliferation and functional maturation in a wide range of tissues through their specific receptor, EGFR (59, 61, 62). In tooth germs and various odontogenic tumors, expression of EGF, TGF- α , and EGFR is mostly located in odontogenic epithelial cells (62–64). Odontogenic tumors express TGF- α but not EGF, whereas expression of both EGF and TGF- α has been detected in tooth germs (62). EGFR expression in ameloblastomas is higher than that in epithelial elements of radicular cysts and granulomas (63). These features suggest that TGF- α and EGFR are involved in odontogenic tumorigenesis.

TGF- β is a multifunctional growth factor involved in the control of cell growth, cell differentiation, cell migration, and the synthesis and degradation of extracellular matrix (59, 60). TGF- β and its receptors (TGF- β receptor types I and II) are expressed in many types of odontogenic tumors, suggesting that they have an important role in cell differentiation and matrix formation via regulation or dysregulation of epithelial-mesenchymal interactions (65–68).

HGF has mitogenic, motogenic, and morphogenic functions in various types of cells and acts via its receptor, c-Met (69). This growth factor is essential for morphogenesis of tooth germs (70). HGF and c-Met are expressed in many types of odontogenic tumors, suggesting that they affect epithelial-mesenchymal interactions not only in developing teeth but also in neoplastic odontogenic tissues (67, 68). Increased expression of HGF and c-Met in ameloblastic carcinomas and clear cell odontogenic carcinomas implies that HGF signaling might be associated with the malignant potential of epithelial odontogenic tumors (67).

FGF is widely distributed and functions in the growth, differentiation, and regeneration of a variety of tissues (71). Several types of FGF are also involved in tooth formation (72, 73). Expression of FGF-1, -2, and their types 2 and 3 receptors has been detected in various odontogenic tumors, suggesting that these molecules are associated with odontogenic differentiation rather than pathogenesis (74, 75).

Ras/MAPK and PI3K/Akt pathways function downstream to growth factor receptors and regulate cell proliferation and differentiation (15, 17). Alterations of involved signaling molecules have been identified in a variety of human tumors (17, 76, 77). In ameloblastomas, Ras/MAPK and PI3K/Akt pathways have been examined, but no distinct aberrations of the signaling molecules have been detected (19, 78).

Telomerase

Telomerase is a specialized reverse transcriptase that synthesizes telomeric DNA at the ends of chromosomes and compensates for its loss with each cell division, participating in cell immortalization (79). Activation of telomerase has been demonstrated in most human malignancies (79, 80). On telomeric repeat amplification protocol (TRAP) assay, ameloblastoma tissues have been consistently positive for telomerase activity, suggesting that telomerase activation is associated with tumorigenesis of odontogenic epithelium (21, 81). Immunohistochemical reactivity for telomerase in ameloblastomas shows a similar distribution pattern to that of c-Myc protein, an oncogene product that directly activates telomerase transcription, suggesting that this oncoprotein induces telomerase activity in ameloblastomas (21).

Cell cycle regulators

68

Cell proliferation follows an orderly progression through the cell cycle, which is governed by numerous factors, including cyclins, cyclin-dependent kinases (CDKs), CDK inhibitors (CKIs), and other critical regulators (82, 83). Uncontrolled cell cycles caused by aberrations of these regulators have been identified in a variety of tumors (82, 84). In many types of odontogenic tumors, cell cycle phase/cell proliferation markers, proliferating cell nuclear antigen (PCNA), Ki-67, DNA topoisomerase IIa, and histone H3, reflect proliferation activity and malignant potential (31, 33, 68, 85-92). Cyclin D1 is strongly implicated in cell cycle progression by regulating the transition from G1 to S phase, while CKIs, p16^{INK4a}, p21^{WAF1/Cip1}, and p27^{Kip1} inhibit cell cycle progression by suppressing cyclin D1 function (83). In ameloblastomas, expression of cyclin D1, $p16^{INK4a}$, $p21^{WAF1/Cip1}$, and $p27^{Kip1}$ is well preserved when compared with tooth germs, suggesting that the proliferation of odontogenic epithelial cells is strictly controlled by these cell cycle regulators (91).

Apoptosis-related factors

Apoptosis, also known as programmed cell death or physiologic cell death, has diverse roles in embryogenesis and normal homeostasis, as well as in a variety of pathologic conditions (93, 94). There are two alternative apoptotic pathways, one mediated by death receptors and the other by mitochondria, and apoptotic processes are modulated by a large family of genes, such as the tumor necrosis factor (TNF) family, the Bcl-2 family, and the inhibitor of apoptosis protein (IAP) family (95–100). Evasion from apoptosis by aberrations of apoptosis regulatory factors has been found to cause the accumulation of neoplastic cells in various tumors (94–96, 98).

In some epithelial odontogenic tumors, apoptotic cells have been detected by TdT-mediated dUTP-biotin nick end-labeling (TUNEL) and immunohistochemistry using single-stranded DNA (ssDNA) antibody, suggesting that apoptotic cell death plays an important role in oncogenesis and cell differentiation in odontogenic epithelium (33, 101-104). Representative death receptors, Fas, TNF receptor I (TNFRI), and TNF-related apoptosis-related ligand (TRAIL) receptors 1 and 2, have been recognized in benign and malignant ameloblastomas, but expression of caspase-8, an apoptosis initiator in the death receptor-mediated apoptotic pathway, is extremely limited, suggesting that apoptotic cell death in odontogenic epithelial components is minimally affected by signaling of death factors (103, 105). Although the apoptosis signaling pathway mediated by mitochondria has not been investigated in odontogenic tumors, Bcl-2 and IAP family proteins, modulators of the mitochondrial apoptotic pathway, have been examined in various types of odontogenic tumors. In these odontogenic tumors, apoptosis inhibitory factors, such as Bcl-2, Bcl-x, survivin, and X chromosome-linked IAP (XIAP), are predominantly expressed, suggesting that these apoptosis modulators are associated with survival and neoplastic transformation of odontogenic epithelial cells (33, 68, 88, 92, 102, 104, 106–110).

Regulators of tooth development

Tooth development is under the strict genetic control of regulators that determine the positions and shapes of the teeth, such as Msx-1, Msx-2, Dlx-2, Barx-1, and Pax-9, or that are involved in the morphogenesis and cytodifferentiation of the teeth, such as Sonic hedgehog (SHH), bone morphogenetic protein (BMP), Wnt, HGF, and FGF (112, 113). Aberrant functions of these specific genes cause various dental anomalies (114, 115).

SHH signals control cell-cell interactions and cell proliferation in tissue patterning of various organs, including the teeth (116, 117). Patched (PTC), whose product is one of the SHH signal transduction molecules, is responsible for basal cell nevus syndrome (BCNS), characterized by basal cell carcinomas and odontogenic keratocysts (118, 119), and mutations of PTC have been identified in both BCNS-associated and sporadic odontogenic keratocysts (120, 121). On cDNA microarray, ameloblastomas show underexpression of SHH and PTC (25). Expression of SHH signaling molecules, SHH, PTC, smoothened (SMO), and GLI1, has been detected in several odontogenic tumors (122, 123). These findings suggest that SHH signaling pathway plays a role in epithelial-mesenchymal interactions and cell proliferation during the growth of odontogenic tumors as well as during tooth development.

Wnt signal transduction controls diverse developmental processes by regulating cell proliferation, morphology, motility, and fate in various organs, including the teeth (41, 124). Mice targeted for the Wnt signaling molecule *LEF-1* show inhibition of tooth morphogenesis (125). Wnt signaling is regulated by the levels of β -catenin, and activation of this pathway results in cytoplasmic accumulation and nuclear translocation of β -catenin (41). Mutations of β -catenin are detected frequently in calcifying odontogenic cysts, but are rare in ameloblastomas (126). Nuclear expression of β-catenin protein has been found in calcifying odontogenic cysts and benign and malignant ameloblastomas (43, 126, 127). APC, a product of tumor-suppressor gene, downregulates Wnt signaling pathway by inducing β -catenin degradation (41, 42), and immunohistochemical reactivity for APC is low in benign and malignant ameloblastomas (43). These findings suggest that aberrations of Wnt signaling pathway are involved in oncogenesis and cytodifferentiation of odontogenic epithelium via dysregulation of cell proliferation.

Other regulators of tooth development, such as HGF and FGF, have been found in odontogenic tumors (67, 68, 74, 75).

Hard tissue-related proteins

Enamel matrix is made from non-collagenous proteins and contains enamel proteins, such as amelogenin,

enamelin, ameloblastin (amelin or sheathlin), and tuftelin, and enzymes, such as matrix metalloproteinase (MMP)-20 (enamelysin) and enamel matrix serine proteinase 1 (128). Genetic alterations of enamel proteins have been implicated in various subtypes of amelogenesis imperfecta, a group of disorders that solely affect enamel formation (129, 130). Expression of amelogenin, enamelin, ameloblastin, and MMP-20 has been recognized in epithelial components of various types of odontogenic tumors (68, 92, 131-139). Recently, mutations of ameloblastin gene have been detected in ameloblastomas, adenomatoid odontogenic tumor and squamous odontogenic tumor (138, 140), and ameloblastin-mutant mice develop odontogenic tumors of odontogenic epithelium origin (141). These findings suggest that aberrations of enamel-related proteins are involved in oncogenesis of odontogenic epithelium.

Mineralized matrices of dentin, cementum, and bone contain type I collagen and numerous non-collagenous proteins, such as bone sialoprotein (BSP), osteonectin, osteocalcin, osteopontin, and dentin matrix protein 1 (DMP1; 142, 143). Two dentin proteins, dentin sialoprotein (DSP) and dentin phosphoprotein (DPP), are tooth-specific (144). Recent studies suggest that DSP, DPP, or DMP1 may cause dentinogenesis imperfecta type II, a hereditary developmental disturbance of dentin without any systemic disorder (129). BSP, osteonectin, osteocalcin, and osteopontin are found in many types of odontogenic tumors, suggesting that these proteins play a role in pathologic mineralization and/or tumor formation (134, 145-147). Furthermore, expression of BMP, which induces bone and cartilage formation, has been recognized in mixed and mesenchymal odontogenic tumors but not in epithelial odontogenic tumors (148).

Molecules involved in progression of odontogenic tumors

Cell adhesion molecules

Cells are neatly glued to each other and their surroundings by a variety of cell adhesion molecules, and aberrations in such interactions can lead to pathologic conditions (10, 149). Invasion and metastasis of various neoplastic lesions have been reported to correlate with altered adhesive systems involving cell adhesion molecules, such as E-cadherin, E-selectin, integrins, and CD44 (10, 150–152).

Vascular endothelium cell adhesion molecules, such as E-selectin, intercellular adhesion molecule (ICAM)-1, and vascular cell adhesion molecule (VCAM)-1, mediate leukocyte adhesion in inflammatory processes (153). Ameloblastomas express these cell adhesion molecules in vascular endothelium, suggesting that stromal blood vessels are activated in the odontogenic tumors (154). A major cell adhesion molecule for homophilic cell–cell adhesion of epithelial cells, E-cadherin, and its undercoat protein, *alpha*-catenin, are found in epithelial odontogenic tumors, and a case of malignant ameloblastoma has shown markedly decreased expression of these molecules (155). Integrins and CD44 mediate cell-cell and cell-extracellular matrix adhesion in various tissues (150, 151). Ameloblastomas have shown expression of several integrins and CD44 predominantly at epithelial-mesenchymal interfaces, suggesting that these cell adhesion molecules might mediate parenchymal-stromal interactions in the odontogenic tumors (156, 157).

Matrix-degrading proteinases

Extracellular matrix degradation occurring during developmental processes, tissue remodeling, inflammatory diseases, and tumor progression requires the action of proteolytic enzymes, such as MMPs and matrix serine proteinases (MSPs) (9, 10, 158). Extracellular matrix components, such as collagens, fibronectin, tenascin, laminin, and proteoglycans, have been detected in various odontogenic tumors (134, 159-164). Ameloblastomas and odontogenic myxomas express MMPs-1, -2, and -9, which degrade extracellular matrix and basement membrane components, and their inhibitors, tissue inhibitors of metalloproteinases (TIMPs)-1 and -2, have also been recognized in ameloblastomas (108, 165, 166). Recently, expression of heparanase, which cleaves proteoglycans, has been detected in ameloblastomas (167). These features suggest that matrix-degrading enzymes may contribute to the local invasiveness of odontogenic tumors.

Angiogenic factors

Angiogenesis is an essential part of a variety of physiologic and pathologic processes, including embryogenesis, wound healing, inflammation, and tumor progression, and these processes are controlled by numerous different molecules, such as VEGF, FGF, HGF, TGF- β , interleukin (IL)-8, and TNF- α (168–170). Immunohistochemical evaluation of microvessel density by means of the vascular endothelial marker CD34 has shown higher vascularity in benign and malignant ameloblastomas than in tooth germs. Simultaneously, increased expression of VEGF, which enhances angiogenesis and vascular permeability, has been found in these odontogenic tumors (171). These features suggest that VEGF is an important mediator of tumor angiogenesis in ameloblastic tumors, and upregulation of VEGF might be associated with tumorigenesis or malignant transformation of odontogenic epithelium.

Osteolytic cytokines

The integrity of bone metabolism is maintained by a delicate balance between bone formation by osteoblasts and bone resorption by osteoclasts, and its dynamics is regulated by a wide variety of hormones, growth factors, and cytokines (172, 173). Inflammatory cytokines with osteolytic activity, such as IL-1, IL-6, and TNF- α , are synthesized in ameloblastomas (105, 154, 174, 175).

Hypercalcemia is the most common metabolic complication of malignancy, and parathyroid hormonerelated protein (PTHrP) has been implicated in malignancy-associated hypercalcemia (173, 174, 176, 177). This peptide shows structural and functional homology with PTH and increases osteoclastic bone resorption and renal tubular calcium reabsorption (177). Several cases of malignant ameloblastoma with hypercalcemia have been attributed to PTHrP (178, 179), and expression of PTHrP has been detected in neoplastic cells of benign and malignant ameloblastomas unassociated with hypercalcemia (180, 181).

Receptor activator of nuclear factor- κB ligand (RANKL) binds to its receptor expressed on osteoclast precursors, RANK, and stimulates osteoclast differentiation and activation, while osteoprotegerin (OPG) functions as a decoy receptor for RANKL and inhibits osteoclastogenesis and osteoclast activation (172). The balance between RANKL and OPG levels regulates osteoclasts and bone metabolism, and abnormalities of the RANKL–OPG system have been implicated in the pathogenesis of various bone lesions (172, 173). Benign and malignant ameloblastomas express RANKL and OPG predominantly in stromal cells rather than tumor cells (181, 182). These observations suggest that osteolytic cytokines have a role in local bone resorption during the progression of odontogenic tumors.

Concluding remarks

The development and progression of odontogenic tumors are affected by alterations of many kinds of genes and molecules. In particular, the characteristics of odontogenic tumors appear to depend on the molecular mechanisms associated with (i) tooth development, (ii) bone metabolism, and (iii) the malignant potential of tumors. Further molecular studies, including genomicand proteomic-based profiling, are required to clarify the etiology and pathogenesis of odontogenic tumors. A better understanding of underlying molecular mechanisms will help to predict the course of odontogenic tumors and lead to the development of new therapeutic applications, such as molecular-targeted treatment and patient-tailored therapy, for odontogenic tumors.

References

- Kramer IRH, Pindborg JJ, Shear M. WHO histological typing of odontogenic tumours. Berlin, Germany: Springer-Verlag, 1992; 1–34.
- 2. Sciubba JJ, Fantasia JE, Kahn LB. *Tumors and cysts of the jaw*. Washington, DC, USA: Armed Forces Institute of Pathology, 2001; 71–160.
- Reichart PA, Philipsen HP. Odontogenic tumors and allied lesions. London, UK: Quintessence Publishing, 2004; 41– 332.
- 4. Eversole LR. Malignant epithelial odontogenic tumors. Semin Diagn Pathol 1999; 16: 317–24.
- Melrose RJ. Benign epithelial odontogenic tumors. Semin Diagn Pathol 1999; 16: 271–87.
- Philipsen HP, Reichart PA. Revision of the 1992edition of the WHO histological typing of odontogenic tumours. A suggestion. J Oral Pathol Med 2002; 31: 253–8.
- Vogelstein B, Fearon ER, Hamilton SR, et al. Genetic alterations during colorectal-tumor development. N Engl J Med 1988; 319: 525–32.
- 8. Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990; **61**: 759–67.

- 9. Liotta LA, Stetler-Stevenson WG. Tumor invasion and metastasis: an imbalance of positive and negative regulation. *Cancer Res* 1991; **51**: 5054s–9s.
- Stetler-Stevenson WG, Aznavoorian S, Liotta LA. Tumor cell interactions with the extracellular matrix during invasion and metastasis. *Annu Rev Cell Biol* 1993; 9: 541–73.
- Stass SA, Mixson J. Oncogenes and tumor suppressor genes: therapeutic implications. *Clin Cancer Res* 1997; 3: 2687–95.
- 12. Haber DA, Fearon ER. The promise of cancer genetics. *Lancet* 1998; **351**: SII1–8.
- Little CD, Nau MM, Carney DN, Gazdar AF, Minna JD. Amplification and expression of the c-myc oncogene in human lung cancer cell lines. *Nature* 1983; **306**: 194–6.
- Bargmann CI, Hung MC, Weinberg RA. The neu oncogene encodes an epidermal growth factor receptorrelated protein. *Nature* 1986; **319**: 226–30.
- Barbacid M. Ras genes. Annu Rev Biochem 1987; 56: 779– 827.
- Kumamoto H, Sasano H, Taniguchi T, Suzuki T, Moriya T, Ichinohasama R. Chromogenic in situ hybridization analysis of HER-2/neu status in breast carcinoma: application in screening of patients for trastuzumab (Herceptin[®]) therapy. *Pathol Int* 2001; **51**: 579–84.
- 17. Schlessinger J. Cell signaling by receptor tyrosine kinases. *Cell* 2000; **103**: 211–25.
- Sandros J, Heikinheimo K, Happonen RP, Stenman G. Expression of p21RAS in odontogenic tumors. *APMIS* 1991; **99**: 15–20.
- Kumamoto H, Takahashi N, Ooya K. K-Ras gene status and expression of Ras/mitogen-activated protein kinase (MAPK) signaling molecules in ameloblastomas. J Oral Pathol Med 2004; 33: 360–7.
- Kumamoto H, Ooya K, Sasano H. Immunohistochemical localization of c-myc oncogene protein correlated with malignancy of oral epithelium. *Jpn J Oral Biol* 1991; 33: 315–9.
- Kumamoto H, Kinouchi Y, Ooya K. Telomerase activity and telomerase reverse transciptase (TERT) expression in ameloblastomas. J Oral Pathol Med 2001; 30: 231–6.
- 22. Gibson CW, Lally E, Herold RC, Decker S, Brinster RL, Sandgren EP. Odontogenic tumors in mice carrying albumin-myc and albumin-ras transgenes. *Calcif Tissue Int* 1992; **51**: 162–7.
- Wright JT, Hansen L, Mahler J, Szczesniak C, Spalding JW. Odontogenic tumours in the v-Ha-ras (TG.AC) transgenic mouse. Arch Oral Biol 1995; 40: 631–8.
- Dodds AP, Cannon RE, Suggs CA, Wright JT. mRNA expression and phenotype of odontogenic tumours in the v-Ha-*ras* transgenic mouse. *Arch Oral Biol* 2003; 48: 843– 50.
- Heikinheimo K, Jee KJ, Niini T, et al. Gene expression profiling of ameloblastoma and human tooth germ by means of a cDNA microarray. *J Dent Res* 2002; 81: 525– 30.
- Knudson AG Jr. Mutation and cancer: statistical study of retinoblastoma. *Proc Natl Acad Sci U S A* 1971; 68: 820– 3.
- 27. Weinberg RA. Tumor suppressor genes. *Science* 1991; **254**: 1138–46.
- 28. Knudson AG. Antioncogenes and human cancer. Proc Natl Acad Sci U S A 1993; 90: 10914–21.
- 29. Lane DP. p53, guardian of the genome. *Nature* 1992; **358**: 15–6.
- 30. Greenblatt MS, Bennett WP, Hollstein M, Harris CC. Mutations in the p53 tumor suppressor gene: clues to

- Slootweg PJ. p53 protein and Ki-67 reactivity in epithelial odontogenic lesions. An immunohistochemical study. J Oral Pathol Med 1995; 24: 393–7.
- McDonald AR, Pogrel MA, Carson J, Regezi J. p53positive squamous cell carcinoma originating from an odontogenic cyst. J Oral Maxillofac Surg 1996; 54: 216–8.
- 33. Kumamoto H. Detection of apoptosis-related factors and apoptotic cells in ameloblastomas: analysis by immunohistochemistry and an *in situ* DNA nick end-labelling method. J Oral Pathol Med 1997; 26: 419–25.
- Sandra F, Nakamura N, Kanematsu T, Hirata M, Ohishi M. The role of MDM2 in the proliferative activity of ameloblastoma. *Oral Oncol* 2002; 38: 153–7.
- 35. Batista de Paula AM, da Costa Neto JQ, da Silva Gusmao E, Guimaraes Santos FB, Gomez RS. Immunolocalization of the p53 protein in a case of ameloblastic fibrosarcoma. *J Oral Maxillofac Surg* 2003; **61**: 256–8.
- 36. Kumamoto H, Izutsu T, Ohki K, Takahashi N, Ooya K. *p53* gene status and expression of p53, MDM2, and p14^{ARF} proteins in ameloblastomas. *J Oral Pathol Med* 2004; **33**: 292–9.
- 37. Shibata T, Nakata D, Chiba I, et al. Detection of TP53 mutation in ameloblastoma by the use of a yeast functional assay. *J Oral Pathol Med* 2002; **31**: 534–8.
- 38. Appel T, Gath R, Wernert N, Martini M, Berge S. Molekularbiologische und immunohistochemische Untersuchung des *tp53*-Gens in menschlichen Ameloblastomen. *Mund Kiefer Gesichtschir* 2004; 8: 167–72.
- 39. Carvalhais J, Aguiar M, Araujo V, Araujo N, Gomez R. p53 and MDM2 expression in odontogenic cysts and tumours. *Oral Dis* 1999; **5**: 218–22.
- Kumamoto H, Ohki K, Ooya K. Expression of p63 and p73 in ameloblastomas. *J Oral Pathol Med* 2005; 34: 220– 6.
- 41. Dale TC. Signal transduction by the Wnt family of ligands. *Biochem J* 1998; **329**: 209–23.
- 42. Fearnhead NS, Britton MP, Bodmer WF. The ABC of APC. *Hum Mol Genet* 2001; 10: 721–33.
- Kumamoto H, Ooya K. Immunohistochemical detection of β-catenin and adenomatous polyposis coli (APC) in ameloblastomas. *J Oral Pathol Med* 2005; 34: 401–6.
- 44. Pierce AM, Schneider-Broussard R, Gimenez-Conti IB, Russell JL, Conti CJ, Johnson DG. E2F1 has both oncogenic and tumor-suppressive properties in a transgenic model. *Mol Cell Biol* 1999; **19**: 6408–14.
- Kolodner RD. Mismatch repair: mechanisms and relationship to cancer susceptibility. *Trends Biochem Sci* 1995; 20: 397–401.
- 46. Karran P. Microsatellite instability and DNA mismatch repair in human cancer. *Semin Cancer Biol* 1996; 7: 15–24.
- 47. Ohki K, Kumamoto H, Ichinohasama R, et al. Genetic analysis of DNA microsatellite loci in salivary gland tumours: comparison with immunohistochemical detection of hMSH2 and p53 proteins. *Int J Oral Maxillofac Surg* 2001; **30**: 538–44.
- Castrilli G, Piantelli M, Artese L, et al. Expression of hMSH2 and hMLH1 proteins of the human DNA mismatch repair system in ameloblastoma. *J Oral Pathol Med* 2001; 30: 305–8.
- Kumar V, Abbas AK, Fausto N. Pathologic basis of disease. Philadelphia, USA: Elsevier Saunders, 2005; 324– 8.
- Lowy DR, Kirnbauer R, Schiller JT. Genital human papillomavirus infection. *Proc Natl Acad Sci U S A* 1994; 91: 2436–40.

- 51. Weinreb M, Day PJ, Niggli F, et al. The consistent association between Epstein-Barr virus and Hodgkin's disease in children in Kenya. *Blood* 1996; **87**: 3828–36.
- 52. Saiki Y, Ohtani H, Naito Y, Miyazawa M, Nagura H. Immunophenotypic characterization of Epstein-Barr virus-associated gastric carcinoma: massive infiltration by proliferating CD8+ T-lymphocytes. *Lab Invest* 1996; 75: 67–76.
- 53. van Heerden WF, van Rensburg EJ, Raubenheimer EJ, Venter EH. Detection of human papillomavirus DNA in an ameloblastoma using the in situ hybridization technique. *J Oral Pathol Med* 1993; **22**: 109–12.
- 54. Fujita S, Shibata Y, Takahashi H, Tsuda N, Okabe H. Latent infection with Epstein-Barr virus in odontogenic disorders: comparison among ameloblastoma, dentigerous cyst and odontogenic keratocyst. *Pathol Int* 1997; **47**: 449–53.
- Sand L, Jalouli J, Larsson PA, Magnusson B, Hirsch JM. Presence of human papilloma viruses in intraosseous ameloblastoma. *J Oral Maxillofac Surg* 2000; 58: 1129– 34.
- 56. Jang HS, Cho JO, Yoon CY, Kim HJ, Park JC. Demonstration of Epstein-Barr virus in odontogenic and nonodontogenic tumors by the polymerase chain reaction (PCR). *J Oral Pathol Med* 2001; **30**: 603–10.
- 57. Namin AK, Azad TM, Eslami B, Sarkarat F, Shahrokhi M, Kashanian F. A study of the relationship between ameloblastoma and human papilloma virus. *J Oral Maxillofac Surg* 2003; **61**: 467–70.
- 58. Migaldi M, Pecorari M, Rossi G, et al. Does HPV play a role in the etiopathogenesis of ameloblastoma? An immunohistochemical, in situ hybridization and polymerase chain reaction study of 18 cases using laser capture microdissection. *Mod Pathol* 2005; **18**: 283–9.
- 59. Heldin CH, Westermark B. Growth factors: mechanism of action and relation to oncogenes. *Cell* 1984; **37**: 9–20.
- Heldin CH, Miyazono K, ten Dijke P. TGF-β signalling from cell membrane to nucleus through SMAD proteins. *Nature* 1997; **390**: 465–71.
- 61. Ullrich A, Coussens L, Hayflick JS, et al. Human epidermal growth factor receptor cDNA sequence and aberrant expression of the amplified gene in A431 epidermoid carcinoma cells. *Nature* 1984; **309**: 418–25.
- 62. Heikinheimo K, Voutilainen R, Happonen RP, Miettinen PJ. EGF receptor and its ligands, EGF and TGF-alpha, in developing and neoplastic human odontogenic tissues. *Int J Dev Biol* 1993; **37**: 387–96.
- Li TJ, Browne RM, Matthews JB. Expression of epidermal growth factor receptors by odontogenic jaw cysts. *Virchows Arch A Pathol Anat Histopathol* 1993; 423: 137– 44.
- 64. Ueno S, Miyagawa T, Kaji R, Mushimoto K, Shirasu R. Immunohistochemical investigation of epidermal growth factor receptor expression in ameloblastomas. *J Pathol* 1994; **173**: 33–8.
- 65. Heikinheimo K, Happonen RP, Miettinen PJ, Ritvos O. Transforming growth factor β2 in epithelial differentiation of developing teeth and odontogenic tumors. *J Clin Invest* 1993; **91**: 1019–27.
- 66. Takata T, Miyauchi M, Ogawa I, et al. Immunoexpression of transforming growth factor β in desmoplastic ameloblastoma. *Virchows Arch* 2000; **436**: 319–23.
- 67. Kumamoto H, Yoshida M, Ooya K. Immunohistochemical detection of hepatocyte growth factor, transforming growth factor- β and their receptors in epithelial odontogenic tumors. *J Oral Pathol Med* 2002; **31**: 539–48.

- Yoshida M, Kumamoto H, Ooya K, Mayanagi H. Immunohistochemical analysis of mixed and mesenchymal odontogenic tumors. *Oral Med Pathol* 2003; 8: 125– 32.
- 69. van der Voort R, Taher TE, Derksen PW, Spaargaren M, van der Neut R, Pals ST. The hepatocyte growth factor/ Met pathway in development, tumorigenesis, and B-cell differentiation. *Adv Cancer Res* 2000; **79**: 39–90.
- 70. Tabata MJ, Kim K, Liu JG, et al. Hepatocyte growth factor is involved in the morphogenesis of tooth germ in murine molars. *Development* 1996; **122**: 1243–51.
- Ornitz DM. FGFs, heparan sulfate and FGFRs: complex interactions essential for development. *Bioessays* 2000; 22: 108–12.
- 72. Niswander L, Martin GR. Fgf-4 expression during gastrulation, myogenesis, limb and tooth development in the mouse. *Development* 1992; **114**: 755–68.
- 73. Vaahtokari A, Aberg T, Thesleff I. Apoptosis in the developing tooth: association with an embryonic signaling center and suppression by EGF and FGF-4. *Development* 1996; **122**: 121–9.
- 74. Myoken Y, Myoken Y, Okamoto T, Sato JD, Takada K. Immunohistochemical localization of fibroblast growth factor-1 (FGF-1) and FGF-2 in cultured human ameloblastoma epithelial cells and ameloblastoma tissues. *J Oral Pathol Med* 1995; 24: 387–92.
- 75. So F, Daley TD, Jackson L, Wysocki GP. Immunohistochemical localization of fibroblast growth factors FGF-1 and FGF-2, and receptors FGFR2 and FGFR3 in the epithelium of human odontogenic cysts and tumors. *J Oral Pathol Med* 2001; **30**: 428–33.
- 76. Seger R, Krebs EG. The MAPK signaling cascade. *FASEB J* 1995; **9**: 726–35.
- 77. Hoshino R, Chatani Y, Yamori T, et al. Constitutive activation of the 41-/43-kDa mitogen-activated protein kinase signaling pathway in human tumors. *Oncogene* 1999; **18**: 813–22.
- 78. Sandra F, Harada H, Nakamura N, Ohishi M. Midkine induced growth of ameloblastoma through MAPK and Akt pathways. *Oral Oncol* 2004; **40**: 274–80.
- Harley CB, Kim NW, Prowse KR, et al. Telomerase, cell immortality, and cancer. *Cold Spring Harb Symp Quant Biol* 1994; **59**: 307–15.
- 80. Shay JW, Bacchetti S. A survey of telomerase activity in human cancer. *Eur J Cancer* 1997; **33**: 787–91.
- Sumida T, Sogawa K, Hamakawa H, Sugita A, Tanioka H, Ueda N. Detection of telomerase activity in oral lesions. *J Oral Pathol Med* 1998; 27: 111–5.
- 82. Weinberg RA. The retinoblastoma protein and cell cycle control. *Cell* 1995; **81**: 323–30.
- Sherr CJ, Roberts JM. CDK inhibitors: positive and negative regulators of G1-phase progression. *Genes Dev* 1999; 13: 1501–12.
- Hall M, Peters G. Genetic alterations of cyclins, cyclindependent kinases, and Cdk inhibitors in human cancer. *Adv Cancer Res* 1996; 68: 67–108.
- 85. Kim J, Yook JI. Immunohistochemical study on proliferating cell nuclear antigen expression in ameloblastomas. *Eur J Cancer B Oral Oncol* 1994; **30B**: 126–31.
- Yamamoto K, Yoneda K, Yamamoto T, Ueta E, Osaki T. An immunohistochemical study of odontogenic mixed tumours. *Eur J Cancer B Oral Oncol* 1995; **31B**: 122–8.
- Sekine J, Kitamura A, Ueno K, et al. Cell kinetics in mandibular ameloblastic fibro-odontoma evaluated by bromodeoxyuridine and proliferating cell nuclear antigen immunohistochemistry: case report. *Br J Oral Maxillofac Surg* 1996; **34**: 450–3.

- Piattelli A, Fioroni M, Di Alberti L, Rubini C. Immunohistochemical analysis of a dentinogenic ghost cell tumour. *Oral Oncol* 1998; 34: 502–7.
- Sano K, Yoshida S, Ninomiya H, et al. Assessment of growth potential by MIB-1 immunohistochemistry in ameloblastic fibroma and related lesions of the jaws compared with ameloblastic fibrosarcoma. *J Oral Pathol Med* 1998; 27: 59–63.
- Takata T, Lu Y, Ogawa I, et al. Proliferative activity of calcifying odontogenic cysts as evaluated by proliferating cell nuclear antigen labeling index. *Pathol Int* 1998; 48: 877–81.
- Kumamoto H, Kimi K, Ooya K. Detection of cell cyclerelated factors in ameloblastomas. J Oral Pathol Med 2001; 30: 309–15.
- 92. Yoshida M, Kumamoto H, Ooya K, Mayanagi H. Histopathological and immunohistochemical analysis of calcifying odontogenic cysts. *J Oral Pathol Med* 2001; 30: 582–8.
- Kerr JF, Wyllie AH, Currie AR. Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer* 1972; 26: 239–57.
- Bellamy CO, Malcomson RD, Harrison DJ, Wyllie AH. Cell death in health and disease: the biology and regulation of apoptosis. *Semin Cancer Biol* 1995; 6: 3–16.
- 95. Cleveland JL, Ihle JN. Contenders in FasL/TNF death signaling. *Cell* 1995; **81**: 479–82.
- Adams JM, Cory S. The Bcl-2 protein family: arbiters of cell survival. *Science* 1998; 281: 1322–6.
- 97. Thornberry NA, Lazebnik Y. Caspases: enemies within. *Science* 1998; **281**: 1312–6.
- Deveraux QL, Reed JC. IAP family proteins-suppressors of apoptosis. *Genes Dev* 1999; 13: 239–52.
- Strasser A, O'Connor L, Dixit VM. Apoptosis signaling. Annu Rev Biochem 2000; 69: 217–45.
- 100. Wang X. The expanding role of mitochondria in apoptosis. *Genes Dev* 2001; **15**: 2922–33.
- Kumamoto H, Ooya K. Immunohistochemical and ultrastructural investigation of apoptotic cell death in granular cell ameloblastoma. *J Oral Pathol Med* 2001; 30: 245–50.
- 102. Kim J, Lee EH, Yook JI, Han JY, Yoon JH, Ellis GL. Odontogenic ghost cell carcinoma: a case report with reference to the relation between apoptosis and ghost cells. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2000; 90: 630–5.
- 103. Kumamoto H, Kimi K, Ooya K. Immunohistochemical analysis of apoptosis-related factors (Fas, Fas ligand, caspase-3 and single-stranded DNA) in ameloblastomas. *J Oral Pathol Med* 2001; **30**: 596–602.
- 104. Sandra F, Nakamura N, Mitsuyasu T, Shiratsuchi Y, Ohishi M. Two relatively distinct patterns of ameloblastoma: an anti-apoptotic proliferating site in the outer layer (periphery) and a pro-apoptotic differentiating site in the inner layer (centre). *Histopathology* 2001; **39**: 93–8.
- 105. Kumamoto H, Ooya K. Expression of tumor necrosis factor α , TNF-related apoptosis-inducing ligand, and their associated molecules in ameloblastomas. *J Oral Pathol Med* 2005; **34**: 287–94.
- 106. Mitsuyasu T, Harada H, Higuchi Y, et al. Immunohistochemical demonstration of bcl-2 protein in ameloblastoma. J Oral Pathol Med 1997; 26: 345–8.
- 107. Kumamoto H, Ooya K. Immunohistochemical analysis of bcl-2 family proteins in benign and malignant ameloblastomas. J Oral Pathol Med 1999; 28: 343–9.
- 108. Bast BT, Pogrel MA, Regezi JA. The expression of apoptotic proteins and matrix metalloproteinases in

73

odontogenic myxomas. J Oral Maxillofac Surg 2003; 61: 1463-6.

- 109. Fregnani ER, Pires FR, Quezada RD, Shih IM, Vargas PA, de Almeida OP. Calcifying odontogenic cyst: clinicopathological features and immunohistochemical profile of 10 cases. J Oral Pathol Med 2003; 32: 163–70.
- 110. Kumamoto H, Ooya K. Expression of survivin and X chromosome-linked inhibitor of apoptosis protein in ameloblastomas. *Virchows Arch* 2004; **444**: 164–70.
- 111. Levine AJ. p53, the cellular gatekeeper for growth and division. *Cell* 1997; **88**: 323–31.
- 112. Tucker AS, Sharpe PT. Molecular genetics of tooth morphogenesis and patterning: the right shape in the right place. *J Dent Res* 1999; **78**: 826–34.
- Jernvall J, Thesleff I. Reiterative signaling and patterning during mammalian tooth morphogenesis. *Mech Dev* 2000; 92: 19–29.
- 114. Thesleff I. Genetic basis of tooth development and dental defects. *Acta Odontol Scand* 2000; **58**: 191–4.
- 115. Miletich I, Sharpe PT. Normal and abnormal dental development. *Hum Mol Genet* 2003; **12**: R69–73.
- 116. Bitgood MJ, McMahon AP. Hedgehog and Bmp genes are coexpressed at many diverse sites of cell-cell interaction in the mouse embryo. *Dev Biol* 1995; **172**: 126–38.
- 117. Dassule HR, McMahon AP. Analysis of epithelialmesenchymal interactions in the initial morphogenesis of the mammalian tooth. *Dev Biol* 1998; **202**: 215–27.
- 118. Hahn H, Wicking C, Zaphiropoulous PG, et al. Mutations of the human homolog of *Drosophila* patched in the nevoid basal cell carcinoma syndrome. *Cell* 1996; **85**: 841– 51.
- 119. Johnson RL, Rothman AL, Xie J, et al. Human homolog of patched, a candidate gene for the basal cell nevus syndrome. *Science* 1996; **272**: 1668–71.
- Levanat S, Gorlin RJ, Fallet S, Johnson DR, Fantasia JE, Bale AE. A two-hit model for developmental defects in Gorlin syndrome. *Nat Genet* 1996; 12: 85–7.
- 121. Ohki K, Kumamoto H, Ichinohasama R, Sato T, Takahashi N, Ooya K. *PTC* gene mutations and expression of SHH, PTC, SMO, and GLI-1 in odontogenic keratocysts. *Int J Oral Maxillofac Surg* 2004; 33: 584–92.
- 122. Barreto DC, Bale AE, De Marco L, Gomez RS. Immunolocalization of PTCH protein in odontogenic cysts and tumors. J Dent Res 2002; 81: 757–60.
- 123. Kumamoto H, Ohki K, Ooya K. Expression of Sonic hedgehog (SHH) signaling molecules in ameloblastomas. *J Oral Pathol Med* 2004; 33: 185–90.
- 124. Sarkar L, Sharpe PT. Expression of Wnt signalling pathway genes during tooth development. *Mech Dev* 1999; 85: 197–200.
- 125. van Genderen C, Okamura RM, Farinas I, et al. Development of several organs that require inductive epithelialmesenchymal interactions is impaired in LEF-1-deficient mice. *Genes Dev* 1994; 8: 2691–703.
- 126. Sekine S, Sato S, Takata T, et al. β-catenin mutations are frequent in calcifying odontogenic cysts, but rare in ameloblastomas. *Am J Pathol* 2003; **163**: 1707–12.
- 127. Hassanein AM, Glanz SM, Kessler HP, Eskin TA, Liu C. β-Catenin is expressed aberrantly in tumors expressing shadow cells. Pilomatricoma, craniopharyngioma, and calcifying odontogenic cyst. Am J Clin Pathol 2003; 120: 732–6.
- 128. Nanci A. Oral histology. Philadelphia, USA: Mosby, 2003; 176–80.
- 129. Kurisu K, Tabata MJ. Human genes for dental anomalies. Oral Dis 1997; **3**: 223–8.

- Hu JC, Yamakoshi Y. Enamelin and autosomal-dominant amelogenesis imperfecta. *Crit Rev Oral Biol Med* 2003; 14: 387–98.
- 131. Mori M, Yamada K, Kasai T, Yamada T, Shimokawa H, Sasaki S. Immunohistochemical expression of amelogenins in odontogenic epithelial tumours and cysts. *Virchows Arch A Pathol Anat Histopathol* 1991; **418**: 319–25.
- Saku T, Okabe H, Shimokawa H. Immunohistochemical demonstration of enamel proteins in odontogenic tumors. *J Oral Pathol Med* 1992; 21: 113–9.
- 133. Snead ML, Luo W, Hsu DD, Melrose RJ, Lau EC, Stenman G. Human ameloblastoma tumors express the amelogenin gene. *Oral Surg Oral Med Oral Pathol* 1992; 74: 64–72.
- 134. Papagerakis P, Peuchmaur M, Hotton D, et al. Aberrant gene expression in epithelial cells of mixed odontogenic tumors. *J Dent Res* 1999; **78**: 20–30.
- 135. Takata T, Zhao M, Nikai H, Uchida T, Wang T. Ghost cells in calcifying odontogenic cyst express enamel-related proteins. *Histochem J* 2000; **32**: 223–9.
- 136. Takata T, Zhao M, Uchida T, Kudo Y, Sato S, Nikai H. Immunohistochemical demonstration of an enamel sheath protein, sheathlin, in odontogenic tumors. *Virchows Arch* 2000; **436**: 324–9.
- 137. Takata T, Zhao M, Uchida T, et al. Immunohistochemical detection and distribution of enamelysin (MMP-20) in human odontogenic tumors. *J Dent Res* 2000; **79**: 1608–13.
- 138. Toyosawa S, Fujiwara T, Ooshima T, et al. Cloning and characterization of the human ameloblastin gene. *Gene* 2000; **256**: 1–11.
- Kumamoto H, Yoshida M, Ooya K. Immunohistochemical detection of amelogenin and cytokeratin 19 in epithelial odontogenic tumors. *Oral Dis* 2001; 7: 171–6.
- 140. Perdigao PF, Gomez RS, Pimenta FJ, De Marco L. Ameloblastin gene (*AMBN*) mutations associated with epithelial odontogenic tumors. *Oral Oncol* 2004; **40**: 841– 6.
- 141. Fukumoto S, Kiba T, Hall B, et al. Ameloblastin is a cell adhesion molecule required for maintaining the differentiation state of ameloblasts. J Cell Biol 2004; 167: 973–83.
- 142. Butler WT, Ritchie H. The nature and functional significance of dentin extracellular matrix proteins. *Int J Dev Biol* 1995; **39**: 169–79.
- 143. Nanci A. Content and distribution of noncollagenous matrix proteins in bone and cementum: relationship to speed of formation and collagen packing density. *J Struct Biol* 1999; **126**: 256–69.
- 144. MacDougall M, Simmons D, Luan X, Nydegger J, Feng J, Gu TT. Dentin phosphoprotein and dentin sialoprotein are cleavage products expressed from a single transcript coded by a gene on human chromosome 4. Dentin phosphoprotein DNA sequence determination. J Biol Chem 1997; 272: 835–42.
- 145. Chen J, Aufdemorte TB, Jiang H, Liu AR, Zhang W, Thomas HF. Neoplastic odontogenic epithelial cells express bone sialoprotein. *Histochem J* 1998; **30**: 1–6.
- 146. Chen J, Sasaguri K, Sodek J, Aufdemorte TB, Jiang H, Thomas HF. Enamel epithelium expresses bone sialoprotein (BSP). *Eur J Oral Sci* 1998; **106**: 331–6.
- 147. Alvarez Perez MA, Pitaru S, Alvarez Fregoso O, Reyes Gasga J, Arzate H. Anti-cementoblastoma-derived protein antibody partially inhibits mineralization on a cementoblastic cell line. *J Struct Biol* 2003; **143**: 1–13.
- 148. Gao YH, Yang LJ, Yamaguchi A. Immunohistochemical demonstration of bone morphogenetic protein in odontogenic tumors. J Oral Pathol Med 1997; 26: 273–7.

- Gumbiner BM. Cell adhesion: the molecular basis of tissue architecture and morphogenesis. *Cell* 1996; 84: 345– 57.
- Albelda SM. Role of integrins and other cell adhesion molecules in tumor progression and metastasis. *Lab Invest* 1993; 68: 4–17.
- 151. Naor D, Sionov RV, Ish-Shalom D. CD44: structure, function, and association with the malignant process. *Adv Cancer Res* 1997; **71**: 241–319.
- Hirohashi S. Inactivation of the E-cadherin-mediated cell adhesion system in human cancers. *Am J Pathol* 1998; 153: 333–9.
- 153. Kluger MS. Vascular endothelial cell adhesion and signaling during leukocyte recruitment. *Adv Dermatol* 2004; **20**: 163–201.
- 154. Pripatnanont P, Song Y, Harris M, Meghji S. In situ hybridisation and immunocytochemical localisation of osteolytic cytokines and adhesion molecules in ameloblastomas. *J Oral Pathol Med* 1998; **27**: 496–500.
- 155. Kumamoto H, Ooya K. Expression of E-cadherin and alpha-catenin in epithelial odontogenic tumors: an immunohistochemical study. J Oral Pathol Med 1999; 28: 152–7.
- 156. Kumamoto H, Ohba S, Suzuki T, Ooya K. Immunohistochemical expression of integrins and CD44 in ameloblastomas. *Oral Med Pathol* 2001; **6**: 73–8.
- 157. Modolo F, Martins MT, Loducca SV, de Araujo VC. Expression of integrin subunits $\alpha 2$, $\alpha 3$, $\alpha 5$, αv , $\beta 1$, $\beta 3$ and $\beta 4$ in different histological types of ameloblastoma compared with dental germ, dental lamina and adult lining epithelium. *Oral Dis* 2004; **10**: 277–82.
- 158. Stetler-Stevenson WG, Liotta LA, Kleiner DE Jr. Role of matrix metalloproteinases in tumor invasion and metastasis. *FASEB J* 1993; 7: 1434–41.
- 159. Thesleff I, Ekblom P. Distribution of keratin and laminin in ameloblastoma. Comparison with developing tooth and epidermoid carcinoma. *J Oral Pathol* 1984; **13**: 85–96.
- 160. Sauk JJ. Basement membrane confinement of epithelial tumor islands in benign and malignant ameloblastomas. *J Oral Pathol* 1985; 14: 307–14.
- Nadimi H, Toto PD. Product identification of ameloblastomas: an immunohistochemical study. *J Oral Pathol* 1986; 15: 439–44.
- 162. Heikinheimo K, Morgan PR, Happonen RP, Stenman G, Virtanen I. Distribution of extracellular matrix proteins in odontogenic tumours and developing teeth. *Virchows Arch B Cell Pathol Incl Mol Pathol* 1991; **61**: 101–9.
- 163. Salo T, Kainulainen T, Parikka M, Heikinheimo K. Expression of laminin-5 in ameloblastomas and human fetal teeth. *J Oral Pathol Med* 1999; **28**: 337–42.
- 164. Ida-Yonemochi H, Ikarashi T, Nagata M, Hoshina H, Takagi R, Saku T. The basement membrane-type heparan sulfate proteoglycan (perlecan) in ameloblastomas: its intercellular localization in stellate reticulum-like foci and biosynthesis by tumor cells in culture. *Virchows Arch* 2002; 441: 165–73.
- 165. Kumamoto H, Yamauchi K, Yoshida M, Ooya K. Immunohistochemical detection of matrix metalloproteinases (MMPs) and tissue inhibitors of metalloproteinases (TIMPs) in ameloblastomas. J Oral Pathol Med 2003; 32: 114–20.
- 166. Pinheiro JJ, Freitas VM, Moretti AI, Jorge AG, Jaeger RG. Local invasiveness of ameloblastoma. Role played

by matrix metalloproteinases and proliferative activity. *Histopathology* 2004; **45**: 65–72.

- 167. Nagatsuka H, Han PP, Tsujigiwa H, et al. Heparanase gene and protein expression in ameloblastoma: possible role in local invasion of tumor cells. *Oral Oncol* 2005; **41**: 542–8.
- 168. Folkman J. Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat Med* 1995; 1: 27–31.
- 169. Toi M, Taniguchi T, Yamamoto Y, Kurisaki T, Suzuki H, Tominaga T. Clinical significance of the determination of angiogenic factors. *Eur J Cancer* 1996; **32A**: 2513–9.
- 170. Ferrara N, Davis-Smyth T. The biology of vascular endothelial growth factor. *Endocr Rev* 1997; 18: 4–25.
- 171. Kumamoto H, Ohki K, Ooya K. Association between vascular endothelial growth factor (VEGF) expression and tumor angiogenesis in ameloblastomas. *J Oral Pathol Med* 2002; **31**: 28–34.
- 172. Suda T, Takahashi N, Udagawa N, Jimi E, Gillespie MT, Martin TJ. Modulation of osteoclast differentiation and function by the new members of the tumor necrosis factor receptor and ligand families. *Endocr Rev* 1999; 20: 345–57.
- 173. Rodan GA, Martin TJ. Therapeutic approaches to bone diseases. *Science* 2000; **289**: 1508–14.
- 174. Macpherson DW, Hopper C, Meghji S. Hypercalcaemia and the synthesis of interleukin-1 by an ameloblastoma. *Br J Oral Maxillofac Surg* 1991; **29**–33.
- 175. Kubota Y, Nitta S, Oka S, Nakagawa S, Ninomiya T, Shirasuna K. Discrimination of ameloblastomas from odontogenic keratocysts by cytokine levels and gelatinase species of the intracystic fluids. *J Oral Pathol Med* 2001; 30: 421–7.
- 176. Kumamoto H, Ichinohasama R, Sawai T. Multiple organ failure associated with extensive metastatic calcification in a patient with an intermediate state of human T lymphotropic virus type I (HTLV-I) infection: report of an autopsy case. *Pathol Int* 1998; **48**: 313–8.
- 177. Suva LJ, Winslow GA, Wettenhall RE, et al. A parathyroid hormone-related protein implicated in malignant hypercalcemia: cloning and expression. *Science* 1987; **237**: 893–6.
- 178. Seward GR, Beales SJ, Jonson NW, Sita Lumsden EG. A metastasising ameloblastoma associated with renal calculi and hypercalcaemia. *Cancer* 1975; **36**: 2277–85.
- 179. Cox DP, Muller S, Carlson GW, Murray D. Ameloblastic carcinoma ex-ameloblastoma of the mandible with malignancy-associated hypercalcemia. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2000; **90**: 716–22.
- 180. Abdelsayed RA, Vartanian RK, Smith KK, Ibrahim NA. Parathyroid hormone-related protein (PTHrP) expression in ameloblastoma. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2004; 97: 208–19.
- 181. Kumamoto H, Ooya K. Expression of parathyroid hormone-related protein (PTHrP), osteoclast differentiation factor (ODF)/receptor activator of nuclear factorkappaB ligand (RANKL) and osteoclastogenesis inhibitory factor (OCIF)/osteoprotegerin (OPG) in ameloblastomas. J Oral Pathol Med 2004; 33: 46–52.
- 182. Tay JY, Bay BH, Yeo JF, Harris M, Meghji S, Dheen ST. Identification of RANKL in osteolytic lesions of the facial skeleton. J Dent Res 2004; 83: 349–53.

This document is a scanned copy of a printed document. No warranty is given about the accuracy of the copy. Users should refer to the original published version of the material.