PERIODONTAL RESEARCH

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Single nucleotide polymorphisms of complement component 5 and periodontitis

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Background and Objective: Polymorphisms of host defence genes might increase one's risks for periodontitis. This study investigated whether tagging single nucleotide polymorphisms (SNPs) of the gene encoding complement component 5 (C5) are associated with periodontitis in a Hong Kong Chinese population.

Material and Methods: Eleven tagging SNPs of 229 patients with at least moderate periodontitis and 207 control subjects without periodontitis were genotyped using an i-plexGOLD MassARRAY mass-spectrometry system.

Results: Genotype AG of SNP rs17611 was more prevalent in the group of periodontitis patients than in the controls (54.6% vs. 41.7%, p = 0.007). The haplotype CGCA of the haplotype block consisting of rs1035029, rs17611, rs25681 and rs992670 was significantly associated with periodontitis in a dominant model (p = 0.001). The SNP rs17611 showed high linkage disequilibrium with rs1035029, rs25681 and rs992670. Smoking was also significantly associated with periodontitis (p = 0.006).

Conclusion: The tagging SNP rs17611 of the C5 gene and smoking may be associated with periodontitis among the Hong Kong Chinese population.

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Periodontitis is a complex human inflammatory disease that is caused by dental plaque, and its clinical manifestations are determined by both environmental influences on, and the genetic makeup of, affected individuals. Polymorphisms of host-defence genes might also increase the risk of periodontitis (1).

The complement system is a lytic effector system that protects the host against microbial pathogens, acts as a key link between innate and specific immune responses (2,3), and has been implicated in the pathogenesis of periodontitis. The complement system components have been detected in the gingival cervical fluid collected from

patients with periodontitis (4,5). It has been reported that increased cleavage of complement is associated with increased severity of inflammation and periodontal destruction (6). Complement component deficiencies seem to be associated with severe chronic periodontitis (7). Complement component 5 (C5) is a pivotal element in the complement system and therefore could take part in the pathogenesis of periodontitis. C5 is cleaved by C5 convertase to yield the C5b fragment and the anaphylatoxin C5a. The latter binds to the G proteincoupled receptor C5aR to trigger intracellular signalling, which results in chemotaxis, respiratory burst and release of proinflammatory mediators from granulocytes (8,9). The C5b fragment combines with complement components C6 and C7 to initiate the formation of the membrane attack complex (MAC) in the membrane of invading microorganisms (10). Because haplotype and tagging polymorphisms of C5 may be associated with some chronic inflammatory diseases such as liver fibrosis, rheumatoid arthritis and bronchial asthma (11-16), C5 polymorphisms may also be important in chronic inflammatory oral diseases such as periodontitis. Published data on the relevance of C5 polymorphisms in periodontitis are not yet available.

The objective of this study was to screen tagging single nucleotide

polymorphisms (SNPs) of the C5 gene in Hong Kong Chinese patients with moderate to severe periodontitis and in periodontitis-free controls. We investigated whether there were any genetic variations in the C5 genes of these two groups and tested whether such variations, together with other possible risk factors, such as smoking status, age and gender, were associated with periodontitis.

Material and methods

This case–control study was approved by the Ethics Committee of the Faculty of Dentistry, The University of Hong Kong. Written informed consent was obtained from all participants.

Study participants

Participants were recruited from new patients attending the Primary Care Clinic, Prince Philip Dental Hospital, Faculty of Dentistry, The University of Hong Kong, from May 2005 to August 2007. Patients' records and radiographs were screened within 1 mo of the first attendance at the clinic, and potentially eligible patients were invited to attend a clinical examination. Demographic information, and medical and dental histories, were obtained from patients' records and were supplemented with information obtained during the day of the clinical examination. Race and ethnicity were self-reported, with a participant being considered Chinese if his or her biological parents, grandparents and great-grandparents were all reported to be ethnic Chinese. Smoking history was self-reported; patients who currently smoked or who had quit smoking in the 12 mo before the start of the study were considered to be smokers, and those who had never smoked or who had quit smoking more than 12 mo before the start of the study were considered to be nonsmokers. All participants needed to be systemically healthy, so those with systemic conditions, including cardiovascular diseases, hypertension, liver diseases, kidney diseases, blood disorders, diabetes mellitus, autoimmune diseases and malignant tumors were excluded, as were pregnant women.

Periodontitis-free and periodontitis groups were defined on the basis of both radiographs and the findings of clinical examinations. Cases or periodontitis subjects were Chinese patients, 18-60 years of age, with radiographic evidence of at least moderate periodontitis, according to the following criteria: the orthopantomogram (OPG) taken at the first visit to the Prince Philip Dental Hospital before recruitment showed more than 50% alveolar bone loss at more than 30% of sites, as measured with a Schei ruler without using the 1 mm space (each tooth contributed to a mesial and a distal site) (17); and subsequent clinical periodontal examination showed at least two teeth in each quadrant that had a probing depth of $\geq 5 \text{ mm}$ and that bled on probing. All the periodontitis subjects showed clinical and radiographic signs of attachment loss and bone loss. respectively, and were further classified into aggressive periodontitis or chronic periodontitis, described as follows. According to the 1999 periodontal disease classification criteria (18), patients \leq 35 years of age and systemic healthy but experiencing > 30% of sites with > 5 mm of clinical attachment loss are classified as having aggressive periodontitis, while the remaining subjects, > 35 years of age, who fulfilled our case recruitment criteria and self-reported nil periodontal disease history before reaching 35 years of age were classified as having chronic periodontitis. Controls were Chinese patients, 18-60 years of age, without periodontitis, according to the following criteria: the OPG taken at recruitment showed no sites with > 15% bone loss or any radiographic evidence of furcation involvement (each tooth contributed to a mesial and a distal site); subsequent clinical periodontal examination confirmed that there were no sites with a probing depth of > 4 mm and gingival recession of > 2 mm; and there was no history of tooth loss caused by periodontal diseases. Control or periodontitis-free subjects included periodontally healthy individuals and those with gingivitis. Bleeding on probing was not recorded in the control group.

DNA isolation and genotyping

Ten millilitres of venous blood was obtained from each participant and stored in ethylene-diamine-tetra-acetic acid at -70°C until DNA extraction. QIAamp DNA blood mini-kits (Qiagen, Hilden, Germany) were used to extract genomic DNA. Tagging SNPs of the C5 gene were selected by using the SNP Tagging Wizard of SNPBROWser software version 3.5 (Applied Biosystems, Foster City, CA, USA). The reason for choosing tagging SNPs was to select a minimum informative subset of SNPs and to eliminate redundant information caused by strong linkage disequilibrium (LD) among the SNPs in the region. Therefore, the SNPs studied here can provide some level of information about the SNPs not selected in this study. Tagging SNPs were those left after (i) eliminating all the SNPs with reported minor allele frequencies under 10% and (ii) reported to be 100% linked with other SNPs in the same region. SNP sequences were checked in the RealSNP Assay Database (Sequenom, San Diego, CA, USA). Genotyping was performed using the i-plexGOLD genotyping assay of the MassARRAY mass-spectrometry system, following the protocol recommended by the manufacturer (Sequenom).

Quality control

As recommended in the genotyping protocol, all template DNA samples needed to have a ratio of spectrophotometer readings at 260 nm and 280 nm (A₂₆₀/A₂₈₀) of between 1.7 and 2.0. Samples were then aliquoted into 96-well plates. For each well of DNA to be tested, five duplicate wells and one water-containing well served as internal and negative controls, respectively. Before genotyping, DNA samples and randomly selected positive controls were electrophoresed in a 1% agarose gel to confirm the quality and quantity of genomic DNA. After genotyping using iPLEX, 25 random

samples of selected SNPs were subjected to concordance testing by direct sequencing to ensure that the genotyping data were reliable.

Statistical analyses

Statistical analysis was performed using SPSS 15.0 (SPSS, Chicago, IL, USA). Differences between the distributions of gender, and reported smoking habit among patients with periodontitis and periodontitis-free individuals were tested using the chisquare test. Differences between cases and controls in the number of standing teeth and age were assessed using the *t*-test.

Three screening steps were adopted for genetic data processing. First, the genotype distribution and allele-type of each SNP were calculated among aggressive periodontitis or chronic periodontitis and periodontitis-free subjects. Second, those SNPs with a call rate of > 80% and minor allele frequencies of > 10% were put into the second step: the Hardy-Weinberg Equilibrium test. Third, those SNPs in the periodontitis-free group with Hardy-Weinberg Equilibrium test *p*-values of ≥ 0.01 were subjected to final statistical analysis. Multiple comparison of genotype and allele-type of aggressive periodontitis, chronic periodontitis and control groups were initially screened using the chi-square test. Single nucleotide polymorphisms fulfilling the criteria for final statistical

Table 1. Demography and clinical profile of study participants

analysis were put under stepwise logistic regression analysis, together with other possible risk indicators of periodontitis - namely age, gender and reported smoking habit. The alpha level was set at 0.05, unless otherwise specified. Risk alleles, odds ratios (ORs) and 95% confidence intervals (CIs) for each SNP were also determined. Linkage disequilibrium (LD) analysis and haplotype association analysis were performed using HAPLO-VIEW 3.32 (http://www.broad.mit.edu/ mpg/haploview/). Haplotype effects in different genetic models were tested using HAPSTAT 3.0 (http://www.bios.unc.edu/~lin/hapstat/).

Results

A total of 436 participants were recruited: 229 patients with at least moderate periodontitis (as defined earlier) and 207 periodontitis-free controls. The mean age (\pm standard deviation) of the participants was 43.1 ± 7.7 years; 249 (57.1%) subjects were women. There were no differences in mean age and gender distribution between patient and control groups. The case group had about twice the proportion of smokers as did the control group and significantly fewer standing teeth (Table 1). Clinical parameters such as probing pocket depth \geq 5 mm, probing attachment level \geq 5 mm and bleeding on probing percentage were also summarized in Table 1.

Eleven tagging SNPs of the C5 gene were identified as candidate SNPs using the SNP Tagging Wizard of SNPBROWSER 3.5. Most were intronic SNPs. Ten of the 11 SNPs were genotyped successfully using the iPLEX assay and seven were found to have more than one genotype (minor allele frequency > 0.001) with an average call rate of 99.2%. One SNP (rs10818491) did not fulfill the Hardy-Weinberg Equilibrium (p < 0.01)(Table 2); hence, analyses of the genotype and allele-type distribution differences of the remaining six SNPs among the subgroups aggressive periodontitis, chronic periodontitis and controls was carried out. None of the six SNPs showed significant differences in the distribution of alleles or genotypes between either of the patient groups vs. the control group (adjusted *p*-value 0.008) (Table 3), which indicated that there was no difference among aggressive periodontitis, chronic periodontitis and control groups. Therefore, in the following stepwise logistic regression analysis, aggressive periodontitis and chronic periodontitis data were combined.

The six C5 SNPs fulfilling the criteria for genetic analysis (Table 3), together with age, gender and smoking habit, were selected for stepwise logistic regression analysis regarding the association between these possible risk indicators and periodontitis.

After the adjustment for age, gender and smoking, only rs17611 and smoking

Characteristic	Categories	Periodontitis-free $(n = 207)$	Periodontitis $(n = 229)$	Test	Statistics	<i>p</i> -value
Age (years)	Mean + SD	42.4 + 8.1	437 + 74	t	-1.69	NS
Gender	Male	87 (42.0)	100 (43.7)	γ^2	0.27	NS
	Female	120 (58.0)	129 (56.3)	7.		
Smoking habit	Nonsmoker Smokar	191 (92.3)	193 (84.3)	χ^2	7.15	< 0.005
	Pack-year (mean \pm SD)	4.7 ± 6.7	16.0 ± 10.2	t	-3.80	< 0.05
Teeth remaining	Mean ± SD	27.0 ± 1.8	$24.7~\pm~3.4$	t	5.46	< 0.001
Percentage of sites with a probing pocket depth $\geq 5 \text{ mm}$	Mean \pm SD	0	$30.3~\pm~18.1$			
Percentage of sites with a probing attachment level of $\geq 5 \text{ mm}$	Mean \pm SD	0	$47.2~\pm~27.2$			
Bleeding on probing percentage	Mean ± SD	ND	72.6 ± 25.0			

Results are no. (%) unless otherwise indicated.

ND, not determined; SD, standard deviation.

Table 2. Candidate complement component 5 (C5) single nucleotide polymorphisms (SNPs) of study participants

SNP ID	Туре	Variation	Call rate (%)	MAF ^a	HWE <i>p</i> -value ^b
rs2300930	Intronic	A/C	98.9	0.197	0.73
rs10818491	Intronic	C/T	98.7	0.406	$< 10^{-5}$
rs7035682	Intronic	C/T	99.3	Single genotype detected	
rs7026551	Intronic	A/C	98.4	Single genotype detected	
rs2269066	Intronic	A/G	98.2	0.237	0.20
rs2269067	Intronic	C/G	99.1	Single genotype detected	
rs1035029	Intronic	C/T	99.1	0.409	0.93
rs17611	Nonsynonymous	A/G	99.8	0.409	0.73
rs25681	Synonymous	C/T	99.8	0.409	0.96
rs992670	Intronic	A/G	99.1	0.230	0.24
rs2300934	Intronic	A/C	0	ND	

 $^{\rm a}{\rm MAF}$ was not determined when the call rate was < 80.0% or only one genotype was detected.

^bFor the periodontitis-free group, the HWE *p*-value was not determined if a call rate of < 80.0% or only one genotype was detected; only SNPs with a HWE *p*-value of ≥ 0.01 are subjected to further analysis.

HWE, Hardy-Weinberg equilibrium; MAF, minor allele frequency; ND, not determined.

habit were significantly associated with periodontitis (p = 0.023 and 0.006, respectively, Table 4); genotype AG of rs17611 seemed to be more frequent in periodontitis (p = 0.007, OR = 6.08, 95% CI = 1.31–28.22). Smoking was also significantly associated with periodontitis in this Hong Kong Chinese sample, having an OR of 2.84 (95% CI = 1.31–6.14, Table 4).

Haploview 3.32 was used to detect any LD block and the association between haplotypes and periodontitis (19) for the six SNPs that had a Hardy-Weinberg Equilibrium test *p*-value of $> 10^{-5}$, a call rate of > 0.8and a minor allele frequency of > 0.001. The pairwise comparisons were designed to identify markers within 500 kb from each other, and one LD block, including SNPs rs1035029, rs17611, rs25681 and rs992670, was detected (Fig. 1). The haplotype CGCA in this block was found to be marginally significantly associated with periodontitis (p = 0.038; Table 5). Then, HAPSTAT 3.0 was used to estimate haplotype effects and haplotype-environment interactions (age, gender and smoking habit, in particular, in this study) under different genetic models such as dominant and additive models (20). While dominant genetic model means that the

heterozygote has the same increased risk as minor homozygous genotypes (21), additive model is a statistical model modified from several regression models (22). No environment or haplotype–environmental interaction was found to be significantly associated with periodontitis. Haplotype CGCA was found to be significantly associated with periodontitis in the dominant model (p = 0.001, OR = 4.85, 95% CI = 1.85–12.71) (Table 5). Other genetic models were not suitable for our data set and therefore were not calculated.

Discussion

It is well known that the complement system is a biochemical cascade that helps to clear pathogens from an organism. Humoral activation of the complement system by the classical, alternative, or lectin pathways results in the cleavage of C3 into C3a and C3b. Subsequent downstream cleavage of C5 generates the anaphylatoxin C5a and fragment C5b, which acts as a nucleus for the MAC (23) by anchoring the assembly of a molecule each of C6, C7 and C8, to guide the polymerization of C9 into a membrane channel in the target cell (24). Not only does pore formation cause direct cell injury and necrosis (25), but it may also amplify the inflammatory response by promoting the expression of proinflammatory mediators (26). Additionally, the MAC can influence the recruitment of inflammatory cells and leukocyte adhesion to the endothelium (27,28), thereby promoting the release of cell stimulants, such as hydrolytic enzymes, reactive oxygen species and cytokines (29,30). Hence, mutations in any of the MAC complement components might modify this inflammatory response to pathogens. In this study of Hong Kong Chinese patients with and without periodontitis, genotype AG of nonsynonymous SNP rs17611 in the C5 gene and the haplotype containing rs17611 were found to be significantly associated with periodontitis, indicating that this genetic variation and haplotype might play a role in the pathogenesis of periodontitis.

The C5 gene is located on chromosome 9q34.1. Eleven SNPs were selected in this study. Although most of the SNPs selected were in introns and their biological function was unclear, the nature of other regional SNPs can be obtained through investigation of their linkage and haplotype status with those tagging SNPs. Therefore, information about the search range of SNPs, and hence the true causative variation(s), could potentially be identified in subsequent experiments. The haplotype and tagging polymorphisms of C5 have been studied in diseases such as liver fibrosis, rheumatoid arthritis and asthma (11-16). The haplotype including the nonsynonymous SNP rs17611 (A/G) in exon 19 has been reported to be associated with bronchial asthma and liver fibrosis (12,14). The A to G variation that was found to be associated with periodontitis in this study causes an amino acid change from isoleucine to valine at position 802. Because these two amino acids are both aliphatic and their structures are similar except for an extra center of asymmetry in the isoleucine side chain, the change from isoleucine to valine can be considered to be conservative.

Classically, it has been assumed that a mutant protein is unstable and leads

		Count ^a (%)				
SNP ID	Genotype/allele type	Periodontitis-free $(n = 207)$	Aggressive periodontitis (n = 32)	Chronic periodontitis $(n = 197)$	<i>p</i> -value	
rs2300930	AA	7 (3.4)	1 (3.1)	7 (3.6)	0.278	
	AC	57 (27.9)	8 (25.0)	73 (37.4)		
	CC	140 (68.6)	23 (71.9)	115 (59.0)		
	А	71 (17.4)	10 (15.6)	87 (22.3)	0.156	
	С	337 (82.6)	54 (84.4)	303 (79.7)		
rs2269066	AA	118 (57.6)	19 (59.4)	110 (56.4)	0.487	
	AG	75 (36.6)	11 (34.4)	80 (41.0)		
	GG	12 (5.9)	2 (6.2)	5 (2.6)		
	А	311 (75.9)	49 (76.6)	300 (76.9)	0.938	
	G	99 (24.1)	15 (23.4)	90 (23.1)		
rs1035029	CC	36 (17.6)	5 (15.6)	30 (15.4)	0.055	
	CT	87 (42.6)	13 (40.6)	110 (56.4)		
	TT	81 (39.7)	14 (43.8)	55 (28.2)		
	С	159 (39.0)	23 (35.9)	170 (43.6)	0.294	
	Т	249 (61.0)	41 (64.1)	220 (56.4)		
rs17611	AA	82 (40.2)	13 (40.6)	56 (28.7)	0.053	
	AG	85 (41.7)	14 (43.8)	110 (56.4)		
	GG	37 (18.1)	5 (15.6)	29 (14.9)		
	А	249 (61.0)	40 (62.5)	222 (56.9)	0.43	
	G	159 (39.0)	24 (37.5)	168 (43.1)		
rs25681	CC	37 (18.1)	6 (18.8)	28 (14.4)	0.033	
	CT	86 (42.2)	12 (37.5)	111 (56.9)		
	TT	81 (39.7)	14 (43.8)	56 (28.7)		
	С	160 (39.2)	24 (37.5)	167 (42.8)	0.685	
	Т	248 (61.8)	40 (62.5)	223 (57.2)		
rs992670	AA	117 (57.1)	19 (59.4)	115 (59.0)	0.178	
	AG	75 (36.6)	11 (34.4)	77 (39.5)		
	GG	13 (6.3)	2 (6.2)	3 (1.5)		
	А	309 (75.4)	49 (76.6)	307 (78.7)	0.529	
	G	101 (24.6)	15 (23.4)	83 (21.3)		

Table 3. Genotype and allele-type of complement component 5 (C5) single nucleotide polymorphisms (SNPs) selected for stepwise logistic regression analysis

^aTotal not adding up because call rates are < 100%.

^bThe level of significance was set at 0.008 after adjustment for multiple comparison.

to a functional defect, so subjects carrying a homozygous mutant genotype would be more susceptible than others to disease. However, a mutated gene may also encode a stable mutant protein that interferes with the formation of a functional form of the wild-type protein, especially for structural proteins or proteins forming dimers (31). An individual carrying a heterozygous genotype in this situation would be more susceptible than one carrying homozygous genotypes (31,32) because products from both genotypes are stable. In this way, the protein with the conservative mutation for valine at position 802, which is expected to be stable, might theoretically affect the wild-type protein. The amino acid mutation in rs17611 is in the MG6

domain of C5, which forms a conserved large cavity with other domains (33). Although the amino acid is far from the cleavage site, this cavity area may be involved in the recognition of C5 by the cleavage enzyme C5 convertase, which is a large macromolecular complex that could conceivably interact with this domain (34). The alteration in rs17611 could also potentially affect the formation of the C5b-9 MAC because the substituted amino acid would theoretically be situated within the C5b peptide after cleavage. Further studies are required on the functions of this gene variant and its effect on periodontitis risk. Additionally, the significant association in the dominant model of a haplotype consisting of SNPs rs1035029,

rs17611, rs25681 and rs992670 indicates that SNPs within this gene region could be associated with periodontitis, and further investigation should focus on this particular region.

Smoking is a well-established risk factor in the incidence and progression of periodontal diseases (35-39). One community study conducted among the Hong Kong population showed that heavy smoking is the major risk factor for periodontitis (OR = 4.61), and that moderate and light smoking are also significant risk factors (ORs = 2.69 and 2.33, respectively)(40). These values are similar to the OR (95% CI) of 2.84 (1.31-6.14) calculated in our study. Nevertheless, we could not subcategorize the small number of smokers as heavy, moderate, light and very light smokers, which may explain why the OR for smoking was lower than the published value for heavy smoking. The value was also lower than the OR (95% CI) of 6.08 (1.31-28.22) calculated for the rs17611 genetic variant in this study. It must be noted, however, that our strict criterion for a case was any patient with at least moderate periodontitis, and smoking and the rs17611 genetic variant could have a different influence on mild to moderate periodontitis. Finally, other factors, such as age and gender, have been reported to be associated with periodontitis in Hong Kong Chinese patients (40), but the effects of these two factors were removed by our case-control study design.

It should be noted that in our study we used extra-oral radiographs instead of intra-oral radiographs, despite the fact that OPG has a sensitivity of only 79% for periodontal pathology (41). Nevertheless, OPG has a reported specificity of 92% for periodontal pathology, and studies have confirmed that bone loss measured on OPG is closely correlated with periodontal status (42,43). In our study, it was necessary for a simple and robust method to be adopted in order to screen a large number of subjects and to eliminate any ambiguous subjects whose periodontal condition did not fulfill the recruitment criteria of either group, and therefore OPG was selected

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Table 4. Odds ratios (ORs) for study participants with periodontitis

Variable	Categories	Unadjusted (univariate) ^a		Adjusted ^b			
		OR	<i>p</i> -value	95% CI	OR	<i>p</i> -value	95% CI
Age		1.02	0.072	0.99-1.05			
Gender	Female	1	0.601				
	Male	0.9		0.62-1.32			
Smoking habit	Nonsmoker	1	0.003		1	0.006	
	Smoker	2.57		1.34-4.92	2.84		1.31-6.14
Genotype (SNP)							
rs2300930	AA	1.00	0.22				
	AC	1.24	0.69	0.43-3.62			
	CC	0.86	0.781	0.3-2.44			
rs2269066	AA	1	0.338				
	AG	1.11	0.605	0.75-1.65			
	GG	0.53	0.202	0.2-1.4			
rs1035029	TT	1	0.051			NS	
	СТ	1.66	0.024	1.09-2.53			
	CC	1.14	0.667	0.68-2.01			
rs17611	AA	1	0.026		1	0.023	
	AG	1.73	0.014	1 14-2 65	6.08	0.007	1 31-28 22
	GG	1.09	0.775	0.62-1.92	1.15	0.707	0.63-1.97
rs25681	TT	1	0.044	0.02 1.02		NS	0102 1197
1525001	CT	1.65	0.024	1.08-2.52		145	
	CC	1.06	0.886	0.6-1.87			
rs990032670	AA	1	0.12	010 1107			
100000000000000000000000000000000000000	AG	1.02	0.905	0.69-1.52			
	GG	0.34	0.044	0.12-0.97			
Allele-type							
rs2300930	C	1					
182300930	Δ	1 28	0.147	0.91_1.8			
rs2260066	G	1.20	0.147	0.91 1.0			
132207000	Δ	0.93	0.663	0.68-1.28			
ma1025020	C	1	0.005	0.00 1.20			
131055027	т	0.86	0.291	0.66-1.13			
rs17611	1	1	0.271	0.00 1.15			
	G	1 15	0.307	0.88 1.51			
m 25691	C	1.15	0.507	0.00 1.51			
1525001	т	0.02	0.531	0712			
rs992670	1 Δ	0.92	0.331	0.7-1.2			
13772070	G	0.84	0.288	0.61 1.16			
	U	0.04	0.200	0.01-1.10			

^aThe OR for each factor was computed from logistic regression.

^bAdjusted ORs were computed using stepwise logistic regression with variables that were significant in the univariate analysis.

CI, confidence interval; NS, not significant; SNP, single nucleotide polymorphism.

for assess the bone loss level of the subjects. We used the Schei ruler, without using the 1 mm space or what was referred to be the 'normal' distance from the alveolar crest to the cemento– enamel junction; instead, a certain level or fraction of 'bone loss' was allowed for the control group, for the reason that OPG typically magnifies tooth-length ranges from 15 to 30% (44,45). Considering that the average permanent tooth root length is about 11–17 mm (46), 1 mm equals about 5–9% root length from the cemento–enamel

junction to the radiographic apex. In order to provide some leeway for measurement, we used 15% root length as the cut-off, when some other studies considered 2 mm as the 'normal' distance from the alveolar crest to the the cemento–enamel junction (47). All these subjects selected by OPG screening were subsequently periodontally examined to confirm their eligibility as per our inclusion criteria.

In conclusion, our study showed that genotype AG of the nonsynonymous SNP rs17611 of the C5 gene was more prevalent in the periodontitis patient group than in periodontitis-free controls and, together with smoking, may be associated with periodontitis among the Hong Kong Chinese population. In addition, one haplotype including SNP rs17611 was associated with periodontitis. An association study with a large sample size and other ethnic populations are needed to confirm this finding. Further studies should also investigate how genetic variation may alter the structure and biological function of C5 and its



Fig. 1. Pairwise linkage disequilibrium (LD) in the complement component 5 (C5) gene. All single nucleotide polymorphisms (SNPs) of C5 were assessed in this study. The horizontal bar represents the relative location of each SNP along the chromosome 9, 9q33-q34, and the numbers above the bar are the corresponding chromosome positions; SNPs bolded are coding SNPs. Diamonds in the haplotype blocks represent pairwise linkage disequilibrium between all SNPs assessed; as shown in the figure key, the darker the diamond, the stronger the linkage disequilibrium between the two SNPs.

Table 5. Haplotype association results in different genetic models (n = 872)

	Count ratio ^b (freque	ount ratio ^b (frequencies)			
Haplotype ^a	Periodontitis-free	Periodontitis	χ^2 <i>p</i> -value ^c	Additive model	Dominant model
TATA	240.9/169.1 (0.588)	245.9/201.1 (0.599)	0.395	0.194	0.078
CGCG	95.7/314.3 (0.233)	93.6/362.4 (0.205)	0.319	0.308	0.323
CGCA	59.3/350.7 (0.145)	90.3/365.7 (0.198)	0.038	0.029	0.001 ^e
TACA	4.8/405.2 (0.012)	5.1/450.9 (0.011)	0.950	0.216	0.845

^aTwo haplotypes are not listed because of low frequencies (fractional likelihood frequency < 0.01).

^bCounts were obtained by summing the fractional likelihoods of each individual for each haplotype.

^cThe level of significance was set at 0.0125 after adjustment for multiple comparison.

^dLogistic regression analysis *p*-values are shown.

eOdds ratio and 95% confidence interval: 4.85 (1.85-12.71).

fragments, and how these alterations modify humoral immune responses and susceptibility to periodontitis among the Hong Kong Chinese population, and possibly among other populations.

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References

- Yoshie H, Kobayashi T, Tai H, Galicia JC. The role of genetic polymorphisms in periodontitis. *Periodontol* 2000 2007; 43:102–132.
- Carroll MC. The complement system in regulation of adaptive immunity. *Nat Immunol* 2004;5:981–986.
- Kohl J, Wills-Karp M. Complement regulates inhalation tolerance at the dendritic cell/T cell interface. *Mol Immunol* 2007; 44:44–56.
- Schenkein HA, Genco RJ. Gingival fluid and serum in periodontal diseases. II. Evidence for cleavage of complement components C3, C3 proactivator (factor B) and C4 in gingival fluid. *J Periodontol* 1977;**48**:778–784.
- Niekrash CE, Patters MR. Simultaneous assessment of complement components C3, C4, and B and their cleavage products in human gingival fluid. I. Reliability of the method. *J Periodontal Res* 1985; 20:260–267.
- Niekrash CE, Patters MR. Assessment of complement cleavage in gingival fluid in humans with and without periodontal disease. J Periodontal Res 1986;21:233– 242.
- Seppanen M, Lokki ML, Notkola IL et al. Complement and C4 null alleles in severe chronic adult periodontitis. Scand J Immunol 2007;65:176–181.
- Gerard NP, Gerard C. The chemotactic receptor for human C5a anaphylatoxin. *Nature* 1991;349:614–617.
- Gerard C, Gerard NP. C5a anaphylatoxin and its seven transmembrane-segment receptor. *Annu Rev Immunol* 1994;12:775– 808.
- Podack ER, Esser AF, Biesecker G, Muller-Eberhard HJ. Membrane attack complex of complement: a structural analysis of its assembly. *J Exp Med* 1980;151:301–313.
- Karp CL, Grupe A, Schadt E *et al.* Identification of complement factor 5 as a susceptibility locus for experimental allergic asthma. *Nat Immunol* 2000;1:221–226.
- Hasegawa K, Tamari M, Shao C *et al.* Variations in the C3, C3a receptor, and C5 genes affect susceptibility to bronchial asthma. *Hum Genet* 2004;115:295–301.
- Hillebrandt S, Wasmuth HE, Weiskirchen R et al. Complement factor 5 is a quantitative trait gene that modifies liver fibrogenesis in mice and humans. Nat Genet 2005;37:835–843.
- 14. Gressner O, Meier U, Hillebrandt S et al. Gc-globulin concentrations and C5

haplotype-tagging polymorphisms contribute to variations in serum activity of complement factor C5. *Clin Biochem* 2007; **40**:771–775.

- Kurreeman FA, Padyukov L, Marques RB et al. A candidate gene approach identifies the TRAF1/C5 region as a risk factor for rheumatoid arthritis. PLoS Med 2007:4:e278.
- Plenge RM, Seielstad M, Padyukov L et al. TRAF1-C5 as a risk locus for rheumatoid arthritis – a genomewide study. N Engl J Med 2007;357:1199–1209.
- Bassiouny MA, Grant AA. The accuracy of the Schei ruler: a laboratory investigation. J Periodontol 1975;46:748–752.
- Armitage GC. Development of a classification system for periodontal diseases and conditions. *Ann Periodontol* 1999;4:1–6.
- Teo YY, Fry AE, Clark TG, Tai ES, Seielstad M. On the usage of HWE for identifying genotyping errors. *Ann Hum Genet* 2007;71:701–703.
- Lin DY, Zeng D, Millikan R. Maximum likelihood estimation of haplotype effects and haplotype – environment interactions in association studies. *Genet Epidemiol* 2005;29:299–312.
- Demchuk E, Yucesoy B, Johnson VJ et al. A statistical model for assessing genetic susceptibility as a risk factor in multifactorial diseases: lessons from occupational asthma. Environ Health Perspect 2007; 115:231–234.
- Hastie TJ, Tibshirani RJ Generalized Additive Models. London: Chapman & Hall/CRC, 1990.
- Schmidt RE, Gessner JE. Fc receptors and their interaction with complement in autoimmunity. *Immunol Lett* 2005;100: 56–67.
- Fondevila C, Shen XD, Tsuchihashi S et al. The membrane attack complex (C5b-9) in liver cold ischemia and reperfusion injury. *Liver Transpl* 2008;14:1133–1141.
- Biancone L, David S, Della Pietra V, Montrucchio G, Cambi V, Camussi G. Alternative pathway activation of complement by cultured human proximal tubular epithelial cells. *Kidney Int* 1994; 45:451–460.
- 26. Schonermark M, Deppisch R, Riedasch G, Rother K, Hansch GM. Induction of mediator release from human glomerular mesangial cells by the terminal

complement components C5b-9. Int Arch Allergy Appl Immunol 1991;**96:**331– 337.

- 27. Hattori R, Hamilton KK, McEver RP, Sims PJ. Complement proteins C5b-9 induce secretion of high molecular weight multimers of endothelial von Willebrand factor and translocation of granule membrane protein GMP-140 to the cell surface. J Biol Chem 1989;264:9053–9060.
- Kilgore KS, Shen JP, Miller BF, Ward PA, Warren JS. Enhancement by the complement membrane attack complex of tumor necrosis factor-alpha-induced endothelial cell expression of E-selectin and ICAM-1. *J Immunol* 1995;155:1434–1441.
- Seeger W, Suttorp N, Hellwig A, Bhakdi S. Noncytolytic terminal complement complexes may serve as calcium gates to elicit leukotriene B4 generation in human polymorphonuclear leukocytes. *J Immunol* 1986;137:1286–1293.
- Hansch GM, Seitz M, Betz M. Effect of the late complement components C5b-9 on human monocytes: release of prostanoids, oxygen radicals and of a factor inducing cell proliferation. *Int Arch Allergy Appl Immunol* 1987;82:317–320.
- Sidransky E. Heterozygosity for a Mendelian disorder as a risk factor for complex disease. *Clin Genet* 2006;**70:**275–282.
- 32. Sanders CR, Ismail-Beigi F, McEnery MW. Mutations of peripheral myelin protein 22 result in defective trafficking through mechanisms which may be common to diseases involving tetraspan membrane proteins. *Biochemistry* 2001; 40:9453–9459.
- Fredslund F, Laursen NS, Roversi P et al. Structure of and influence of a tick complement inhibitor on human complement component 5. Nat Immunol 2008;9:753– 760.
- 34. Sandoval A, Ai R, Ostresh JM, Ogata RT. Distal recognition site for classical pathway convertase located in the C345C/ netrin module of complement component C5. J Immunol 2000;165:1066–1073.
- Grossi SG, Zambon JJ, Ho AW et al. Assessment of risk for periodontal disease.
 Risk indicators for attachment loss. J Periodontol 1994;65:260–267.
- 36. Grossi SG, Genco RJ, Machtei EE *et al.* Assessment of risk for periodontal disease.

II. Risk indicators for alveolar bone loss. *J Periodontol* 1995;66:23–29.

- Johnson GK. Position paper: tobacco use and the periodontal patient. Research, Science and Therapy Committee of the American Academy of Periodontology. *J Periodontol* 1999;**70**:1419–1427.
- Johnson GK, Hill M. Cigarette smoking and the periodontal patient. J Periodontol 2004;75:196–209.
- Ryder MI. The influence of smoking on host responses in periodontal infections. *Periodontol 2000* 2007;43:267–277.
- Ng SK, Leung WK. A community study on the relationship between stress, coping, affective dispositions and periodontal attachment loss. *Community Dent Oral Epidemiol* 2006;**34**:252–266.
- Balis S. Error and accuracy rates of panoramic radiography as a screening method for mass surveying of children. J Public Health Dent 1981;41:220–234.
- Walsh TF, al-Hokail OS, Fosam EB. The relationship of bone loss observed on panoramic radiographs with clinical periodontal screening. *J Clin Periodontol* 1997;24:153–157.
- 43. Persson RE, Tzannetou S, Feloutzis AG, Bragger U, Persson GR, Lang NP. Comparison between panoramic and intra-oral radiographs for the assessment of alveolar bone levels in a periodontal maintenance population. J Clin Periodontol 2003;30: 833–839.
- Thanyakarn C, Hansen K, Rohlin M, Akesson L. Measurements of tooth length in panoramic radiographs. 1. The use of indicators. *Dentomaxillofac Radiol* 1992; 21:26–30.
- Thanyakarn C, Hansen K, Rohlin M. Measurements of tooth length in panoramic radiographs. 2: Observer performance. *Dentomaxillofac Radiol* 1992; 21:31–35.
- Ash M, Wheeler R, Nelson S Wheeler's Dental Anatomy, Physiology and Occlusion, 8th edition. Philadelphia, PA, London: Sauders, 2003.
- 47. Lanning SK, Best AM, Temple HJ, Richards PS, Carey A, McCauley LK. Accuracy and consistency of radiographic interpretation among clinical instructors in conjunction with a training program. *J Dent Educ* 2006;**70:**545–557.

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