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Evaluation of three selective media for isolation of *Aggregatibacter actinomycetemcomitans*

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Background and Objective: Aggregatibacter actinomycetemcomitans is a pathogen in oral and nonoral infections. Detection and quantification of this pathogen can be performed using selective culture techniques. The aim of this study was to establish the efficacy of two known selective media in their ability to select and support the growth of *A. actinomycetemcomitans*.

Material and Methods: Trypticase soy bacitracin vancomycin (TSBV) medium and brain-heart infusion agar with vancomycin (Dentaid-1), as well as a modified Dentaid-1 medium (in which the brain-heart infusion agar was substituted with brain-heart infusion broth), were compared. Two-hundred and eighteen clinical samples were used to establish the recovery rate, the number of colonyforming units (CFUs) of *A. actinomycetemcomitans* as well as the total number of CFUs on the three different types of medium. In addition, the numbers of gram-negative aerobic rods and yeasts were determined.

Results: Both types of Dentaid-1 medium showed a higher recovery of *A. actinomycetemcomitans* compared with TSBV. However, these differences did not reach statistical significance. The total number of CFUs of *A. actinomycetemcomitans* recovered was significantly higher on Dentaid-1 compared with TSBV (p = 0.029). The mean number of gram-negative aerobic rods recovered was statistically higher on both types of Dentaid-1 medium in comparison with TSBV. Low numbers of yeasts were recovered occasionally on all test plates.

Conclusion: Dentaid-1 is a low-cost effective alternative to TSBV for the isolation and growth of *A. actinomycetemcomitans* from clinical samples, such as dental plaque, which contain a complex microflora.

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Aggregatibacter actinomycetemcomitans is a capnophilic, gram-negative rod that is primarily associated with human periodontal disease. The organism can also be involved in nonoral infections such as endocarditis, prosthetic heart valve infection (1) and brain abscesses (2). A. actinomycetem*comitans* has a low prevalence in periodontal health (3,4). However, high levels of the organism are found in aggressive periodontitis (5). Detection of *A. actinomycetemcomitans* has been facilitated by the use of selective isolation media. Mandel & Socransky (6) described on a trypticase soy agar based-medium supplemented with sheep blood, malachite green and bacitracin for the isolation if *A. actinomycetemcomitans*. Slots (7) reported on a tryptic soy agar elective medium supplemented with horse serum, yeast, bacitracin and vancomycin (TSBV), and this medium has become the standard medium for the isolation and growth of A. actinomycetemcomitans. The TSBV medium was modified by Holm et al. (8), who added carbenicillin, fusidic acid and spiramycin to give better suppression of bacterial species other than A. actinomycetemcomitans. This medium is, however, complex and expensive to prepare. In order to produce a low-cost, but effective, isolation medium for A. actinomycetemcomitans, Alsina et al. (9) developed a new medium, Dentaid-1, which has brainheart infusion agar as a base and is supplemented with yeast, sodium fumarate, sodium formate and vancomycin. This new medium was shown to support the growth of A. actinomycetemcomitans and the number of contaminants was comparable with the TSBV medium. Although detection of A. actinomycetemcomitans is currently often performed using molecular techniques, such as the PCR, isolation of A. actinomycetemcomitans by culture is still necessary for antibiotic-susceptibility testing (10) and strain typing (11). The purpose of the present study was to evaluate the recovery of A. actinomycetemcomitans from a large sample of clinical specimens (n = 218)after culture on Dentaid-1 medium prepared using two different brainheart infusion bases and to compare the recovery of A. actinomycetemcomitans on these media with recovery on the TSBV medium. Attention was also paid to the concomitant growth of yeasts and gram-negative aerobic rods on the three selective media.

Material and methods

Selective media

Dentaid-1 and modified Dentaid-1 (mDentaid-1) were prepared with brain-heart infusion agar and brainheart infusion broth, respectively, and were supplemented with the compounds listed in Table 1. TSBV medium was prepared according to the original description by Slots (7) and checked for the ability to support the growth of *A. actinomycetemcomitans* using a reference strain (DSM 11123). Plates were used within 2 d of preparation. *Table 1.* Composition of Dentaid-1, modified Dentaid-1 (mDentaid-1) and trypticase soy bacitracin vancomycin (TSBV) isolation media

TSBV	Dentaid-1	mDentaid-1
Tryptic soy agar: 40 g/L	BHI agar: 52 g/L	BHI broth: 37 g/L
Yeast extract: 1.0 g/L	Yeast extract: 5 g/L	Agar: 15 g/L
Autoclave: 121°C for 15 min	Sodium fumarate: 1.5 g/L	Yeast extract: 5 g/L
рН 7.2	Sodium formate: 1 g/L	Sodium fumarate: 1.5 g/L
Horse serum: 10%	pH 7.2	Sodium formate: 1 g/L
Bacitracin: 75 µg/mL	Autoclave: 121°C for	рН 7.2
Vancomycin: 5 µg/mL	15 min	Autoclave: 121°C for
	Vancomycin: 9 µg/mL	15 min
		Vancomycin: 9 µg/mL

BHI, brain-heart infusion.

Design

The study was performed in two separate experiments: in the first experiment, the recovery of A. actinomycetemcomitans on TSBV and Dentaid-1 media was compared; and in the second experiment, mDentaid-1 medium was compared with TSBV medium for the recovery of A. actinomycetemcomitans. The outcome variables of these experiments included: total numbers of colony-forming units (CFUs); total counts of A. actinomycetemcomitans; total counts of gramnegative aerobic rods; and total counts of yeasts.

Clinical samples

Subgingival samples (n = 218) were obtained from patients with untreated, chronic or aggressive destructive periodontal disease with deep and bleeding pockets (> 5 mm). Samples were obtained after mechanical removal of supragingival plaque deposits and air drying of the sampling sites. Sampling sites were than isolated with cotton rolls. Two sterile paper points were subsequently inserted into the deepest and bleeding pocket in each quadrant of the dentition and left *in situ* for 15 s. Paper points were pooled in 2 mL of reduced transport fluid (12).

Sample processing

Upon arrival in the laboratory, the samples were vortexed for 45 s to dislodge bacteria from the paper points. Tenfold serial dilutions were prepared in sterile saline (1/10, 1/100 and 1/1000 dilutions) and $100-\mu\text{L}$ aliquots of the dilutions were spread on the three selective isolation media. The plates were incubated in air + 5% CO₂ at 35°C for up to 5 d.

Identification and counting

A. actinomycetemcomitans was identified on the basis of its typical colony morphology (a star-like inner structure) and the ability to degrade 3% hydrogen peroxide by catalase production (7). For further identification, the API-ZYM (Biomérieux, Marcy l'Etoile, France) was used (13).

The total number of CFUs was determined on plates containing at least 100 CFUs. Then, the numbers of CFUs of yeast species and of gram-negative rods were established using gram staining. The total number of CFUs of gram-negative rods was recorded using the following semiquantitative scale: 0, no gram-negative rods; 1, < 100 CFUs; 2, 10–300 CFUs; and 3, > 300 CFUs.

Data analyses

The numbers of total CFUs, total counts of *A. actinomycetemcomitans* and total counts of gram-negative rods were log-transformed (Graph-Pad Instat 3; GraphPad Software, Inc., La Jolla, CA, USA). Differences in mean counts were tested for statistical significance using the nonparametric Mann–Whitney *U*-test. Detection frequencies on the different plates were compared using Fisher's

exact test. The level of significance was set at 5% (p < 0.05).

Results

A. actinomycetemcomitans was isolated more frequently from Dentaid-1 (24%) than from TSBV medium (19%; p = not significant). Also, in the second experiment the number of actinomycetemcomitans-positive A samples was higher on mDentaid-1 (18%) than on TSBV medium (16%; p = not significant (Table 2). Comparison of A. actinomycetemcomitans counts revealed a significantly higher mean count on Dentaid-1 in comparison with TSBV medium (p = 0.029, Table 3). The mean total A. actinomycetemcomitans count on the mDentaid-1 medium was also higher than on TSBV medium but the difference did not reach significance. Significantly higher counts of gram-negative aerobic rods were observed on both Dentaid-1 (p = 0.0142) and mDentaid-1 (p = 0.0001) media in comparison with TSBV medium. No significant differences were noted in the total numbers of CFUs between the two types of Dentaid-1 medium and TSBV medium. The number of samples with detectable yeast was low on Dentaid-1 and TSBV medium (12/111 vs. 7/111; p = not significant) and on mDentaid-1 and TSBV medium (15/107 vs. 14/107; p = not significant).

Discussion

The aim of this study was to compare the efficacy of three selective media for the isolation and growth of *A. actinomycetemcomitans*. The high costs of the TSBV medium were a major reason for this comparative study. The TSBV medium contains 10% horse serum and two antibiotics, whereas Dentaid-1 needs no serum for growth of *A. actinomycetemcomitans*. To further reduce the costs of Dentaid-1 we replaced the brain-heart infusion agar with brain-heart infusion broth. *A. actinomycetemcomitans*

Table 2. Frequency of isolation of Aggregatibacter actinomycetemcomitans on Dentaid-1, modified Dentaid-1 (mDentaid-1) and trypticase soy bacitracin vancomycin (TSBV) isolation media

Experiment 1	No. of samples	Experiment 2	No. of samples
Dentaid-1 (+), TSBV (+)	22	mDentaid-1 (+), TSBV (+)	14
Dentaid-1 (+), TSBV (-)	5	mDentaid-1 (+), TSBV (-)	5
Dentaid-1 (-), TSBV (+)	0	mDentaid-1 (-), TSBV (+)	3
Dentaid-1 (-), TSBV (-)	84	mDentaid-1 (-), TSBV (-)	85
Total samples	111		107

(+), *A. actinomycetemcomitans* recovered. (-), *A. actinomycetemcomitans* not recovered. Two different sets of TSBV plates were used in the experiments.

Table 3. Total counts of *Aggregatibacter actinomycetemcomitans*, gram-negative aerobic rods and total colony-forming units on Dentaid-1, modified Dentaid-1 (mDentaid-1) and trypticase soy bacitracin vancomycin (TSBV) isolation media

Bacterial species	Experiment 1		Experiment 2				
	Dentaid-1	TSBV	mDentaid-1	TSBV			
Total <i>A. actinomycetemcomitans</i> counts	4.73(0.62) ^a	4.54(1.12)	4.92(1.14)	4.84(0.65)			
Gram-negative aerobic rods	1.3(1.1) ^b	0.96(1.1)	1.8(1.0) °	1.4(1.2)			
Total counts	4.25(1.19)	4.48(0.92)	5.07(0.85)	5.08(0.89)			

Values are mean (standard deviation) of log-transformed total counts.

^aSignificant difference between Dentaid-1 and TSBV in experiment 1 (p = 0.029). ^bSignificant difference between Dentaid-1 and TSBV in experiment 1 (p = 0.0142). ^cSignificant difference between mDentaid-1 and TSBV in experiment 2 (p = 0.0001).

was recovered more frequently on both types of Dentaid-1 medium than on TSBV medium. The reason why A. actinomycetemcomitans may not be isolated on TSBV medium in some cases is unclear. A. actinomycetemcomitans isolates from samples recovered from Dentaid-1 plates, but not from TSBV plates, were transferred to freshly prepared TSBV medium and in all instances these isolates produced growth on TSBV. Therefore, it is unlikely that suppression of the growth of feeder bacteria on TSBV medium is the explanation for the lack of initial growth of A. actinomycetemcomitans in some samples. We also checked A. actinomycetemcomitans strains isolated from Dentaid-1 media, but not from TSBV medium, for susceptibility to bacitracin and vancomycin; however, none of these isolates were susceptible to these antibiotics at the concentrations used in the isolation media. Reduced growth (i.e. 2-5 logs of inhibition) of A. actinomycetemcomitans on TSBV medium, but not on blood agar, has been described (9).

Compared with TSBV medium, a significantly higher number of *A. ac-tinomycetemcomitans* colonies were isolated from Dentaid-1 but not from mDentaid-1. One potentially negative characteristic was the recovery of a significantly higher number of gramnegative aerobic rods from both Dentaid-1 and mDentaid-1. Although this phenomenon is a disadvantage, it did not result in a lower recovery rate of *A. actinomycetemcomitans* and did not reduce the total numbers of *A. actinomycetemcomitans*.

We did not include the malachitegreen bacitracin selective medium in this study because it has been shown that the recovery rate of A. actinomycetemcomitans on this medium is much lower than on TSBV medium (14). We incubated all plates in air + 5% CO₂ because it has been shown that anaerobic incubation significantly reduces the recovery of A. actinomycetemcomitans on all selective media tested (14). Our results are in full agreement with the observations of Alsina et al. (9), who first described the low-cost Dentaid-1 culture medium and who also found a significantly lower recovery (79%) of *A. actinomycetemcomitans* on TSBV medium.

We confirm that Dentaid-1 medium is highly selective and cost effective (approximately 50% of the cost of TSBV medium) and may therefore be a useful tool for the isolation and growth of *A. actinomycetemcomitans* from both oral and nonoral clinical samples.

References

- Van Winkelhoff AJ, Slots J. Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in nonoral infections. Periodontol 2000 1999;20:122–135.
- Rahamat Langendoen JC, Van Vonderen MG, Engstrom LJ, Manson WL, Van Winkelhoff AJ, Mooi-Kokenberg EA. Brain abscess associated with *Aggregatibacter actinomycetemcomitans*: case report and review of literature. *J Clin Periodont* 2011;38:702–706.
- Boutaga K, Van Winkelhoff AJ, Vandenbroucke-Grauls CMJE, Savelkoul

PHM. The additional value of real-time PCR in the quantitative detection of periodontal pathogens. *J Clin Periodont* 2006;**33**:427–433.

- Slots J. Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in periodontal disease: introduction. Periodontol 2000 1999;20:7–13.
- Rodenburg JP, Van Winkelhoff AJ, Winkel EG, Goené RJ, Abbas F, De Graaff J. Occurence of *Bacteroides gingivalis*, *Bacteroides intermedius and Actinobacillus actinomycetemcomitans* in severe periodontitis in relation to age and treatment history. *J Clin Periodontol* 1990;17: 392–399.
- Mandell RL, Socransky SS. A selective medium for *Actinobacillus actinomycetemcomitans* and the incidence of the organism in juvenile periodontitis. *J Periodontol* 1981;52:593–598.
- Slots J. Selective medium for isolation of Actinobacillus actinomycetemcomitans. J Clin Microbiol 1982;15:606–609.
- Holm A, Rabe P, Kalfas S, Edwardsson S. Improved selective culture media for Actinobacillus actinomycetemcomitans and Haemophilus aphrophilus. J Clin Microbiol 1987;25:1985–1988.

- Alsina M, Olle E, Frias J. Improved, low-cost selective culture medium for Actinobacillus actinomycetemcomitans. J Clin Microbiol 2001;39:509–513.
- Ardila CM, Granada MI, Guzman IC. Antibiotic resistance of subgingival species in chronic periodontitis patients. J Periodontal Res 2010;45:557–563.
- Claesson R, Lagervall M, Höglund-Aberg C, Johansson A, Haubek D. Detection of the highly leucotoxic JP2 clone of Aggregatibacter actinomycetemcomitans in members of a Caucasian family living in Sweden. J Clin Periodontol 2011;38:115–121.
- Syed SA, Loesche WJ. Survival of human dental plaque flora in various transport media. *Appl Microbiol* 1972;24:638–644.
- Slots J. Enzymatic characterization of some oral and nonoral gram-negative bacteria with the API ZYM system. J Clin Microbiol 1981;14:288–294.
- Martijn van Steenbergen TJM, Van Winkelhoff AJ, Van der Mispel L, Van der Velden U, Abbas F, de Graaff J. Comparison of two selective media for *Actinobacillus actinomycetemcomitans*. J Clin Microbiol 1986;24:636–638.

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