Copyright © Blackwell Munksgaard Ltd

JOURNAL OF PERIODONTAL RESEARCH doi:10.1111/j.1600-0765.2005.00860.x

# T. E. Rams<sup>1</sup>, M. A. Listgarten<sup>2</sup>, J. Slots<sup>3</sup>

<sup>1</sup>Temple University School of Dentistry, Philadelphia, PA, USA, <sup>2</sup>University of California at San Francisco School of Dentistry, CA, USA, <sup>3</sup>University of Southern California School of Dentistry, Los Angeles, CA, USA

# Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis subgingival presence, species-specific serum immunoglobulin G antibody levels, and periodontitis disease recurrence

Rams TE, Listgarten MA, Slots J. Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis subgingival presence, species-specific serum immunoglobulin G antibody levels, and periodontitis disease recurrence. J Periodont Res 2006; 41: 228–234. © Blackwell Munksgaard 2006

*Background and Objective:* The biological and clinical effects of antibody against periodontal pathogenic bacteria are incompletely understood. This study evaluated the inter-relationships among periodontal levels of cultivable *Actinobacillus actinomycetemcomitans* and *Porphyromonas gingivalis*, species-specific serum immunoglobulin G (IgG) antibody levels, and periodontitis disease activity.

*Material and Methods:* Forty-three adults who had previously been treated for periodontitis and who also harbored cultivable *A. actinomycetemcomitans* or *P. gingivalis* were evaluated semiannually for clinical disease recurrence over a 36-month period. Each patient provided subgingival microbial samples, for the recovery of *A. actinomycetemcomitans* and *P. gingivalis*, from the two deepest pockets in each dentition sextant. *A. actinomycetemcomitans* and *P. gingivalis* serum IgG antibody levels were assessed using enzyme-linked immunosorbent assay (ELISA), together with whole-cell sonicate extracts from *A. actinomycetemcomitans* serotypes a–c and *P. gingivalis* ATCC 33277. Data were analyzed using the Mantel–Haenszel chi-square and Fisher exact two-tailed tests.

*Results:* Eighteen (60.0%) of 30 *A. actinomycetemcomitans*-positive subjects, and 10 (76.9%) of 13 *P. gingivalis*-positive subjects, exhibited recurrent periodontal breakdown within 36 months of periodontal therapy. Nineteen (67.9%) of the 28 patients with active periodontitis had *A. actinomycetemcomitans* or *P. gingivalis* serum antibody levels below designated threshold values. In comparison, 10 (66.7%) of 15 culture-positive clinically stable subjects showed *A. actinomycetemcomitans* or *P. gingivalis* serum antibody levels above threshold values. The difference between specific antibody levels in periodontitis-active and periodontitis-stable patients was statistically significant (p = 0.032).

Jørgen Slots, DDS, DMD, PhD, MS, MBA, University of Southern California School of Dentistry, MC-0641, Los Angeles, CA 90089– 0641, USA

email: jslots@usc.edu

Key words: Actinobacillus actinomycetemcomitans; antibody; periodontitis; Porphyromonas gingivalis

Accepted for publication November 1, 2005

*Conclusions:* Serum levels of IgG antibodies against *A. actinomycetemcomitans* or *P. gingivalis* in periodontitis-stable patients were higher than those in patients with active periodontitis. The results suggest that elevated levels of IgG antibody against *A. actinomycetemcomitans* and *P. gingivalis* have a detectable protective effect against periodontal infections with these microorganisms.

The periodontal pocket is densely populated with bacteria, making it an important site for host-microbe interactions. Periodontal bacteria stimulate B-cell proliferation, mostly through a classical antigen-specific immune response, but stimulation may also take place through B-cell superantigens or through the innate immune system. Several major periodontal pathogens, including Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis, are capable of inducing a robust serum antibody response (1). In fact, individuals infected with these two species of bacteria can exhibit serum antibody levels which exceed those found for many bacteria in medical infections. Particularly high antibody levels against A. actinomycetemcomitans are detected in juveniles with localized aggressive periodontitis (2) and against P. gingivalis in adults with advanced periodontitis (3). As A. actinomycetemcomitans and P. gingivalis only seem to inhabit the human oral cavity, the predominant antibody response against these two bacteria is derived from an oral source of infection, probably periodontal sites.

Preventing periodontitis through immunological approaches is appealing, but mechanisms of protective immunity in periodontal tissues are incompletely understood. A recent hypothesis suggests that periodontitis occurs in part because of a genetically determined nonefficacious immune response to periodontopathic bacteria (4,5). Despite decades of research, the relationship between specific serum immunogloblin G (IgG) antibody level and protection against periodontal pathogens, including A. actinomycetemcomitans and P. gingivalis, remains obscure. Some studies found high antibody levels against these two periodontopathic bacteria to be associated

with relative disease stability, whereas other studies detected no discernible antibody effect on disease activity (1). Discrepancy in research findings may be a consequence of using different methods measure to antibody responses and difficulties in assessing periodontitis disease activity. Delineating the role of host immune responsiveness in periodontal disease may provide new approaches for the control of periodontitis through the development of safe and effective vaccines.

To further elucidate the extent to which periodontal pathogens may stimulate systemic immune responses, and to understand how specific antibody responses may affect periodontal disease activity, the present study evaluated the serum IgG antibody response of periodontitis patients to whole-cell sonicates from A. actinomycetemcomitans and P. gingivalis. This study also aimed to relate periodontitis disease activity to levels of cultivable *A. actinomycetemcomitans* and P. gingivalis and to the levels of serum IgG antibodies against the two microorganisms.

### Material and methods

#### Subjects and clinical examinations

The patient population data were obtained from a prospective study of periodontitis disease recurrence. The experimental design, patient selection criteria, examination techniques, and clinical and microbiological findings have been described in detail previously (6,7). In brief, 98 adults, previously treated for moderate to advanced periodontitis, were enrolled in a systematic 3-month maintenance care program, and were clinically evaluated semiannually for disease recurrence over a 36-month period. Recurrent periodontitis disease activity was defined as any site, identified by two independent examiners, which exhibited either a probing depth increase of 3 mm or greater from baseline, or a probing depth increase of 2 mm or greater, together with a loss of relative periodontal attachment level of 2 mm or greater from baseline, as measured from the margin of an occlusal reference stent.

Baseline data on culture recovery of subgingival *A. actinomycetemcomitans* and *P. gingivalis* were compared with baseline serum levels of *A. actinomycetemcomitans* or *P. gingivalis* IgG antibodies in all 98 study subjects. In addition, baseline microbiological and immunological parameters were related to disease recurrence over 36 months in a subset of 43 *A. actinomycetemcomitans* or *P. gingivalis* culture-positive subjects.

#### **Microbiological procedures**

As previously described (6), A. actinomycetemcomitans and P. gingivalis were recovered from subgingival specimens obtained by paper points from the two pockets with the deepest probing depths in each of the study subject's six dentition sextants. Following a 2-h transport in VMGA III (8), and after dispersion using a Vortex mixer, 10-fold serial dilutions of plaque specimens were plated onto nonselective enriched Brucella blood agar (EBBA) for the recovery of P. gingivalis and the determination of total anaerobic viable counts (6), onto a selective medium for the isolation of black-pigmented gramnegative anaerobic species (9), and onto TSBV medium for the selective recovery of A. actinomycetemcomitans (10). EBBA plates and the selective medium for black-pigmented gramnegative anaerobic species were

incubated at 35°C in a Coy anaerobic chamber (Coy Laboratory Products, Inc., Ann Arbor, MI, USA), containing 85% N<sub>2</sub>, 10% H<sub>2</sub> and 5% CO<sub>2</sub>, for 10 days. TSBV plates were incubated at 35°C in 10% CO2/90% N2 for 4 days. Identification of A. actinomycetemcomitans and P. gingivalis isolates was carried out using the criteria and techniques of Slots (11,12). The proportional recovery of subgingival A. actinomycetemcomitans and P. gingivalis was determined from their cultivable counts in relation to total anaerobic viable counts on nonselective EBBA plates.

# Serum levels of IgG antibodies against *A. actinomycetemcomitans* and *P. gingivalis*

The immunological assays used were as previously described (7). In brief, an enzyme-linked immunosorbent assay (ELISA) was used to determine the levels of IgG antibodies to A. actinomycetemcomitans and P. gingivalis in serum collected by venepuncture from each of the 98 study subjects. Antigens for ELISA were prepared from wholecell extracts from A. actinomycetemcomitans serotypes a (ATCC 29523), b (Y4) and c (NCTC 9710), and from P. gingivalis ATCC 33277, as previously described (7). The coating and processing of ELISA plates, dilution of patient sera, incubation conditions, and determination of the optical density of ELISA plate wells, followed procedures previously described (7). A. actinomycetemcomitans and P. gingivalis antibody levels were expressed as the area in square millimeters under the dose-response dilution curve, using the formula described by Sedgwick et al. (13) and Pussinen et al. (14). The 90th percentile of all ELISA A. actinomycetemcomitans and P. gingivalis antibody measurements, determined by using an Excel 2000 spreadsheet (Microsoft Corp., Redmond, WA, USA), served as a threshold for the identification of study subjects demonstrating high vs. low serum IgG antibody levels to the two study organisms.

All microbiological and immunological laboratory procedures were performed blind, without knowledge of the clinical status of the study subjects or of the periodontal sites sampled.

#### Data analysis

Odds ratio analysis (15) was used with the Mantel-Haenszel chi-square test (16) or, for cell numbers less than 5, with the Fisher exact two-tailed test, to examine the relationship in subjects between the cultivable subgingival recovery of A. actinomycetemcomitans and P. gingivalis and the homologous serum IgG antibody level, and to assess the relationship between serum levels of IgG antibodies to A. actinomycetemcomitans or P. gingivalis and recurrent periodontitis disease activity. Positive predictive values (17) were calculated for the relationship between serum A. actinomycetemcomitans or P. gingivalis IgG antibody levels and clinical periodontal status over 36 months. The Student's t-test was used to determine significant differences in mean cultivable proportional recovery of subgingival A. actinomycetemcomitans and P. gingivalis between subjects with high- and low-serum A. actinomycetemcomitans and P. gingivalis IgG antibody levels. A *p*-value of  $\leq 0.05$ was required for statistical significance.

# Results

Culture analysis revealed the presence of subgingival *A. actinomycetemcomitans* in 31 (31.6%) of the 98 study subjects, and a mean of 2.1% (range: 0.0002–8.1%) *A. actinomycetemcomitans* isolates in culture-positive patients. Subgingival *P. gingivalis* was recovered from 13 (13.4%) study subjects, yielding an average of 3.3% (range: 0.1–16.5%) culturable *P. gingivalis*. No study subject revealed a dual infection of subgingival *A. actinomycetemcomitans* and *P. gingivalis*.

A total of 95 (96.9%) study subjects was found to have serum IgG antibody to one or more of the three A. actinomycetemcomitans serotypes tested, with the area under the ELISA curve ranging from 0 to 34.5 mm<sup>2</sup> per subject. Thirty (30.6%) subjects were found to have high serum A. actinomycetemcomitans IgG antibody levels to one or more of the three serotypes, using a threshold of 90th percentile and greater ( $\geq 15.1 \text{ mm}^2$ ) of all A. actinomycetemcomitans ELISA antibody values. High anti-A. actinomycetemcomitans antibody levels were most frequently detected against serotype c, either alone (14 subjects), or in joint high occurrence with serotype a (one subject). High IgG antibody levels against A. actinomycetemcomitans serotype b occurred in eight subjects and against serotype a in seven subjects.

Forty-two (42.9%) study subjects revealed serum IgG antibody reactivity to *P. gingivalis*, with ELISA antibody values ranging from 0 to 7.2 mm<sup>2</sup> per subject. Eleven (11.2%) subjects showed high serum *P. gingivalis* IgG antibody levels when the 90th percentile and greater ( $\geq$  3.3 mm<sup>2</sup>) of all *P. gingivalis* ELISA antibody levels was used as threshold.

Table 1 shows the relationship between subgingival presence and serum IgG antibody levels for *A. actinomycetemcomitans* and *P. gingivalis.* 

*Table 1.* Relationship between the subgingival presence of *Actinobacillus actinomycetemcomitans* and *Porphyromonas gingivalis* and homologous serum immunoglobulin G (IgG) antibody levels in 98 adults with treated periodontitis

	Culture-positive patients	Culture-negative patients
A. actinomycetemcomitans		
serum IgG antibody level		
$\geq 15.1 \text{ mm}^2$	16 <sup>a</sup>	14
$< 15.1 \text{ mm}^2$	15	53
P. gingivalis		
serum IgG antibody level		
$\geq 3.3 \text{ mm}^2$	$4^{\mathrm{a}}$	7
$< 3.3 \text{ mm}^2$	9	78

<sup>a</sup>Number of subjects; IgG, Immunoglobulin G (see Table 2).

Sixteen (51.6%) of the 31 A. actinomvcetemcomitans culture-positive subjects exhibited a high serum IgG antibody level to one or more of the three A. actinomycetemcomitans serotypes tested (ELISA curve threshold area of 15.1 mm<sup>2</sup>). Odds ratio analysis revealed a statistically significant positive subject relationship between the cultivable occurrence of subgingival A. actinomycetemcomitans and high homologous serum IgG level (odds ratio 4.0; p = 0.002, Mantel-Haenszel chi-square test). However, the mean recovery of A. actinomycetemcomitans was not significantly different in 16 subjects (standard error:  $6.0 \pm 1.8\%$ ) exhibiting high, and in 14 subjects  $(5.7 \pm 1.9\%)$  showing low, anti-A. actinomycetemcomitans serum IgG antibody levels. Four (30.8%) of 13 P. gingivalis culture-positive subjects showed a high IgG antibody level to the organism (ELISA curve threshold area of 3.3 mm<sup>2</sup>). A statistically significant positive subject association was found between the occurrence of subgingival P. gingivalis and a high homologous serum IgG antibody level (odds ratio 5.0; p = 0.037, Fisher exact two-tailed test). The mean recovery of *P. gingivalis* was not significantly different in three subjects  $(1.5 \pm 0.4\%)$  demonstrating high, and in 10 subjects  $(3.9 \pm 1.5\%)$  revealing a low, anti-*P. gingivalis* serum IgG antibody level.

Table 2 presents the relationship between A. actinomycetemcomitans or P. gingivalis serum IgG antibody levels and recurrent periodontitis disease activity in the 43 culture-positive subjects over the 36 study months. Eighteen (60.0%) of the 30 A. actinomycetemcomitans-positive subjects and 10 (76.9%) of the 13 P. gingivalis-positive subjects experienced recurrent periodontal breakdown in the posttreatment follow-up period. Perhaps, because of the relatively small number of study patients, when A. actinomycetemcomitans and P. gingivalis data sets were analyzed individually, no statistically significant relationship was found between culture-positive subjects with low serum antibody levels against either of the organisms and periodontitis disease activity (data not shown). However, when the A. actin-

*Table 2.* Periodontitis recurrence over 36 months in 43 subjects harboring either subgingival *Actinobacillus actinomycetemcomitans* or *Porphyromonas gingivalis*<sup>a</sup>

	Periodontitis disease-active patients	Clinically stable patients
A. actinomycetemcomitans		
culture-positive patients		
A. actinomycetemcomita	Ins	
serum IgG antibody lev	els	
$\geq 15.1 \text{ mm}^2$	8 <sup>b</sup>	8
$< 15.1 \text{ mm}^2$	10	4
P. gingivalis		
culture-positive patients		
P. gingivalis		
serum IgG antibody lev	els	
$\geq 3.3 \text{ mm}^2$	1 <sup>b</sup>	2
$< 3.3 \text{ mm}^2$	9	1
A. actinomycetemcomitans		
or P. gingivalis culture-pos	sitive	
patients		
High A. actinomyceteme	comitans	
or P. gingivalis serum Ig	gG antibody	
levels to homologous su	ıbgingival organism <sup>c</sup>	
+	9 <sup>b</sup>	10
_	19	5

<sup>a</sup>No subject yielded both *A. actinomycetemcomitans* and *P. gingivalis*.

<sup>b</sup>Number of subjects.

<sup>c</sup>Values were  $\ge 15.1 \text{ mm}^2$  for *A. actinomycetemcomitans* and  $\ge 3.3 \text{ mm}^2$  for *P. gingivalis.* IgG, immunoglobulin G.

omycetemcomitans and P. gingivalis data sets were combined, it was found that among the 28 culture-positive and disease-active subjects, 19 (67.9%) exhibited A. actinomycetemcomitans or P. gingivalis serum antibody levels below the designated threshold values. Patients who demonstrated cultivable A. actinomycetemcomitans or P. gingivalis, together with low homologous serum antibody levels, showed a positive predictive value of 79.2 and an odds ratio of 4.2 (p = 0.032, Mantel-Haenszel chi-square test) for periodontitis disease recurrence. Also, the difference between specific antibody levels in periodontitis-active and periodontitis-stable patients was statistically significant (p = 0.032,Mantel-Haenszel chi-square test).

Table 2 also shows that 10 (66.7%) of the 15 culture-positive subjects, who were clinically stable, revealed *A. ac-tinomycetemcomitans* or *P. gingivalis* serum IgG antibody levels above the designated threshold values. Patients who demonstrated cultivable *A. actinomycetemcomitans* or *P. gingivalis*, together with high homologous serum antibody levels, showed a positive predictive value of 52.6 and an odds ratio of 4.2 (p = 0.032, Mantel–Haenszel chi-square test) for exhibiting nonprogressing periodontal disease.

# Discussion

Virtually all previous studies on infectious periodontal disease have been limited to examining relationships between bacterial occurrence and periodontitis disease severity (18-20), presence between bacterial and homologous antibodies (21,22), or between specific antibody levels and periodontitis disease severity (2,23–28). Moreover, the design of most previous studies has been cross-sectional (21,29), and some of the few longitudinal studies have included relatively short follow-up periods (30,31). Studies that only examine a few aspects of the complex inter-relationships of bacteria-host resistance-periodontitis may be unable to elucidate critical pathogenic determinants of destructive periodontal disease. To overcome some of the weaknesses of previous

studies, the present investigation simultaneously determined the occurrence of subgingival *A. actinomycetemcomitans* and *P. gingivalis*, systemic IgG antibody levels against the two species in infected subjects, and periodontitis disease activity assessed over a period of 36 months.

Almost one-third of the study patients yielded A. actinomycetemcomitans. Although A. actinomycetemcomitans is commonly regarded as a pathogen of localized aggressive periodontitis (32), it is also frequently recovered from adult periodontitis lesions that are refractory to conventional mechanical therapy (33). The ability of A. actinomycetemcomitans to invade gingival tissue (34) may explain why the organism is largely unaffected by thorough scaling and root planing (35). Despite the importance of P. gingivalis in adult periodontitis (33), the organism was cultured from only a few study patients, a finding confirmed by immunofluorescence staining of direct subgingival smears (36). As periodontal P. gingivalis is confined to the pocket area, it has the potential to be markedly suppressed or eradicated by thorough mechanical periodontal therapy (35).

The major findings of the present study were the detection of relatively low A. actinomycetemcomitans and *P. gingivalis* antibody levels in patients with progressing periodontitis, and of relatively high antibody levels to both species in stable periodontitis patients. Those data are consistent with the notion that specific serum antibody responses towards A. actinomycetemcomitans and P. gingivalis confer protection for patients at risk of periodontitis. Studies have also shown that immunization with *P. gingivalis* can inhibit the progression of experimental periodontitis in nonhuman primates (37,38), rats (39,40) and mice (41), and can significantly reduce bacterial counts and tissue reactions in inoculated tissue cages of rabbits (42). Antibodies may neutralize toxins of A. actinomycetemcomitans (43) and proteolytic enzymes of P. gingivalis (44,45), as well as promote opsonization, agglutination/precipitation, steric hindrance of attachment, antibodydependent cell-mediated cytotoxicity, and complement activation (1). Thus, antibodies may not necessarily need to decrease periodontal pathogen loads in order to exert a protective effect, a notion that may help to explain the present finding of no relationship between specific serum antibody levels and the proportional recovery of the two test microorganisms in patients with stable periodontitis. Parenthetically, the observation that elevated antibody responses against A. actinomycetemcomitans and P. gingivalis seemed to help prevent further periodontal destruction provides additional evidence of the importance of the two organisms in the pathogenesis of periodontitis.

The finding of a positive relationship between subgingival A. actinomycetemcomitans counts and A. actinomycetemcomitans IgG antibody levels in serum has been described previously (46). A high pathogen load will generally induce a vigorous host immune response. It is less clear why eight A. actinomycetemcomitans-positive patients and one *P. gingivalis*-positive patient experienced continuing periodontal destruction, despite high specific antibody levels. In refractory human periodontitis (47), and in nonhuman primates immunized with P. gingivalis antigens (48,49), periodontal disease progression has previously been reported to occur in the presence of high anti-pathogen antibody levels. First, as discussed previously (50), some periodontitis cases may have involved herpesviruses, the control of which is dependent upon cellular immunity rather than upon antibodies. Second, the presence of subgingival microorganisms, other than A. actinomycetemcomitans and P. gingivalis, may have sustained further periodontal breakdown (20). Third, in spite of high specific antibody levels, the elicited antibody may vary widely in functional capacity (51) and a major portion may have been directed against bacterial antigens of little or no pathogenic relevance rather than towards critical antigens. A novel means of identifying critical virulence determinants in periodontopathic bacteria has recently been described (52). Fourth, the specific serum antibody may have been predominantly of the IgG2 isotype (53), which is mostly elicited by polysaccharide antigens, and which possesses relatively low avidity (54,55) and lacks strong complement fixation and opsonic properties (55,56). There may exist a degree of genetic control over the immunoglobulin phenotype response to P. gingivalis (57). Fifth, serum antibodies towards one periodontal pathogen may alter the functional antibody response against another periodontopathic microorganism (58,59). Sixth, some periodontal lesions may have harbored A. actinomycetemcomitans or P. gingivalis loads too heavy to be effectively controlled by the antibody threshold level used in the present study (60). Seventh, bacterial proteases in periodontal sites may have degraded antipathogen antibodies (61) or the C3 protein (62). Eighth, although less likely, some patients may have experienced periodontitis as a result of antibody-mediated immunopathological reactions (63).

Atypical were also the four culturepositive A. actinomycetemcomitans patients and the one culture-positive P. gingivalis patient, who exhibited low levels of homologous serum antibody but still did not experience periodontal breakdown during the 36-month study period. As A. actinomycetemcomitans and P. gingivalis may require a supporting active herpesvirus infection to produce periodontal destruction (50), the stable periodontitis lesions may have been devoid of periodontopathic herpesviruses. The possibility also exists that the infecting A. actinomycetemcomitans and P. gingivalis strains belonged to a less pathogenic serotype (64,65), or lacked one or more critical virulence factors (66,67). A. actinomycetemcomitans serotype c, which evoked a high antibody response in 15 of our study patients, may exert less periodontopathogenicity than serotype b (68,69), perhaps because serotype c strains tend to elicit high-avidity IgG antibodies (70). Finally, the available antibody may have effectively neutralized key virulence determinants and therefore sufficed in protecting the periodontal tissues.

In conclusion, the present study provides evidence that IgG antibodies against *A. actinomycetemcomitans* and *P. gingivalis* exert a protective role in periodontitis. After delineating critical antigens of periodontal pathogenic bacteria, studies are warranted to evaluate the feasibility of developing a vaccine against periodontitis.

# Acknowledgements

The study was supported by grants RO1-DE06085 and RR-01224 from the National Institutes of Health, Bethesda, MD, USA.

#### References

- Kinane DF, Mooney J, Ebersole JL. Humoral immune response to Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in periodontal disease. Periodontol 2000 1999;20:289–340.
- Taubman MA, Haffajee AD, Socransky SS, Smith DJ, Ebersole JL. Longitudinal monitoring of humoral antibody in subjects with destructive periodontal diseases. *J Periodont Res* 1992;27:511–521.
- Mouton C, Hammond PG, Slots J, Genco RJ. Serum antibodies to oral *Bacteroides* asaccharolyticus (Bacteroides gingivalis): relationship to age and periondontal disease. *Infect Immun* 1981;31:182–192.
- Schenkein HA. Finding genetic risk factors for periodontal diseases: is the climb worth the view? *Periodontol 2000* 2002;**30**:79–90.
- Diehl SR, Wu T, Burmeister JA, et al. Evidence of a substantial genetic basis for IgG2 levels in families with aggressive periodontitis. J Dent Res 2003;82:708–712.
- Listgarten MA, Slots J, Rosenberg J, Nitkin L, Sullivan P, Oler J. Clinical and microbiological characteristics of treated periodontitis patients on maintenance care. J Periodontol 1989;60:452–459.
- Listgarten MA, Slots J, Nowotny AH, et al. Incidence of periodontitis recurrence in treated patients with and without cultivable Actinobacillus actinomycetemcomitans, Prevotella intermedia, and Porphyromonas gingivalis: a prospective study. J Periodontol 1991;62:377–386.
- Möller ÅJR. Microbiological examination of root canals and periapical tissues of human teeth. Odont Tidskr 1966;74:1–380.
- Zambon JJ, Reynolds HS, Slots J. Blackpigmented *Bacteroides* spp. in the human oral cavity. *Infect Immun* 1981;32:198–203.
- Slots J. Selective medium for isolation of Actinobacillus actinomycetemcomitans. J Clin Microbiol 1982;15:606–609.

- Slots J. Rapid identification of important periodontal microorganisms by cultivation. Oral Microbiol Immunol 1986;1:49– 55.
- Slots J. Detection of colonies of *Bactero-ides gingivalis* by a rapid fluorescence assay for trypsin-like activity. *Oral Microbiol Immunol* 1987;2:139–141.
- Sedgwick AK, Ballow M, Sparks K, Tilton RC. Rapid quantitative microenzymelinked immunosorbent assay for tetanus antibodies. *J Clin Microbiol* 1983;18:104– 109.
- Pussinen PJ, Vilkuna-Rautianen T, Alfthan G, Mattila K, Asikainen S. Multiserotype enzyme-linked immunosorbent assay as a diagnostic aid for periodontitis in large-scale studies. J Clin Microbiol 2002;40:512–518.
- Fleiss JL. Statistical Methods for Rates and Proportions, 2nd edn. New York: John Wiley, 1981: 160–176.
- Mantel N, Haenszel W. Statistical aspects of the analysis of data from retrospective studies of disease. J Natl Cancer Inst 1959;22:719–748.
- McNeil BJ, Keeler E, Adelstein SJ. Primer on certain elements of medical decision making. N Engl J Med 1975;293:211–215.
- Bragd L, Dahlén G, Wikström M, Slots J. The capability of Actinobacillus actinomycetemcomitans, Bacteroides gingivalis and Bacteroides intermedius to indicate progressive periodontitis; a retrospective study. J Clin Periodontol 1987;14:95–99.
- Kamma JJ, Nakou M, Gmür R, Baehni PC. Microbiological profile of early onset/ aggressive periodontitis patients. Oral Microbiol Immunol 2004;19:314–321.
- Socransky SS, Haffajee AD. Periodontal microbial ecology. *Periodontol 2000* 2005;38:135–187.
- Ebersole JL, Taubman MA, Smith DJ, Frey DE, Haffajee AD, Socransky SS. Human serum antibody responses to oral microorganisms. IV. Correlation with homologous infection. *Oral Microbiol Immunol* 1987;2:53–59.
- Ebersole JL, Cappelli D, Sandoval MN. Subgingival distribution of *A. actin*omycetemcomitans in periodontitis. *J Clin Periodontol* 1994;21:65–75.
- Celenligil H, Ebersole JL. Analysis of serum antibody responses to periodontopathogens in early-onset periodontitis patients from different geographical locations. *J Clin Periodontol* 1998;25:994– 1002.
- Celenligil-Nazliel H, Kansu E, Ebersole JL. Periodontal findings and systemic antibody responses to oral microorganisms in Behcet's disease. *J Periodontol* 1999;70:1449–1456.
- Furuichi Y, Shimotsu A, Ito H, et al. Associations of periodontal status with general health conditions and serum anti-

body titers for *Porphyromonas gingivalis* and *Actinobacillus actinomycetemcomitans. J Periodontol* 2003;**74**:1491–1497.

- Nguyen KA, DeCarlo AA, Paramaesvaran M, Collyer CA, Langley DB, Hunter N. Humoral responses to *Porphyromonas* gingivalis gingipain adhesin domains in subjects with chronic periodontitis. *Infect Immun* 2004;72:1374–1382.
- Yang HW, Asikainen S, Dogan B, Suda R, Lai CH. Relationship of *Actinobacillus actinomycetemcomitans* serotype b to aggressive periodontitis: frequency in pure cultured isolates. *J Periodontol* 2004; 75:592–599.
- Nakashima K, Kobayashi T, Yoshihara A, Fujiwara J, Miyazaki H, Kowashi Y. Periodontal conditions in an elderly Japanese population influenced by smoking status and serum immunoglobulin G2 levels. J Periodontol 2005;76:582–589.
- Craig RG, Boylan R, Yip J, et al. Serum IgG antibody response to periodontal pathogens in minority populations: relationship to periodontal disease status and progression. J Periodont Res 2002;37:132– 146.
- O'Dell DS, Ebersole JL. Longitudinal changes in antibody avidity to Actinobacillus actinomycetemcomitans in periodontitis. J Clin Periodontol 1996;23: 203–211.
- Apatzidou DA, Riggio MP, Kinane DF. Impact of smoking on the clinical, microbiological and immunological parameters of adult patients with periodontitis. *J Clin Periodontol* 2005;**32**:973–983.
- 32. Slots J, Schonfeld SE. Actinobacillus actinomycetemcomitans in localized juvenile periodontitis. In: Hamada S, Holt SC, McGhee JR, eds. Periodontal Disease: Pathogens and Host Immune Responses. Tokyo, Japan: Quintessence Publishing Co., Ltd., 1991: 53–64.
- Slots J, Ting M. Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in human periodontal disease: occurrence and treatment. Periodontol 2000 1999;20:82–121.
- Christersson LA, Albini B, Zambon JJ, Wikesjo UM, Genco RJ. Tissue localization of *Actinobacillus actinomycetemcomitans* in human periodontitis. I. Light, immunofluorescence and electron microscopic studies. *J Periodontol* 1987;58:529– 539.
- Renvert S, Wikström M, Dahlén G, Slots J, Egelberg J. Effect of root debridement on the elimination of *Actinobacillus actinomycetemcomitans* and *Bacteroides gingivalis* from periodontal pockets. J Clin Periodontol 1990;17:345–350.
- Lai C-H, Azimzadeh R, Slots J, Listgarten MA. Occurrence of *B. gingivalis* (Bg), *B. intermedius* (Bi) and *A. actinomycetem-comitans* (Aa) in human periodontal

disease. J Dent Res 1990; 69 (Spec. Issue): 242 (Abstr 1071).

- Persson GR, Engel D, Whitney C, et al. Immunization against Porphyromonas gingivalis inhibits progression of experimental periodontitis in nonhuman primates. Infect Immun 1994;62:1026–1031.
- Moritz AJ, Cappelli D, Lantz MS, Holt SC, Ebersole JL. Immunization with *Porphyromonas gingivalis* cysteine protease: effects on experimental gingivitis and ligature-induced periodontitis in *Macaca fascicularis*. J Periodontol 1998;69:686– 697.
- Evans RT, Klausen B, Sojar HT, et al. Immunization with Porphyromonas (Bacteroides) gingivalis fimbriae protects against periodontal destruction. Infect Immun 1992;60:2926–2935.
- Rajapakse PS, O'Brien-Simpson NM, Slakeski N, Hoffmann B, Reynolds EC. Immunization with the RgpA-Kgp proteinase-adhesin complexes of *Porphyromonas gingivalis* protects against periodontal bone loss in the rat periodontitis model. *Infect Immun* 2002;**70:**2480– 2486.
- Gibson FC III, Genco CA. Prevention of *Porphyromonas gingivalis*-induced oral bone loss following immunization with gingipain R1. *Infect Immun* 2001;69:7959– 7963.
- Dahlén G, Slots J. Experimental infections by *Bacteroides gingivalis* in non-immunized and immunized rabbits. *Oral Microbiol Immunol* 1989;4:6–11.
- Engstrom PE, George M, Larsson P, Lally ET, Taichman NS, Norhagen G. Oral and systemic immunoglobulin G-subclass antibodies to Actinobacillus actinomycetemcomitans leukotoxin. Oral Microbiol Immunol 1999;14:104–108.
- 44. Nakagawa T, Sims T, Fan Q, et al. Functional characteristics of antibodies induced by Arg-gingipain (HRgpA) and Lys-gingipain (Kgp) from Porphyromonas gingivalis. Oral Microbiol Immunol 2001;16:202–211.
- 45. Yonezawa H, Ishihara K, Okuda K. Arg-gingipain a DNA vaccine induces protective immunity against infection by *Porphyromonas gingivalis* in a murine model. *Infect Immun* 2001;69:2858–2864.
- Ebersole JL, Capelli D, Steffen MJ. Longitudinal dynamics of infection and serum antibody in *A. actinomycetemcomitans* periodontitis. *Oral Dis* 1995;1:129–138.
- Haffajee AD, Socransky SS, Dzink JL, Taubman MA, Ebersole JL. Clinical, microbiological and immunological features of subjects with refractory perio-

dontal diseases. *J Clin Periodontol* 1988;**15**:390–398.

- Ebersole JL, Brunsvold M, Steffensen B, Wood R, Holt SC. Effects of immunization with *Porphyromonas gingivalis* and *Prevotella intermedia* on progression of ligature-induced periodontitis in the nonhuman primate *Macaca fascicularis*. *Infect Immun* 1991;59:3351–3359.
- Holt SC, Brunsvold M, Jones A, Wood R, Ebersole JL. Cell envelope and cell wall immunization of *Macaca fascicularis*: effect on the progression of ligature-induced periodontitis. *Oral Microbiol Immunol* 1995;10:321–333.
- Slots J. Herpesviruses in periodontal diseases. *Periodontol 2000* 2005;38:33–62.
- Anderson DM, Ebersole JL, Novak MJ. Functional properties of nonhuman primate antibody to *Porphyromonas gingivalis. Infect Immun* 1995;63:3245–3252.
- Handfield M, Progulske-Fox A, Hillman JD. In vivo induced genes in human diseases. *Periodontol 2000* 2005;38:123–134.
- Nakashima K, Schenkein HA, Califano JV, Tew JG. Heterogeneity of antibodies reactive with the dominant antigen of *Actinobacillus actinomycetemcomitans. Infect Immun* 1997;65:3794–3798.
- Lopatin DE, Blackburn E. Avidity and titer of immunoglobulin G subclasses to *Porphyromonas gingivalis* in adult periodontitis patients. *Oral Microbiol Immunol* 1992;7:332–337.
- 55. Whitney C, Ant J, Moncla B, Johnson B, Page RC, Engel D. Serum immunoglobulin G antibody to *Porphyromonas gingi*valis in rapidly progressive periodontitis: titer, avidity, and subclass distribution. *Infect Immun* 1992;**60**:2194–2200.
- Ling TY, Sims TJ, Chen HA, et al. Titer and subclass distribution of serum IgG antibody reactive with Actinobacillus actinomycetemcomitans in localized juvenile periodontitis. J Clin Immunol 1993;13:101–112.
- Gemmell E, Winning TA, Carter CL, et al. Differences in mouse strain influence leukocyte and immunoglobulin phenotype response to Porphyromonas gingivalis. Oral Microbiol Immunol 2003;18:364–370.
- Choi JI, Kim US, Kim SJ, Son WS, Park HR. Fusobacterium nucleatum impairs serum binding to Porphyromonas gingivalis biofilm. Oral Microbiol Immunol 2003;18:92–94.
- Gemmell E, Bird PS, Ford PJ, et al. Modulation of the antibody response by Porphyromonas gingivalis and Fusobacterium nucleatum in a mouse model. Oral Microbiol Immunol 2004;19:247–251.

- Ebersole JL, Feuille F, Kesavalu L, Holt SC. Host modulation of tissue destruction caused by periodontopathogens: effects on a mixed microbial infection composed of *Porphyromonas gingivalis* and *Fusobacterium nucleatum*. *Microb Pathog* 1997;23:23–32.
- Grenier D, Mayrand D, McBride BC. Further studies on the degradation of immunoglobulins by black-pigmented *Bacteroides. Oral Microbiol Immunol* 1989;4:12–18.
- Sundqvist GK, Carlsson J, Herrmann BF, Höfling JF, Vaatainen A. Degradation in vivo of the C3 protein of guinea-pig complement by a pathogenic strain of *Bacteroides gingivalis. Scand J Dent Res* 1984;92:14–24.
- Brandtzaeg P. Inflammatory bowel disease: clinics and pathology. Do inflammatory bowel disease and periodontal disease have similar immunopathogeneses? Acta Odontol Scand 2001;59: 235–243.
- Ebersole JL, Steffen MJ. Human antibody responses to outer envelope antigens of *Porphyromonas gingivalis* serotypes. *J Periodontal Res* 1995;**30**:1–14.
- 65. Yang HW, Huang YF, Chan Y, Chou MY. Relationship of *Actinobacillus actinomycetemcomitans* serotypes to periodontal condition: prevalence and proportions in subgingival plaque. *Eur J Oral Sci* 2005;113:28–33.
- Fives-Taylor PM, Meyer DH, Mintz KP, Brissette C. Virulence factors of Actinobacillus actinomycetemcomitans. Periodontol 2000 1999;20:136–167.
- Holt SC, Ebersole JL. Porphyromonas gingivalis, Treponema denticola, and Tannerella forsythia: the 'red complex', a prototype polybacterial pathogenic consortium in periodontitis. Periodontol 2000 2005;38:72–122.
- Zambon JJ, Slots J, Genco RJ. Serology of oral Actinobacillus actinomycetemcomitans and serotype distribution in human periodontal disease. Infect Immun 1983;41:19–27.
- Asikainen S, Lai CH, Alaluusua S, Slots J. Distribution of *Actinobacillus actinomyce*temcomitans serotypes in periodontal health and disease. Oral Microbiol Immunol 1991;6:115–118.
- Wang D, Kawashima Y, Nagasawa T, et al. Elevated serum IgG titer and avidity to Actinobacillus actinomycetemcomitans serotype c in Japanese periodontitis patients. Oral Microbiol Immunol 2005;20:172–179.

This document is a scanned copy of a printed document. No warranty is given about the accuracy of the copy. Users should refer to the original published version of the material.