J Periodont Res 2011; 46: 397–406 All rights reserved

Mini Review

FoxOs, Wnts and oxidative stress-induced bone loss: new players in the periodontitis arena?

Galli C, Passeri G, Macaluso GM. FoxOs, Wnts and oxidative stress-induced bone loss: new players in the periodontitis arena? J Periodont Res 2011; 46: 397-406. © 2011 John Wiley & Sons A/S

Background and Objective: Chronic periodontitis is a widespread disease affecting tooth-supporting structures that can lead to extensive loss of periodontal ligament and bone, ultimately resulting in tooth loss. Extensive evidence has demonstrated a strong association between age, metabolic disorders such as type II diabetes, oxidative stress and alveolar bone loss. The molecular players controlling bone maintenance and underlying age-related bone loss and its links to the general metabolism are currently the object of intense research.

Material and Methods: Recent findings are summarized to elucidate the molecular mechanisms linking oxidative stress, bone loss and metabolic factors.

Results: It is well known that reactive oxygen species are an inevitable consequence of cellular respiration and that organisms have developed an efficient array of defenses against them. The core of this complex defense line is a family of transcription factors, known as FoxOs, which can bind to β -catenin and initiate a transcriptional programme regulating cell apoptosis, DNA repair and degradation of reactive oxygen species. An increase in reactive oxygen species due, for example, to age or insulin resistance, generates a situation in which bone formation is impaired by activation of FoxO, and a decrease in Wnt signaling and bone resorption are promoted.

Conclusion: The balance between FoxO and the Wnt pathway is finely tuned by systemic and local factors, creating a far-reaching mechanism that dictates the fate of mesenchymal progenitors and regulates the homeostasis of bone, providing a rationale for the impairment of systemic and alveolar bone maintenance clinically observed with age and metabolic diseases.

Periodontis is an inflammatory disease that affects tooth-supporting structures and can lead to loss of periodontal ligament and alveolar bone, with increased tooth mobility and eventually tooth loss. Its pathogenesis is multifactorial and involves the periodontal microbiota, the host's immune responses and behavioral or concomitant medical conditions. There appears to be an epidemiological association between periodontitis and metabolic diseases, including diabetes, and an association between periodontitis and aging has long been suggested, although not conclusively proved (1–13). Importantly, the main factor that seems to be linking all these medical

C. Galli^{1,2}, G. Passeri¹, G. M. Macaluso² ¹Department of Internal Medicine and ²Unit of Periodontology, University of Parma, Parma,

© 2011 John Wiley & Sons A/S JOURNAL OF PERIODONTAL RESEARCH doi:10.1111/j.1600-0765.2011.01354.x

Italy

Carlo Galli, DDS, PhD, Unit of Periodontics, University of Parma, Via Gramsci 14, 43100 Parma, Italy Tel: +39 0521 986722 Fax: +39 0521 292955 e-mail: carlo.galli@unipr.it

Key words: periodontal disease; oxidative stress; FoxO; Wnt

Accepted for publication December 21, 2010

conditions is increased oxidative stress (3,14–16). Humans live in an oxygenrich environment, and their survival depends on a subtle mitochondrial electric current that generates ATP, the common energy source for all cells. Electrons, however, can escape from the mitochondrial transport chain and generate reactive oxygen species

Fig. 1. Diagram showing generation of reactive oxygen species (ROS) in the mitochondria and antioxidant cellular defenses. Hypoxia, aging and inflammation can increase the formation of highly reactive oxygen species, such as superoxide radicals (O_2^-) . These can be converted to H_2O_2 by the enzyme superoxide dismutase (SOD). Hydrogen peroxide can then be converted to H_2O by glutathione or the enzyme catalase. Accumulation of ROS in the cells can lead to cellular damage. GSSG, oxidized glutathione; GSH, reduced glutathione.

(ROS), such as hydrogen peroxide $(H₂O₂)$, oxygen superoxide $(O₂⁻)$ or the hydroxyl radical (OH⁻; Fig. 1; 17-19). These are highly reactive oxygen radicals that are responsible for most oxidative stress in cells (20–22). Although low levels of ROS are, in fact, necessary for several cellular signaling pathways, such as ERKs, p38 MAPK and tyrosine phosphatases (23,24), their continuous production within cellular organelles causes cellular microdamage that accumulates over time (25–28). Broadly speaking, oxidative stress can cause cellular damage and death in many tissues, and it has been inversely correlated with lifespan in nematodes, flies and mammals (29), to the point of being considered the primary force driving aging. Increased oxidative stress is a hallmark of inflammation, because it can be produced by neutrophils as a defense against invading microorganisms or by the microorganisms themselves, and this is well known to contribute to tissue damage (30–34). Importantly, several pathologic conditions, such as wound healing, ischemia, or diabetesrelated changes in the microvasculature, are associated with hypoxia, a shortage of oxygen supply (35,36). Although apparently in contrast, hypoxia and generation of ROS are coupled processes. During hypoxia, ROS can be released into the cytoplasm as a consequence of mitochondrial failure. Reactive oxygen species have also been demonstrated to impair the immune responses to microorganisms, and several reports directly suggest that oxidative stress is an important factor in the pathogenesis of periodontitis (30,37–39). Moreover, recent clinical studies have demonstrated a higher concentration of oxygen metabolites in the serum of periodontal patients (40,41), a reduction in the serum levels of oxidized low-density lipoproteins (42) or an increase in total antioxidant capacity after periodontal treatment (43–45). Some studies showed a relation between serum (46,47) or salivary antioxidants (48,49) as predictors of the insurgence of periodontitis or periimplantitis.

As aging and a wealth of metabolic disorders have been associated with both systemic and alveolar bone loss, and the same conditions are also characterized by a marked increase in oxidative stress, the idea that oxidative stress is the culprit for bone loss is becoming increasingly attractive (3,50). To support this hypothesis, some recent studies have shown that bone formation in young mice is decreased by inhibiting the antioxidant glutathione, that increased lipid oxidation may reduce pro-osteogenic stimuli in the skeleton (51) and that administration of the antioxidant N-acetylcysteine reverses bone loss in a murine model of estrogen deficiency (52–55). Interestingly, Toker et al. (31) showed that the same antioxidant, N-acetylcysteine, can reduce alveolar bone loss in a rat periodontitis model, while Tomofuji et al. (56) demonstrated that a cocoaenriched diet protected rats from periodontitis-induced alteration in serum antioxidant levels and inhibited alveolar bone loss.

The molecular mechanisms that link oxidative stress and bone loss are complex and still not fully understood. However, understanding them can provide a critical key to hamper or prevent bone loss in clinical conditions of increased oxidative stress. Recent exciting discoveries about the signal pathways controlling the balance between cell fate and the cellular defenses against ROS can provide a rationale for many important clinical observations and help identify potential therapeutic targets.

An association between periodontal disease and metabolic syndrome has been established. These medical conditions are characterized by increased oxidative stress. It has been shown that Reactive oxygen species can induce bone loss, thus providing a rationale for tissue destruction in periodontitis.

The FoxO family of transcription factors

Since oxidative stress is an inevitable and, within certain limits, not undesired side-effect of cell respiration, cells have developed several antioxidant mechanisms to contain ROS-mediated damage, while allowing them to function in cell signaling. Some of these defenses rely on thiol-containing peptides, such as glutathione and thioredoxine, which can reduce ROS into harmless alcohols (57). Cells can, however, also resort to more sophisticated transcriptional programmes, such as the ones controlled by FoxOs.

FoxOs are members of the O ('other') class of the Forkhead superfamily (58), originally called FKHRs (forkhead in rhabdomyosarcomas). Four members of this class are known at present, FoxO1, FoxO3, FoxO4 and FoxO6, of which FoxO1–3 are broadly expressed, whereas FoxO6 is restricted to the developing brain (60). They are all characterized by a 100 amino-acid helix–loop–helix DNA binding domain, called the Forkhead domain (61,62), which is recognized by a consensus sequence $(G/T)(T/A)AA$ (C/T)AA, called the FoxO-recognized element (FRE; 63–65). FoxOs can move from the nucleus, where they act as transcription factors, to the cytoplasm, where they are inactive, depending on their phosphorylation state (62). In the absence of growth factors or insulin or in the presence of stress stimuli, FoxOs reside in the nucleus and actively transcribe several target genes. Some of them are involved in apoptosis signaling, like the tumor-necrosis-factor-related apoptosis-inducing ligand (TRAIL), Fas ligand and Bim (66,67). Interestingly, it has been shown that the expression of a phosphorylation-resistant form of FoxO3 induces cell apoptosis (61). FoxOs can also induce cell cycle arrest (68–70) by up-regulating the expression of the cyclin-dependent kinase inhibitor, $p27^{KIP1}$ (71), of the retinoblastoma protein p130 (72) and downregulating cyclin D1 and D2 (73,74), a critical function, because an inactivation of FoxO activity can be found in 60–80% of prostate cancers (75). Fox-Os antagonize oxidative stress through the transcription of manganese superoxide dismutase (MnSOD), which catalyzes the transformation of O_2 ⁻ into H2O2, catalase, which neutralizes H2O2, and Growth Arrest and DNA Damage 45, which repairs DNA damage (68,76,77). By promoting cell cycle arrest and quiescence, FoxOs help cells to survive and counteract the action of ROS (67,78). The balance between the pro-apoptotic and the prosurvival actions of FoxOs depends on mechanisms not yet completely clear. However, it is known that the acetylation state of FoxO plays a role in the preferential expression of a subset of genes (79). Sirtuins (SIRTs) have been shown to deacetylate FoxOs in response to oxidative stress, facilitating the expression of genes involved in defense against ROS and cell cycle arrest (80–82).

The pivotal importance of FoxOs for the organism is highlighted by the dramatic consequences of FoxO deletion in mice. FoxO1 null mice die during embryonic development because of defects in angiogenesis (83,84), whereas FoxO3 null mice present with impaired fertility and lymphoprolipherative disorders (85,86). Most interestingly, conditional FoxO1,2,3 triple knockout mice show hemopoietic defects, because of increased ROS levels in the hemopoietic stem cells. Reactive oxygen species, in turn, deplete the stem cell reservoir in the bone marrow, by driving hemopoietic stem cells into the cell cycle and differentiation, which is reversed by administration of N-acetylcysteine (NAC), a potent antioxidant (78).

Two very recent works have greatly elucidated the importance of FoxO for bone maintenance. Rached et al. (87) showed that α 1(I) Collagen-Cre-mediated conditional deletion of FoxO1 from osteoblasts decreased bone mineral density in the spine and femur. Reduced osteoblast number, bone formation rate and bone volume were observed at histology. Moreover, FoxO1^{-/-} bone phenotype did not progress with age, so these mice lacked agerelated bone changes. At the cellular level, the authors showed a decrease in osteoblast proliferation and in antioxidant defense responses and, noticeably, FoxO1 overexpression or NAC administration reversed the bone phenotype. Ambrogini et al. (88) conditionally deleted FoxO1,3,4 using the interferoninducible transgene Mx-Cre, and demonstrated a reduction in bone mineral density, osteoblast number, bone formation rate and an increase in osteoblast apoptosis 5 wk after deletion. Conversely, FoxO3 overexpression increased bone mass and reduced oxidative-stress-induced apoptosis (88).

FoxO transcription factors induce the expression of genes controlling defenses against oxidative stress and cell survival. Genetic ablation of Fox-Os decreases bone mass and osteoblast numbers. Treatment with antioxidants can reverse this phenotype.

FoxO control by Akt: the metabolic link

FoxOs possess evolutionarily conserved phosphorylation sites for the survival kinase Akt near the Forkhead domain, on threonine 24, serine 256 and serine 319 (62,68). Akt, also known as protein kinase B, is a serine/ threonine kinase activated by a number of receptor tyrosine kinases and G protein-coupled receptors (89,90) through phosphatidyl inositol 3-kinase (PI3K) (91) and its product, phosphatidylinositol 3,4,5-triphosphate (PIP₃). PIP3 recruits Akt to the cell membrane, where it phosphorylates FoxO, which is bound by the 14-3-3 chaperone proteins and is thus retained in the cytoplasm. Akt activation is antagonized by the protein Klotho and by phosphatase and tensin homologue deleted on chromosome 10 (PTEN), which removes the 3' phosphate from $PIP₃$ and thereby attenuates $PI3K$ signaling (92). In the presence of ROS, PTEN is activated, and as a consequence Akt is down-regulated, removing its inhibition on FoxO, which is then free to initiate the transcription of defense factors against oxidative stress. Akt, in contrast, can also increase ROS generation by controlling cell metabolism and oxygen consumption. Akt1 and 2 double knockout mouse embryonic fibroblasts (MEFs) had significantly lower intracellular ROS levels than wild-type MEFs, whereas cells expressing activated Akt or MEFs from $PTEN^{-/-}$ mice showed increased ROS (93). Moreover, activation of Akt provides protection from apoptosis, leads to uncontrolled cell replication and hyperplastic lesions in SCID mice (94). Not unexpectedly then, it also sensitizes cells to oxidative damage (93,95).

The Akt–FoxO axis is of critical importance in the regulation of cell metabolism. Insulin, glucagon-like peptide 1 or insulin-like growth factor induce Akt activation, which in turns retains FoxOs outside the nucleus. The removal of FoxO-mediated inhibition of cyclins allows β -cells in the pancreas to proliferate actively, to supply more insulin when needed, but at the same time weakens cellular defenses against

ROS. Reactive oxygen species, however, can also activate FoxO through c-Jun Kinase (JNK)-mediated phosphorylation, regardless of Aktmediated insulin signaling, thus decreasing the cell responses to insulin, and therefore insulin sensitivity (Fig. 2). Notably, JNK deletion, FoxO1 (96) or PTEN haploinsufficiency (97), as well as NAC administration (98,99) or overexpression of ROS scavengers (100,101) improved insulin sensitivity in mice. Conversely, FoxO1 overexpression or the expression of a constitutively active FoxO1 impaired glucose metabolism and induced diabetes (102). To make things worse, hyperglycemia and insulin resistance create a situation in which the increased metabolism, due to increased presence of glucose, and proinflammatory cytokines generate more ROS. This requires higher levels of FoxO transcriptional activity to oppose cellular damage, at the expense of the response to insulin.

Metabolic stimuli, such as insulin or insulin-like growth factor, can activate Akt, an inhibitor of FoxOs, thus allowing pancreatic β -cells to proliferate and supply the required amount of insulin, but at the same time reducing defenses against oxidative stress. Reactive oxygen species, however, can activate FoxOs via JNK, independently of Akt, thus reducing insulin sensitivity. This might help explain why periodontal disease worsens diabetes control and vice versa.

FoxO and T-cell factor (TCF) signaling: β -catenin at the helm

To better understand the consequences of FoxO activation for bone, it must be remembered that FoxO transcriptional activity requires β -catenin, a protein of the Armadillo family and a normal constituent of cell-to-cell junctions. b-Catenin is an essential mediator of several pathways that control the cell fate. One of the best known is the Wnt canonical pathway (103–105). The canonical or Wnt/b-catenin pathway is activated upon binding of some secreted glycoproteins, called Wnt proteins, to Frizzled (Fz) and LRP5/6 receptor (106,107). This induces the activation of dishevelled (Dvl; 108), which releases β -catenin from a multimolecular complex it forms with gly-

Fig. 2. Insulin binding to membrane receptors in the pancreatic islets leads to Akt activation through PIP3. This in turn phosphorylates FoxO transcription factors, excluding them from the nucleus and promoting the activation of alternative pathways that induce cell proliferation, to produce more insulin. The presence of ROS, as a consequence of periodontitis, however, can activate FoxOs independently of Akt, leading to insulin resistance. PIP3, Phosphatidylinositol (3,4,5)-trisphosphate; JNK, c-Jun N-terminal Kinase; TCF, T-cell factor.

cogen synthase kinase 3 and casein kinase 1a, and two scaffold proteins, axin and adenomatous polyposis coli (109), which target β -catenin for proteosomal degradation (110,111). Once released, b-catenin can shuttle to the nucleus, where it binds to a member of the T-cell factor/lymphoid enhancer factor (TCF/Lef1) transcription factor family (112), thus promoting the expression of several target genes (113).

The β-catenin-mediated Wnt signaling is a pivotal pathway in the development of the embryo (114,115) and has been shown to control stem cell proliferation and differentiation (116). The canonical Wnt signaling is also required for osteoblast differentiation (117) and bone formation (118). Patients suffering with van Buchem disease present with dramatic osteopetrosis due to unopposed Wnt canonical signaling in bone owing to the lack of an osteocyte-derived β -catenin inhibitor, sclerostin (119–121). Furthermore, b-catenin is a co-mediator of the action of parathyroid hormone on bone formation (122), and it regulates bone remodeling by increasing the expression of osteoprotegerin (123), which acts as an antagonist of RANKL, the main inducer of osteoclastogenesis, and thus bone resorption. Akt can promote Wnt canonical signaling by phosphorylating glycogen synthase kinase 3 (124) and thus releasing β -catenin, and it has been demonstrated that Akt can exert a broad function in controlling skeletal development in the embryo by tuning the activation of the Akt–glycogen synthase kinase 3 or the Akt–FoxO axis (125).

b-Catenin is then at the crossroad between two alternative pathways with juxtaposed effects, the former, mediated by TCF, which promotes cells proliferation and commitment, and the latter, mediated by FoxOs, that induces quiescence, preservation of stemness and defenses against ROS (53,126).

To activate cellular defenses against ROS, therefore, FoxOs must compete with TCF/Lef1 to bind B-catenin, and it has been shown that induction of oxidative stress by ROS antagonizes Wnt canonical signaling, by diverting the pool of free cytosolic β -catenin away from TCF (52,127) to bind FoxO

in humans and rodents (137–139). PPAR- γ can bind to B-catenin and degrade it, while at the same time the increase in oxidative stress promotes $FoxO/\beta$ -catenin transcriptional activity, diverting b-catenin from binding TCF.

Oxidative stress and bone loss 401

Moreover, ROS play an important role in the formation, survival and resorbing activity of osteoclasts by up-regulating RANKL and tumor necrosis factor-a expression through ERK and nuclear factor- κ B activation (140–142). Inhibition of ROS by the antioxidant NAC prevents osteoclastogenesis (143). Furthermore, recent reports have shown that RANKL induces osteoclast formation by generating ROS in osteoclast precursors, and mice lacking the antioxidant gene peroxiredoxin II have reduced bone mass (143,144). Likewise, it has been reported that overexpression of chloroplastic glutathione peroxidase, an enzyme responsible for H_2O_2 degradation in osteoclast precursors, impairs osteoclast formation and RANKL signaling (142). Consistent with these results, Srinivasan et al. (145) demonstrated that hypoxia promoted osteoclast formation by increasing ROS and that homocysteine enhanced bone resorption through induction of ROS (146). Notably, there is important evidence that the effects on bone mass of estrogens and androgens, hormones that have been long associated with maintenance of bone homeostasis, are mediated by antioxidant effects, and that their effect in preventing ovariectomy-induced bone loss can be recapitulated by NAC (50,147,148). A recent report by Jilka *et al.* (149) also showed that the effects of intermittent parathyroid hormone on bone anabolism are more marked in aged bone compared with the young skeleton because of its antioxidant activity.

It is therefore possible to envisage a scenario where, in the presence of periodontitis, a localized and systemic state of oxidative stress is created (39,40), which activates a cascade of oxidation products and stimulates bone resorption while inhibiting bone formation. The cellular mechanisms that antagonize ROS in the presence of

Fig. 3. FoxOs and TCF compete for the co-transcription factor β -catenin. β -Catenin can thus initiate two alternative pathways with opposite effects, leading to cell quiescence, defenses against ROS and apoptosis or cell proliferation and differentiation. External stimuli, such as the presence of ROS, hormones and growth factors, can affect the balance between these two pathways.

(Fig. 3). The balance between the two pathways is regulated by the presence of stress factors; ROS activate JNK both independently of and via Dvl, and JNK in turn also phosphorylates FoxO, increasing its activity at the expense of b-catenin/TCF (128). JNK could also directly inactivate TCF by phosphorylating it, much like a similar kinase, Nemo-like kinase (129).

This may provide a rationale for the impaired bone formation and increased osteoblast apoptosis in diabetic rodents and for alveolar bone loss in diabetic rodents (1,130–132).

FoxOs and TCF compete for binding b-catenin. This co-transcription can thus participate in the activation of transcription programmes for defenses against oxidative stress or alternatively for osteoblast differentiation. Continuous stimulation of $FoxO/\beta$ -catenin by ROS results, therefore, in reduced bone formation.

Reactive oxygen species, lipid oxidation and bone resorption

An increase in ROS can oppose osteoblastogenesis and bone formation by another mechanism. Reactive oxygen species have been proved to enhance the activity of the lipoxygenase Alox-15 (51), converting polyunsaturated fatty acids, such as linoleic acid, to oxidized fatty acids, including 9-hydroxy-10,12-octadecadienoic acid., a high affinity ligand of the adipogenic transcription factor Peroxisome proliferator-activated receptor gamma (PPAR- γ) (133). A vicious cycle may ensue, because these oxidized compounds can also break into unsaturated aldehydes, such as 4-hydroxynonenal, which can deplete glutathione and thus further increase ROS (134). Sheikhi et al. (33) hypothesized that lipid peroxidation by activated neutrophils could be responsible for tissue damage in periodontal disease. As a matter of fact, pharmacologic inhibition of Alox15 has been shown to improve bone mass in mice (135). Adipogenesis and osteoblastogenesis are balanced and almost antagonistic processes in the skeleton. It has been demonstrated that PPAR- γ haploinsufficiency in adipocytes increases bone mass and osteoblastogenesis (136), whereas PPAR- γ stimulation by rosiglitazone or

Table 1. List of studies investigating the effects of antioxidants on periodontitis, the model used and their effect

Study	Year	Model	Antioxidant	Effect
Abou Sulaiman et al. (43)	2010	Periodontitis, human	Vitamin C	n.S.
Maruyama et al. (150)	2011	Periodontitis, rat	Green tea catechins	$^{+}$
Govindaraj et al. (151)	2010	Periodontitis, rat	Proanthocyanidin	$^{+}$
Toker et al. (31)	2009	Periodontitis, rat	N-Acetylcysteine	$^{+}$
Tomofuji et al. (152)	2009	Periodontitis, rat	Vitamin C	$+$ $(*)$
Tomofuji et al. (56)	2009	Periodontitis, rat	Cocoa	$^{+}$
Hirasawa et al. (154)	2002	Periodontitis, human	Green Tea catechins	$^{+}$
Khmelevskii et al. (153)	1985	Periodontitis, human	Vitamin A, E, K	$^+$

n.s., no significant effect observed; +, significant difference between treatment in the presence or in the absence of antioxidant; *, only oxidative parameters considered, no clinical outcome.

sustained oxidative stress are detrimental to bone formation and facilitate progressive damage to alveolar and periodontal structures.

Oxidative stress can exert a detrimental effect on bone by generating oxidized fatty acids, which stimulate adipogenesis and inhibit osteoblastogenesis, and by directly stimulating osteoclast formation and activity. Taken together, these mechanisms provide a framework in which periodontal and bone damage ensues as a consequence of a periodontitis-related highly oxidative state.

What next?

Attempts to counteract oxidative stress to improve the outcome of treatment of periodontitis with local or systemic factors have been reported (31,43,56, 150–154), and most of them appeared successfully to hamper periodontal destruction, at least in rodent models (Table 1). The available antioxidants have some limitations, however. N-Acetylcysteine, though effective in rodents, has been shown to inhibit Wnt canonical signaling, which might decrease its overall benefits (149). Improved antioxidants should be developed to avoid undesired effects, and novel therapeutic approaches should take advantage of known molecular pathways underlying cell defenses against ROS.

Conclusions

Oxidative stress is a central event for the fate of cells. Its ubiquitous presence has led to the development of a complex genetic network, co-ordinated by FoxOs and β -catenin, aiming to protect cells and balance cell activities to best oppose ROS. An increase in ROS, which may result from age or insulin resistance, generates a situation in which bone formation is impaired and bone resorption is strongly promoted. Understanding the mechanisms underlying ROS-mediated bone loss is the key to developing new therapeutic approaches to systemic and localized bone disorders, but also to periodontitis and possibly peri-implantitis.

Acknowledgements

The authors would like to thank Dr Simone Lumetti, Universiity of Parma, Dental School, for providing the clinical material for the illustrations.

References

- 1. Liu R, Bal HS, Desta T et al. Diabetes enhances periodontal bone loss through enhanced resorption and diminished bone formation. J Dent Res 2006;85:510-514.
- 2. Nishimura F, Taniguchi A, Yamaguchi-Morimoto M et al. Periodontal infection and dyslipidemia in type 2 diabetics: association with increased HMG-CoA reductase expression. Horm Metab Res 2006;38:530–535.
- 3. Bullon P, Morillo JM, Ramirez-Tortosa MC, Quiles JL, Newman HN, Battino M. Metabolic syndrome and periodontitis: is oxidative stress a common link? J Dent Res 2009;88:503–518.
- 4. Chen L, Wei B, Li J et al. Association of periodontal parameters with metabolic

level and systemic inflammatory markers in patients with type 2 diabetes. J Periodontol 2010;81:364–371.

- 5. Li P, He L, Sha YQ, Luan QX. Relationship of metabolic syndrome to chronic periodontitis. J Periodontol 2009;80:541–549.
- 6. D'Aiuto F, Sabbah W, Netuveli G et al. Association of the metabolic syndrome with severe periodontitis in a large U.S. population-based survey. J Clin Endocrinol Metab 2008;93:3989–3994.
- 7. Shimazaki Y, Saito T, Yonemoto K, Kiyohara Y, Iida M, Yamashita Y. Relationship of metabolic syndrome to periodontal disease in Japanese women: the Hisayama Study. J Dent Res 2007;86:271–275.
- 8. Nishimura F, Murayama Y. Periodontal inflammation and insulin resistance – lessons from obesity. J Dent Res 2001;80:1690–1694.
- 9. Mealey BL, Ocampo GL. Diabetes mellitus and periodontal disease. Periodontol 2000 2007;44:127–153.
- 10. Graves DT, Liu R, Oates TW. Diabetesenhanced inflammation and apoptosis: impact on periodontal pathosis. Periodontol 2000 2007;45:128–137.
- 11. Huttner EA, Machado DC, de Oliveira RB, Antunes AG, Hebling E. Effects of human aging on periodontal tissues. Spec Care Dentist 2009;29:149–155.
- 12. Arai K, Tanaka S, Yamamoto-Sawamura T, Sone K, Miyaishi O, Sumi Y. Aging changes in the periodontal bone of F344/N rat. Arch Gerontol Geriatr 2005;40:225–229.
- 13. Streckfus CF, Parsell DE, Streckfus JE, Pennington W, Johnson RB. Relationship between oral alveolar bone loss and aging among African-American and Caucasian individuals. Gerontology 1999;45:110–114.
- 14. Ritchie CS. Mechanistic links between type 2 diabetes and periodontitis. J Dent 2009;37:S578–S579.
- 15. Chapple IL. Reactive oxygen species and antioxidants in inflammatory diseases. J Clin Periodontol 1997;24:287–296.
- 16. Chapple IL. Role of free radicals and antioxidants in the pathogenesis of the inflammatory periodontal diseases. Clin Mol Pathol 1996;49:M247– M255.
- 17. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 2007;39:44–84.
- 18. Battino M, Bullon P, Wilson M, Newman H. Oxidative injury and inflammatory periodontal diseases: the challenge of anti-oxidants to free radicals and reactive oxygen species. Crit Rev Oral Biol Med 1999;10:458–476.
- 19. Halliwell B. Mechanisms involved in the generation of free radicals. Pathol Biol (Paris) 1996;44:6–13.
- 20. Sies H. Biochemistry of oxidative stress. Angew Chem Int Ed Engl 1986; 25: 1058– 1071.
- 21. Halliwell B, Gutteridge JM. Free radicals and antioxidant protection: mechanisms and significance in toxicology and disease. Hum Toxicol 1988;7:7–13.
- 22. Halliwell B, Gutteridge JM. Role of free radicals and catalytic metal ions in human disease: an overview. Methods Enzymol 1990;186:1–85.
- 23. Janssen-Heininger YM, Mossman BT, Heintz NH et al. Redox-based regulation of signal transduction: principles, pitfalls, and promises. Free Radic Biol Med 2008;45:1–17.
- 24. Zhang DX, Gutterman DD. Mitochondrial reactive oxygen species-mediated signaling in endothelial cells. Am J Physiol Heart Circ Physiol 2007;292: H2023–H2031.
- 25. Sedelnikova OA, Redon CE, Dickey JS, Nakamura AJ, Georgakilas AG, Bonner WM. Role of oxidatively induced DNA lesions in human pathogenesis. Mutat Res 2010;704:152–159.
- 26. Cataldi A, Di Giulio C. ''Oxygen supply'' as modulator of aging processes: hypoxia and hyperoxia models for aging studies. Curr Aging Sci 2009;2: 95–102.
- 27. Steinboeck F, Hubmann M, Bogusch A, Dorninger P, Lengheimer T, Heidenreich E. The relevance of oxidative stress and cytotoxic DNA lesions for spontaneous mutagenesis in non-replicating yeast cells. Mutat Res 2010;688:47–52.
- 28. Rossi P, Marzani B, Giardina S, Negro M, Marzatico F. Human skeletal muscle aging and the oxidative system: cellular events. Curr Aging Sci 2008;1:182–191.
- 29. Quarrie JK, Riabowol KT. Murine models of life span extension. Sci Aging Knowledge Environ 2004; 2004: re5.
- 30. Chapple IL, Brock GR, Milward MR, Ling N, Matthews JB. Compromised GCF total antioxidant capacity in periodontitis: cause or effect? J Clin Periodontol 2007;34:103–110.
- 31. Toker H, Ozdemir H, Eren K, Ozer H, Sahin G. N-acetylcysteine, a thiol antioxidant, decreases alveolar bone loss in experimental periodontitis in rats. J Periodontol 2009;80:672–678.
- 32. Matthews JB, Wright HJ, Roberts A, Ling-Mountford N, Cooper PR, Chapple IL. Neutrophil hyper-responsiveness in periodontitis. J Dent Res 2007;86:718– 722.
- 33. Sheikhi M, Bouhafs RK, Hammarstrom KJ, Jarstrand C. Lipid peroxidation caused by oxygen radicals from Fusobacterium-stimulated neutrophils as a

possible model for the emergence of periodontitis. Oral Dis 2001;7:41–46.

- 34. Sheikhi M, Gustafsson A, Jarstrand C. Cytokine, elastase and oxygen radical release by Fusobacterium nucleatumactivated leukocytes: a possible pathogenic factor in periodontitis. J Clin Periodontol 2000;27:758–762.
- 35. Keely S, Glover LE, MacManus CF et al. Selective induction of integrin beta1 by hypoxia-inducible factor: implications for wound healing. FASEB J 2009;23:1338–1346.
- 36. Rehman A, Nourooz-Zadeh J, Moller W, Tritschler H, Pereira P, Halliwell B. Increased oxidative damage to all DNA bases in patients with type II diabetes mellitus. FEBS Lett 1999;448:120–122.
- 37. Chapple IL, Brock G, Eftimiadi C, Matthews JB. Glutathione in gingival crevicular fluid and its relation to local antioxidant capacity in periodontal health and disease. Mol Pathol 2002;55:367–373.
- 38. Waddington RJ, Moseley R, Embery G. Reactive oxygen species: a potential role in the pathogenesis of periodontal diseases. Oral Dis 2000;6:138–151.
- 39. Chapple IL, Matthews JB. The role of reactive oxygen and antioxidant species in periodontal tissue destruction. Periodontol 2000 2007;43:160–232.
- 40. D'Aiuto F, Nibali L, Parkar M, Patel K, Suvan J, Donos N. Oxidative stress, systemic inflammation, and severe periodontitis. J Dent Res 2010;89:1241–1246.
- 41. Brock GR, Butterworth CJ, Matthews JB, Chapple IL. Local and systemic total antioxidant capacity in periodontitis and health. J Clin Periodontol 2004;31:515-521.
- 42. Tamaki N, Tomofuji T, Ekuni D, Yamanaka R, Morita M. Periodontal treatment decreases plasma oxidized LDL level and oxidative stress. Clin Oral Investig 2010. [Epub ahead of print].
- 43. Abou Sulaiman AE, Shehadeh RM. Assessment of total antioxidant capacity and the use of vitamin C in the treatment of non-smokers with chronic periodontitis. J Periodontol 2010;81:1547–1554.
- 44. Wei D, Zhang XL, Wang YZ, Yang CX, Chen G. Lipid peroxidation levels, total oxidant status and superoxide dismutase in serum, saliva and gingival crevicular fluid in chronic periodontitis patients before and after periodontal therapy. Aust Dent J 2010;55:70–78.
- 45. Grant MM, Brock GR, Matthews JB, Chapple IL. Crevicular fluid glutathione levels in periodontitis and the effect of non-surgical therapy. J Clin Periodontol 2010;37:17–23.
- 46. Linden GJ, McClean KM, Woodside JV et al. Antioxidants and periodontitis in 60–70-year-old men. J Clin Periodontol 2009;36:843–849.
- 47. Chapple IL, Milward MR, Dietrich T. The prevalence of inflammatory periodontitis is negatively associated with serum antioxidant concentrations. J Nutr 2007;137:657–664.
- 48. Patel SP, Pradeep AR, Chowdhry S. Crevicular fluid levels of plasma glutathione peroxidase (eGPx) in periodontal health and disease. Arch Oral Biol 2009;54:543–548.
- 49. Liskmann S, Vihalemm T, Salum O, Zilmer K, Fischer K, Zilmer M. Characterization of the antioxidant profile of human saliva in peri-implant health and disease. Clin Oral Implants Res 2007;18:27–33.
- 50. Manolagas SC. From estrogen-centric to aging and oxidative stress: a revised perspective of the pathogenesis of osteoporosis. Endocr Rev 2010;31:266–300.
- 51. Almeida M, Ambrogini E, Han L, Manolagas SC, Jilka RL. Increased lipid oxidation causes oxidative stress, increased peroxisome proliferatoractivated receptor-gamma expression, and diminished pro-osteogenic Wnt signaling in the skeleton. J Biol Chem 2009;284:27438–27448.
- 52. Almeida M, Han L, Martin-Millan M, O-Brien CA, Manolagas SC. Oxidative stress antagonizes Wnt signaling in osteoblast precursors by diverting beta-catenin from T cell factor- to forkhead box O-mediated transcription. J Biol Chem 2007;282:27298– 27305.
- 53. Manolagas SC, Almeida M. Gone with the Wnts: beta-catenin, T-cell factor, forkhead box O, and oxidative stress in age-dependent diseases of bone, lipid, and glucose metabolism. Mol Endocrinol 2007;21:2605–2614.
- 54. Jagger CJ, Lean JM, Davies JT, Chambers TJ. Tumor necrosis factor-alpha mediates osteopenia caused by depletion of antioxidants. Endocrinology 2005;146:113–118.
- 55. Lean JM, Davies JT, Fuller K et al. A crucial role for thiol antioxidants in estrogen-deficiency bone loss. J Clin Invest 2003;112:915–923.
- 56. Tomofuji T, Ekuni D, Irie K et al. Preventive effects of a cocoa-enriched diet on gingival oxidative stress in experimental periodontitis. J Periodontol 2009;80:1799–1808.
- 57. Halliwell B. Antioxidant defence mechanisms: from the beginning to the end (of the beginning). Free Radic Res 1999;31:261–272.
- 58. Barthel A, Schmoll D, Unterman TG. FoxO proteins in insulin action and metabolism. Trends Endocrinol Metab 2005;16:183–189.
- 59. Kaestner KH, Knochel W, Martinez DE. Unified nomenclature for the winged

helix/forkhead transcription factors. Genes Dev 2000;14:142–146.

- 60. Greer EL, Brunet A. FOXO transcription factors at the interface between longevity and tumor suppression. Oncogene 2005;24:7410–7425.
- 61. Brunet A, Bonni A, Zigmond MJ et al. Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. Cell 1999;96:857– 868.
- 62. Huang H, Tindall DJ. Dynamic FoxO transcription factors. J Cell Sci 2007;120:2479–2487.
- 63. Biggs WH 3rd, Meisenhelder J, Hunter T, Cavenee WK, Arden KC. Protein kinase B/Akt-mediated phosphorylation promotes nuclear exclusion of the winged helix transcription factor FKHR1. Proc Natl Acad Sci USA 1999;96:7421–7426.
- 64. Furuyama T, Nakazawa T, Nakano I, Mori N. Identification of the differential distribution patterns of mRNAs and consensus binding sequences for mouse DAF-16 homologues. Biochem J 2000;349:629–634.
- 65. Gilley J, Coffer PJ, Ham J. FOXO transcription factors directly activate bim gene expression and promote apoptosis in sympathetic neurons. J Cell Biol 2003;162:613–622.
- 66. Dijkers PF, Medema RH, Lammers JW, Koenderman L, Coffer PJ. Expression of the pro-apoptotic Bcl-2 family member Bim is regulated by the forkhead transcription factor FKHR-L1. Curr Biol 2000;10:1201–1204.
- 67. Nakamura T, Sakamoto K. Forkhead transcription factor FOXO subfamily is essential for reactive oxygen species-induced apoptosis. Mol Cell Endocrinol 2008;281:47–55.
- 68. Kops GJ, Dansen TB, Polderman PE et al. Forkhead transcription factor FOXO3a protects quiescent cells from oxidative stress. Nature 2002;419:316– 321.
- 69. Martinez-Gac L, Marques M, Garcia Z, Campanero MR, Carrera AC. Control of cyclin G2 mRNA expression by forkhead transcription factors: novel mechanism for cell cycle control by phosphoinositide 3-kinase and forkhead. Mol Cell Biol 2004;24:2181–2189.
- 70. Seoane J, Le HV, Shen L, Anderson SA, Massague J. Integration of Smad and forkhead pathways in the control of neuroepithelial and glioblastoma cell proliferation. Cell 2004;117:211–223.
- 71. Nakamura N, Ramaswamy S, Vazquez F, Signoretti S, Loda M, Sellers WR. Forkhead transcription factors are critical effectors of cell death and cell cycle arrest downstream of PTEN. Mol Cell Biol 2000;20:8969–8982.
- 72. Kops GJ, Medema RH, Glassford J et al. Control of cell cycle exit and entry by protein kinase B-regulated forkhead transcription factors. Mol Cell Biol 2002;22:2025–2036.
- 73. Ramaswamy S, Nakamura N, Sansal I, Bergeron L, Sellers WR. A novel mechanism of gene regulation and tumor suppression by the transcription factor FKHR. Cancer Cell 2002;2:81–91.
- 74. Schmidt M, Fernandez de Mattos S, van der Horst A et al. Cell cycle inhibition by FoxO forkhead transcription factors involves downregulation of cyclin D. Mol Cell Biol 2002;22:7842–7852.
- 75. Modur V, Nagarajan R, Evers BM, Milbrandt J. FOXO proteins regulate tumor necrosis factor-related apoptosis inducing ligand expression. Implications for PTEN mutation in prostate cancer. J Biol Chem 2002;277:47928–47937.
- 76. Nemoto S, Finkel T. Redox regulation of forkhead proteins through a p66shcdependent signaling pathway. Science 2002;295:2450–2452.
- 77. Tran H, Brunet A, Grenier JM et al. DNA repair pathway stimulated by the forkhead transcription factor FOXO3a through the Gadd45 protein. Science 2002;296:530–534.
- 78. Tothova Z, Kollipara R, Huntly BJ et al. FoxOs are critical mediators of hematopoietic stem cell resistance to physiologic oxidative stress. Cell 2007;128:325–339.
- 79. Vogt PK, Jiang H, Aoki M. Triple layer control: phosphorylation, acetylation and ubiquitination of FOXO proteins. Cell Cycle 2005;4:908–913.
- 80. Yang Y, Hou H, Haller EM, Nicosia SV, Bai W. Suppression of FOXO1 activity by FHL2 through SIRT1-mediated deacetylation. EMBO J 2005;24:1021– 1032.
- 81. Brunet A, Sweeney LB, Sturgill JF et al. Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. Science 2004;303:2011–2015.
- 82. Lam EW, Francis RE, Petkovic M. FOXO transcription factors: key regulators of cell fate. Biochem Soc Trans 2006;34:722–726.
- 83. Furuyama T, Kitayama K, Shimoda Y et al. Abnormal angiogenesis in Foxo1 (Fkhr)-deficient mice. J Biol Chem 2004;279:34741–34749.
- 84. Hosaka T, Biggs WH 3rd, Tieu D et al. Disruption of forkhead transcription factor (FOXO) family members in mice reveals their functional diversification. Proc Natl Acad Sci USA 2004;101:2975– 2980.
- 85. Castrillon DH, Miao L, Kollipara R, Horner JW, DePinho RA. Suppression of ovarian follicle activation in mice by the transcription factor Foxo3a. Science 2003;301:215–218.
- 86. Lin L, Hron JD, Peng SL. Regulation of NF-kappaB, Th activation, and autoinflammation by the forkhead transcription factor Foxo3a. Immunity 2004;21: 203–213.
- 87. Rached MT, Kode A, Xu L et al. FoxO1 is a positive regulator of bone formation by favoring protein synthesis and resistance to oxidative stress in osteoblasts. Cell Metab 2010;11:147–160.
- 88. Ambrogini E, Almeida M, Martin-Millan M et al. FoxO-mediated defense against oxidative stress in osteoblasts is indispensable for skeletal homeostasis in mice. Cell Metab 2010;11:136–146.
- 89. Kandel ES, Hay N. The regulation and activities of the multifunctional serine/ threonine kinase Akt/PKB. Exp Cell Res 1999;253:210–229.
- 90. Lawlor MA, Alessi DR. PKB/Akt: a key mediator of cell proliferation, survival and insulin responses? J Cell Sci 2001;114:2903–2910.
- 91. Cantley LC. The phosphoinositide 3-kinase pathway. Science 2002;296: 1655–1657.
- 92. Jiang BH, Liu LZ. PI3K/PTEN signaling in tumorigenesis and angiogenesis. Biochim Biophys Acta 2008;1784:150– 158.
- 93. Nogueira V, Park Y, Chen CC et al. Akt determines replicative senescence and oxidative or oncogenic premature senescence and sensitizes cells to oxidative apoptosis. Cancer Cell 2008;14:458– 470.
- 94. Korkaya H, Paulson A, Charafe-Jauffret E et al. Regulation of mammary stem/ progenitor cells by PTEN/Akt/ beta-catenin signaling. PLoS Biol 2009;7:e1000121.
- 95. Kandel ES, Skeen J, Majewski N et al. Activation of Akt/protein kinase B overcomes a G(2)/m cell cycle checkpoint induced by DNA damage. Mol Cell Biol 2002;22:7831–7841.
- 96. Nakae J, Biggs WH 3rd, Kitamura T et al. Regulation of insulin action and pancreatic beta-cell function by mutated alleles of the gene encoding forkhead transcription factor Foxo1. Nat Genet 2002;32:245–253.
- 97. Kushner JA, Simpson L, Wartschow LM et al. Phosphatase and tensin homolog regulation of islet growth and glucose homeostasis. J Biol Chem 2005;280:39388–39393.
- 98. Kaneto H, Kajimoto Y, Miyagawa J et al. Beneficial effects of antioxidants in diabetes: possible protection of pancreatic beta-cells against glucose toxicity. Diabetes 1999;48:2398–2406.
- 99. Tanaka Y, Gleason CE, Tran PO, Harmon JS, Robertson RP. Prevention of glucose toxicity in HIT-T15 cells and Zucker diabetic fatty rats by antioxi-

dants. Proc Natl Acad Sci USA 1999;96:10857–10862.

- 100. Yang H, Roberts LJ, Shi MJ et al. Retardation of atherosclerosis by overexpression of catalase or both Cu/Znsuperoxide dismutase and catalase in mice lacking apolipoprotein E. Circ Res 2004;95:1075–1081.
- 101. Menini S, Amadio L, Oddi G et al. Deletion of p66Shc longevity gene protects against experimental diabetic glomerulopathy by preventing diabetesinduced oxidative stress. Diabetes 2006;55:1642–1650.
- 102. Kamei Y, Miura S, Suzuki M et al. Skeletal muscle FOXO1 (FKHR) transgenic mice have less skeletal muscle mass, down-regulated Type I (slow twitch/red muscle) fiber genes, and impaired glycemic control. J Biol Chem 2004;279: 41114–41123.
- 103. Yang-Snyder J, Miller JR, Brown JD, Lai CJ, Moon RT. A frizzled homolog functions in a vertebrate Wnt signaling pathway. Curr Biol 1996;6:1302–1306.
- 104. Moon RT, Brown JD, Yang-Snyder JA, Miller JR. Structurally related receptors and antagonists compete for secreted Wnt ligands. Cell 1997;88:725–728.
- 105. van Amerongen R, Mikels A, Nusse R. Alternative wnt signaling is initiated by distinct receptors. Sci Signal 2008;1:re9.
- 106. Hartmann C. A Wnt canon orchestrating osteoblastogenesis. Trends Cell Biol 2006;16:151–158.
- 107. Mikels AJ, Nusse R. Purified Wnt5a protein activates or inhibits beta-catenin-TCF signaling depending on receptor context. PLoS Biol 2006;4:e115.
- 108. Gordon MD, Nusse R. Wnt signaling: multiple pathways, multiple receptors, and multiple transcription factors. J Biol Chem 2006;281:22429–22433.
- 109. Angers S, Moon RT. Proximal events in Wnt signal transduction. Nat Rev Mol Cell Biol 2009;10:468–477.
- 110. Clevers H. Wnt/beta-catenin signaling in development and disease. Cell 2006;127: 469–480.
- 111. Verheyen EM, Gottardi CJ. Regulation of Wnt/beta-catenin signaling by protein kinases. Dev Dyn 2009;239:34–44.
- 112. Mosimann C, Hausmann G, Basler K. Beta-catenin hits chromatin: regulation of Wnt target gene activation. Nat Rev Mol Cell Biol 2009;10:276–286.
- 113. Tutter AV, Fryer CJ, Jones KA. Chromatin-specific regulation of LEF-1-betacatenin transcription activation and inhibition in vitro. Genes Dev 2001;15: 3342–3354.
- 114. Huelsken J, Vogel R, Brinkmann V, Erdmann B, Birchmeier C, Birchmeier W. Requirement for beta-catenin in anterior-posterior axis formation in mice. J Cell Biol 2000;148:567-578.
- 115. Martin BL, Kimelman D. Wnt signaling and the evolution of embryonic posterior development. Curr Biol 2009;19:R215– R219.
- 116. Nusse R. Wnt signaling and stem cell control. Cell Res 2008;18:523–527.
- 117. Rodda SJ, McMahon AP. Distinct roles for Hedgehog and canonical Wnt signaling in specification, differentiation and maintenance of osteoblast progenitors. Development 2006;133:3231–3244.
- 118. Krishnan V, Bryant HU, Macdougald OA. Regulation of bone mass by Wnt signaling. *J Clin Invest* 2006;116:1202-1209.
- 119. Balemans W, Ebeling M, Patel N et al. Increased bone density in sclerosteosis is due to the deficiency of a novel secreted protein (SOST). Hum Mol Genet 2001;10:537–543.
- 120. Brunkow ME, Gardner JC, Van Ness J et al. Bone dysplasia sclerosteosis results from loss of the SOST gene product, a novel cystine knot-containing protein. Am J Hum Genet 2001;68:577–589.
- 121. van Bezooijen RL, Bronckers AL, Gortzak RA et al. Sclerostin in mineralized matrices and van Buchem disease. J Dent Res 2009;88:569–574.
- 122. Wan M, Yang C, Li J et al. Parathyroid hormone signaling through low-density lipoprotein-related protein 6. Genes Dev 2008;22:2968–2979.
- 123. Glass DA 2nd, Bialek P, Ahn JD et al. Canonical Wnt signaling in differentiated osteoblasts controls osteoclast differentiation. Dev Cell 2005;8:751–764.
- 124. Allard D, Figg N, Bennett MR, Littlewood TD. Akt regulates the survival of vascular smooth muscle cells via inhibition of FoxO3a and GSK3. J Biol Chem 2008;283:19739–19747.
- 125. Rokutanda S, Fujita T, Kanatani N et al. Akt regulates skeletal development through GSK3, mTOR, and FoxOs. Dev Biol 2009;328:78–93.
- 126. Hoogeboom D, Burgering BM. Should I stay or should I go: beta-catenin decides under stress. Biochim Biophys Acta 2009;1796:63–74.
- 127. Hoogeboom D, Essers MA, Polderman PE, Voets E, Smits LM, Burgering BM. Interaction of FOXO with beta-catenin inhibits beta-catenin/T cell factor activity. J Biol Chem 2008;283:9224– 9230.
- 128. Essers MA, Weijzen S, de Vries-Smits AM et al. FOXO transcription factor activation by oxidative stress mediated by the small GTPase Ral and JNK. EMBO J 2004;23:4802–4812.
- 129. Ishitani T, Ninomiya-Tsuji J, Nagai S et al. The TAK1-NLK-MAPK-related pathway antagonizes signalling between beta-catenin and transcription factor TCF. Nature 1999;399:798–802.
- 130. Lu H, Kraut D, Gerstenfeld LC, Graves DT. Diabetes interferes with the bone formation by affecting the expression of transcription factors that regulate osteoblast differentiation. Endocrinology 2003;144:346–352.
- 131. Fujii H, Hamada Y, Fukagawa M. Bone formation in spontaneously diabetic Torii-newly established model of non-obese type 2 diabetes rats. Bone 2008;42:372– 379.
- 132. Hamada Y, Kitazawa S, Kitazawa R, Fujii H, Kasuga M, Fukagawa M. Histomorphometric analysis of diabetic osteopenia in streptozotocin-induced diabetic mice: a possible role of oxidative stress. Bone 2007;40:1408–1414.
- 133. Kuhn H, Borchert A. Regulation of enzymatic lipid peroxidation: the interplay of peroxidizing and peroxide reducing enzymes. Free Radic Biol Med 2002;33:154–172.
- 134. Lee JY, Jung GY, Heo HJ et al. 4-Hydroxynonenal induces vascular smooth muscle cell apoptosis through mitochondrial generation of reactive oxygen species. Toxicol Lett 2006;166:212–221.
- 135. Klein RF, Allard J, Avnur Z et al. Regulation of bone mass in mice by the lipoxygenase gene Alox15. Science 2004;303:229–232.
- 136. Akune T, Ohba S, Kamekura S et al. PPARgamma insufficiency enhances osteogenesis through osteoblast formation from bone marrow progenitors. J Clin Invest 2004;113:846–855.
- 137. Lecka-Czernik B, Moerman EJ, Grant DF, Lehmann JM, Manolagas SC, Jilka RL. Divergent effects of selective peroxisome proliferator-activated receptorgamma 2 ligands on adipocyte versus osteoblast differentiation. Endocrinology 2002;143:2376–2384.
- 138. Soroceanu MA, Miao D, Bai XY, Su H, Goltzman D, Karaplis AC. Rosiglitazone impacts negatively on bone by promoting osteoblast/osteocyte apoptosis. J Endocrinol 2004;183:203–216.
- 139. Ali AA, Weinstein RS, Stewart SA, Parfitt AM, Manolagas SC, Jilka RL. Rosiglitazone causes bone loss in mice by suppressing osteoblast differentiation and bone formation. Endocrinology 2005;146:1226–1235.
- 140. Garrett IR, Boyce BF, Oreffo RO, Bonewald L, Poser J, Mundy GR. Oxygen-derived free radicals stimulate osteoclastic bone resorption in rodent bone in vitro and in vivo. J Clin Invest 1990;85:632–639.
- 141. Bai XC, Lu D, Liu AL et al. Reactive oxygen species stimulates receptor activator of NF-kappaB ligand expression in osteoblast. J Biol Chem 2005;280:17497– 17506.
- 142. Lean JM, Jagger CJ, Kirstein B, Fuller K, Chambers TJ. Hydrogen peroxide is essential for estrogen-deficiency bone loss and osteoclast formation. Endocrinology 2005;146:728–735.
- 143. Lee NK, Choi YG, Baik JY et al. A crucial role for reactive oxygen species in RANKL-induced osteoclast differentiation. Blood 2005;106:852–859.
- 144. Kim MS, Yang YM, Son A et al. RANKL-mediated reactive oxygen species pathway that induces long lasting Ca2+ oscillations essential for osteoclastogenesis. J Biol Chem 2010;285:6913– 6921.
- 145. Srinivasan S, Avadhani NG. Hypoxiamediated mitochondrial stress in RAW264.7 cells induces osteoclast-like TRAP-positive cells. Ann N Y Acad Sci 2007;1117:51–61.
- 146. Koh JM, Lee YS, Kim YS et al. Homocysteine enhances bone resorption by stimulation of osteoclast formation and activity through increased intracellular

ROS generation. J Bone Miner Res 2006;21:1003–1011.

- 147. AlmeidaM,Martin-MillanM, Ambrogini E et al. Estrogens attenuate oxidative stress and thedifferentiation and apoptosis of osteoblasts by dna binding-independent actions of the ER alpha. J Bone Miner Res 2010;25:769–781.
- 148. Almeida M, Han L, Ambrogini E, Bartell SM, Manolagas SC. Oxidative stress stimulates apoptosis and activates NF-kappaB in osteoblastic cells via a PKCbeta/p66shc signaling cascade: counter regulation by estrogens or androgens. Mol Endocrinol 2010;24: 2030–2037.
- 149. Jilka RL, Almeida M, Ambrogini E et al. Decreased oxidative stress and greater bone anabolism in the aged, when compared to the young, murine skeleton with parathyroid hormone administration. Aging Cell 2010;9:851–867.
- 150. Maruyama T, Tomofuji T, Endo Y et al. Supplementation of green tea catechins

in dentifrices suppresses gingival oxidative stress and periodontal inflammation. Arch Oral Biol 2011;56:48–53.

- 151. Govindaraj J, Emmadi P, Deepalakshmi, Rajaram V, Prakash G, Puvanakrishnan R. Protective effect of proanthocyanidins on endotoxin induced experimental periodontitis in rats. Indian J Exp Biol 2010;48:133–142.
- 152. Tomofuji T, Ekuni D, Sanbe T et al. Effects of vitamin C intake on gingival oxidative stress in rat periodontitis. Free Radic Biol Med 2009;46:163–168.
- 153. Khmelevskii IuV, Danilevskii NF, Borisenko AV, Poberezkina NV. [Effect of vitamins A, E and K on the indices of the glutathione antiperoxide system in gingival tissues in periodontosis]. Vopr Pitan 1985;4:54–56.
- 154. Hirasawa M, Takada K, Makimura M, Otake S. Improvement of periodontal status by green tea catechin using a local delivery system: a clinical pilot study. J Periodontal Res 2002;37:433–438.

This document is a scanned copy of a printed document. No warranty is given about the accuracy of the copy. Users should refer to the original published version of the material.