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INVITED REVIEW

Sjögren's syndrome: why autoimmune epithelitis?

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The expression 'autoimmune epithelitis' has been proposed as an alternative for Sjögren's syndrome (SS) based on data pointing out the central role of the epithelial cell in the pathogenesis of the syndrome. Clinically, apart from exocrine glands that are the main target, the epithelial component of the other organs such as kidneys, liver, lungs or thyroid is commonly affected resulting in various extraglandular manifestations. On the other hand, at the molecular and cellular level, the epithelial cell plays a major role in the initiation and perpetuation of the autoimmune lesion. Mechanisms such as antigen presentation, apoptosis, chemokine production or germinal center formation lie in the center of SS pathogenesis and the epithelial cell has a very important role. Herein, we present both aspects, review the data that support the proposed terminology and finally, suggest a unifying theory for the pathogenesis of SS. Oral Diseases (2006) 12, 523-532

Keywords: Sjögren's syndrome; autoimmune epithelitis; extraglandular manifestations; epithelial cell; antigen-presenting cell; apoptosis

Introduction

Autoimmune attack of epithelial tissues is the main characteristic of Sjögren's syndrome (SS). Ocular and oral dryness are the main presenting symptoms as the lymphocytic invasion of salivary and lacrimal glands results in their functional impairment. SS is a relatively common chronic autoimmune disease affecting approximately 1–2% of total population and may occur either alone (primary SS) or along with other autoimmune diseases (mainly systemic lupus erythematosus or rheumatoid arthritis, secondary SS). Its progress is benign unless certain adverse prognostic factors are present, such as low C4 complement levels, mixed monoclonal cryoglobulinemia or palpable purpura. Patients with these factors are at high risk for B-cell (mucosaassociated lymphoid tissue; MALT) lymphoma and/or glomerulonephritis (GMN) leading to increased mortality (Skopouli *et al*, 2000).

Sjögren's syndrome has been characterized as a systemic disorder with features of both organ-specific and systemic autoimmunity (Kassan and Moutsopoulos, 2004). Indeed, apart from exocrine glands, other organs such as kidney, liver, lungs or thyroid may be involved with the primary target, their epithelial component. This observation has led to the alternative term, 'autoimmune epithelitis' (Moutsopoulos, 1994) pointing out the fact that SS may be considered as a cell-specific autoimmune disease.

In this review, we summarize the data that support the new terminology and organize them in two axes. The first focuses on the clinical and pathological studies regarding SS and the target organs; emphasis is given on extraglandular sites and their participation in the pathogenesis of the syndrome. Secondly, we review studies that reveal the key role of the epithelial cell in the initiation and perpetuation of the autoimmune lesion through specific molecular mechanisms. Finally, we propose a theory that unifies these data and addresses the question of the initiating offending factor that leads to epithelial dysfunction.

Epithelial cell as a target – clinical manifestations

Exocrine glands

Sjögren's syndrome is classically characterized by oral (xerostomia) and ocular dryness (xeropthalmia). In a recent analysis, the presence of both dry mouth and dry eyes classified patients with 93% sensitivity and 97.7% specificity (Al-Hashimi et al, 2001). According to the American-European consensus group, six criteria are proposed for the SS diagnosis (Table 1), four of which include subjective symptoms and objective signs of oral and ocular dryness (Vitali et al, 2002). Chronic persistence of decreased salivary flow leads to depapillation of the tongue, increased incidence of dental caries, superficial mouth ulcers and cheilitis (Figure 1a) (Ravald and List, 1998). Likewise, ocular manifestations may include erosions of the cornea, conjunctivitis and in severe cases corneal ulceration (Kassan and Moutsopoulos, 2004). The most important sequel of SS, although, is the

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 Table 1
 Classification criteria for Sjögren's syndrome (SS) as proposed by the American–European consensus group (Vitali *et al*, 2002)

1. Ocular symptoms

2. Oral symptoms

(a)

- 3. Ocular signs positive Schirmer's test or rose bengal stain
- 4. Histopathology positive minor salivary gland biopsy
- Oral signs positive whole salivary flow or parotid sialography or scintigraphy
- 6. Serology positive anti- Ro/SS-A or anti-La/SS-B or both Definite primary SS:
- Presence of any 4 out of 6 (obligatory no. 4 or no. 6) Presence of any 3 out of 4 objective (no. 3 to no. 6) criteria



Figure 1 (a) A female patient with primary Sjögren's syndrome; angular cheilitis and dry tongue with atrophic papillae are evident. Note also that this patient lost many teeth while the rest are with fillings, (b) Primary Sjögren's syndrome patient with persistent left parotid gland enlargement (arrow)

progression of benign lymphoproliferation to B-cell lymphoma (mainly marginal zone lymphoma of the MALT); the relative risk is approximately 20 and major risk factors are low C4 levels, palpable purpura and the presence of mixed monoclonal cryoglobulinemia (Skopouli *et al*, 2000; Zintzaras *et al*, 2005). Clinically, the presence of significant salivary gland enlargement (Figure 1b), lymphadenopathy and skin vasculitis **Table 2** Characteristics of lymphoma development in Sjögren'ssyndrome (Voulgarelis et al, 1999; Skopouli et al, 2000)

Clinical findings
Major (mainly, bilateral parotid) gland enlargement
Lymphadenopathy
Skin vasculitis
Peripheral nerve involvement
Low-grade fever
Splenomegaly
Laboratory findings
Mixed monoclonal cryoglobulinemia
Low C4 levels
Anemia/lymphopenia
Hypogammaglobulinemia

should raise the suspicion of lymphoproliferation (Table 2).

The hallmark of SS, as it appears in labial minor salivary gland (LMSG) biopsies, is the lymphocytic infiltration of the tissues (Figure 2a). In early lesions, the infiltrate primarily consists of activated T cells (increased $CD4^+$ to $CD8^+$ ratio) and tends to form around the ductal epithelium. In advanced chronic lesions, B cells predominate and the infiltrate extends to occupy the acinar epithelium as well. Macrophage and dendritic cells appear in intense lesions with germinal center (GC) formation (Xanthou et al, 1999). Decreased production of saliva is associated with ongoing accumulation of immune cells around the epithelia and the progressive destruction of the glandular architecture. Similar changes seem to be responsible for lacrimal gland dysfunction as well, as the lacrimal glands of SS patients are enlarged and prolapse with concomitant lymphocytic infiltration (Parkin et al, 2005).

Kidnev

One of the target organs that is very often involved in SS is the kidney (Table 3). The major clinicopathologic entity is interstitial nephritis (IN) which may appear early in the progress of the disease or even may precede the onset of subjective sicca symptoms (Bossini et al, 2001). Distal renal tubular acidosis (RTA) type I, decreased concentrating ability and RTA type II are the most frequent clinical presentations. However, it should be noted that overt renal disease due to IN is relatively infrequent (< 3%) and follows a rather benign course (Goules et al, 2000). Histologically, IN resembles the lesion of LMSG (Figure 2b). Lymphocytic infiltration (mainly CD4⁺) predominates around tubular epithelial cells, plasma cells and monocytes are present and tubular atrophy and interstitial fibrosis may ensue (Rosenberg et al, 1988; Goules et al, 2000). Moreover, apoptotic death of tubular epithelia seems to play important role (Matsumura et al, 1998a); this is another common feature with LMSG pathology, as will be discussed in detail later.

Apart from IN, to a lesser degree, GMN is present in patients with SS and renal involvement. Hypertension, mild proteinuria and microscopic hematuria are the major signs and in contrast to IN, tend to present late in the progress of the disease (Skopouli, 2001). The



Figure 2 Lymphocytic infiltrates around the epithelial components (arrowheads) of various tissues, characteristic of Sjögren's syndrome. (a) Minor labial salivary gland biopsy, (b) tubular epithelia of a renal biopsy, (c) lung biopsy and (d) liver biopsy

 Table 3 Extraglandular (systemic) manifestations of Sjögren's syndrome (Kassan and Moutsopoulos, 2004)

Fatigue
Hypothyroidism
Musculoskeletal involvement
Arthralgias
Myalgias
Fibromyalgia
Skin involvement
Dryness
Skin vasculitis
Raynaud phenomenon
Renal involvement
Tubulo-interstitial nephritis
Glomerulonephritis
Pulmonary involvement
Xerotrachea and chronic cough
Bronchiolitis (small airway obstruction)
Interstitial pneumonia (IP) – non-specific IP, lymphocytic IP, usual IP
Pulmonary lymphoma
Gastrointestinal involvement
Atrophic gastritis
Autoimmune cholangitis (similar to primary biliary cirrhosis
stage I)
Autoimmune pancreatitis (commonly subclinical)
Peripheral neuropathy
Hematologic involvement
Lymphadenopathy, splenomegaly
Leukopenia, thrombocytopenia
Benign monoclonal B cell proliferation
Non-Hodgkin's lymphoma (low- or intermediate-grade B cell
MALT lymphoma)

pathogenesis in this case seems to be completely different as it may involve immune complex deposition and complement activation (Moutsopoulos *et al*, 1978) as it is underlined by the strong association of GMN with low C4 levels and the presence of mixed monoclonal cryoglobulinemia (Skopouli *et al*, 2000). Of importance, the latter two are also strong predictors for lymphoma formation and, not surprisingly, GMN and lymphoma tend to coexist in SS patients. In conclusion, renal involvement in the form of IN, although frequent, is largely subclinical for the majority of the patients with SS. Of those who present with overt renal disease (approximately 5% in a cohort of 471 patients) the percentages of IN and GMN are almost the same (Goules *et al*, 2000). The latter should be seen more as part of a generalized vasculitic syndrome, and due to the grave prognosis promptly receive the proper therapeutic intervention.

Lungs

The exact nature of pulmonary involvement in patients with SS has been a matter of debate over the past years. Depending on the methodology applied, pulmonary findings were evident in up to 75% of patients (Constantopoulos et al, 1985; Hatron et al, 1987; Matsuyama et al, 2003) but clinically significant disease approximates 10% (Constantopoulos and Moutsopoulos, 1986; Davidson et al, 2000). Dry cough is the most common and usually the only symptom and is a consequence of xerotrachea. Subclinically, abnormal pulmonary tests reveal small airway obstruction, while a detailed high-resolution CT scan detected predominantly wall thickening at the segmental bronchi (Papiris et al, 1999). Early studies using bronchoalveolar lavage showed increased numbers of CD4⁺ lymphocytes (Wallaert et al, 1987; Dalavanga et al, 1991) and subsequent reports with bronchial biopsies focalized the inflammatory cells subepithelially, involving not only the glandular but the extraglandular tissues as well (Papiris et al, 1997). Once again, the characteristics of this peribronchial inflammation highly resemble the histological picture of LMSG biopsies of SS patients (Figure 2c).

The histological pattern of pulmonary lesion changes radically when the population under study has clinically evident lung involvement. In this case, interstitial expansion of the inflammatory process is frequent and may take various forms. Non-specific interstitial pneumonia seems to be the most frequent (Ito *et al*, 2005), but lymphocytic IP and the usual IP are also present (Deheinzelin *et al*, 1996; Yamadori *et al*, 2002). Notably, these patterns are not clear-cut as they may coexist or evolve in the same individual along with the progress of the disease.

Whether IP comes on as a result of more severe bronchiolitis in those patients who finally become symptomatic is not known. It would be rational, however, to consider that an initial subclinical insult in the form of subepithelial peribronchial inflammation, in a selected minor percentage of patients, expands to the interstitial tissues and becomes clinically apparent.

Finally, ongoing lymphocytic proliferation may progress to pulmonary MALT lymphoma. It involves approximately 1–2% of primary SS patients (Cain *et al*, 1998) and as in the case of salivary glands the main histopathologic pattern is that of low-grade marginal zone lymphoma (Royer *et al*, 1997; Ito *et al*, 2005).

Liver

Liver involvement in patients with autoimmune diseases is not very common and, when present, it usually follows a benign progress. Moreover, even in the case of hepatic injury, it is very difficult to classify the lesion as primary with associated autoimmune manifestations or as an autoimmune disease with secondary liver involvement (Youssef and Tavill, 2002). This is especially prominent in the case of SS and primary biliary cirrhosis (PBC). In a study of 300 SS patients, approximately 7% had elevated liver enzymes and were positive for antimitochondrial antibodies and almost all had histological changes similar to stage-I PBC (Figure 2d). None, though, proceeded to liver damage compatible with cirrhosis (Skopouli et al, 1994). On the other hand, 38% of PBC patients were positive for antibodies against the SS antigen SS-B/La and 95% had histological changes in LMSG similar to focal sialadenitis (Hansen et al, 1988). Of special importance is the similarity of the histological lesion in the two diseases. Both are characterized by lymphocytic infiltrates (with predominance of CD4⁺ cells) that initiate around ductal epithelium (salivary or bile ducts), while these epithelial cells inappropriately express HLA class II molecules (Abraham et al, 2004). Conclusively, in view of the preservation of hepatic architecture and the benign progress of the lesion, the hepatic involvement in SS can be characterized mainly as autoimmune cholangitis (AIC) (Moutsopoulos, 1994), especially given the fact that the immunohistochemical findings in AIC and PBC are practically identical (Kaserer et al, 1998).

Thyroid

Since the first description of thyroid dysfunction in SS 40 years ago (Bertram and Halberg, 1965), many studies point out that autoimmune thyroid insult is relatively common, ranging from 10% to almost 50% (Karsh *et al*, 1980; Hansen *et al*, 1991; Perez *et al*, 1995; D'Arbonneau *et al*, 2003). Discrepancies regarding the exact percentages or even the relation between the two (Ramos-Casals *et al*, 2000) can be attributed to methodological and/or laboratory differences and more

importantly, to the fact that both entities refer to the same population (that is middle-aged women). In any case, autoimmune thyroiditis (AT) (primarily, Hashimoto thyroiditis and to a lesser extend, Graves' disease) is the main clinical form and may precede or follow the SS diagnosis by many years. Primary finding is the presence of autoantibodies against thyroid peroxidase (anti-TPO), thyroglobulin (anti-TG), T3 and/or T4. The presence of anti-TPO/anti-TG or rheumatoid factor and anti-Ro/SS-A in the sera of SS patients can be used as primary indicators of those that are prone to develop thyroid disorders in the future (D'Arbonneau *et al*, 2003).

Histological studies of AT specifically in SS patients are very sparse (Hansen et al, 1991). The histological picture of Hashimoto thyroiditis per se however, is highly similar to that of SS (D'Arbonneau et al, 2003). The infiltrate consists primarily of CD4⁺ T lymphocytes, the thyroid epithelial cells express HLA molecules class II and adhesion molecules while GC formation and progress to B cell MALT lymphoma is probable (Hanafusa et al, 1983; Aichinger et al, 1985; Tandon et al, 1992); Hashimoto thyroiditis may evolve to lymphoma in 0.5% of patients (Thieblemont et al, 2002). Moreover, one-third of patients with AT have SS features (Coll et al, 1997) and one of 10 ANApositive AT patients shares the diagnosis of SS (Tektonidou et al, 2004). Therefore, as in the case of SS and PBC, SS and AT are two autoimmune entities closely related pathogenetically, as the target in both cases seems to be the epithelial cell.

Epithelial cell as a conductor – molecular mechanisms

As aforementioned, the main target organs of SS are exocrine and especially salivary glands. MLSG biopsies are routinely performed to establish the diagnosis (Vitali *et al*, 2002), they can be easily obtained and therefore the majority of the studies regarding the pathogenetic mechanisms of SS are carried out using this tissue.

Epithelial cell: a non-professional antigen-presenting cell Professional antigen-presenting cells (APC) are dendritic cells, macrophages and B cells. These cells present antigen in the context of MHC molecules and express co-stimulatory and adhesion molecules to form immunologic synapsis with T cells (Montoya *et al*, 2002). Extensive data show that epithelial cells in SS are activated and capable to act as non-professional APC (Figure 3).

Indeed, salivary gland epithelial cells (SGEC) express MHC class I and MHC class II (HLA-DR) molecules as well as functional co-stimulatory B7.1 (CD80) and B7.2 (CD86) molecules (Moutsopoulos *et al*, 1986; Manoussakis *et al*, 1999; Kapsogeorgou *et al*, 2001a; Matsumura *et al*, 2001). Thus, they provide both signals necessary for T cell activation. B7 molecules bind CD28 on T cells (leading to activation), as well as CTLA-4, which is a negative regulator of T cells; the affinity of B7 for CTLA-4 is higher compared with CD28 and leads to



Figure 3 Epithelial T-cell interaction. The epithelial cell expresses all the molecules necessary for optimal T-cell response and therefore is a potent non-professional antigen-presenting cell

self-limitation of immune reaction. Interestingly, B7.2 molecules expressed by SGEC were found to have higher affinity for CD28 compared with the negative regulator CTLA-4 (Kapsogeorgou et al, 2001a). Intercellular adhesion molecule-1 (ICAM1) and vascular cell adhesion molecule (VCAM) are expressed on APCs and ensure stabilization of the APC/T cell synapsis. Epithelial cells on LMSG biopsies and cultured SGEC lines from SS patients express both ICAM1 and VCAM (Kapsogeorgou et al, 2001b; Tsunawaki et al, 2002). Moreover, SGEC express functional CD40 molecules and CD40L is highly expressed on the infiltrating lymphocytes (Dimitriou et al, 2002; Ohlsson et al, 2002a). The consequences of this interaction are not fully clarified but are thought to augment the inflammatory response either by directly activating T cells or by enhancing co-stimulatory and adhesion molecules, such as ICAM1, on the APC (Grewal and Flavell, 1998). Toll-like receptors (TLRs) recognize distinct pathological molecular patterns, and upon binding, lead to cytokine production and upregulation of co-stimulatory and adhesion molecules. In this way, they link innate to adaptive immunity (Takeda et al, 2003). Recent data from our group indicate that SGEC express functional TLRs, further implicating the epithelial cell in the local inflammatory responses (Spachidou et al, 2005). Finally, SGEC produce proinflammatory cytokines such as tumor necrosis factor-a, interleukin (IL)-1 and IL-6, significantly contributing to the inflammatory milieu (Boumba et al, 1995).

Interestingly, similar properties are identified for tubular epithelial cells. Indeed, B7.2 as well as ICAM and VCAM were present in renal biopsies from SS patients with IN (Matsumura *et al*, 1998b, 2001), linking the extraglandular manifestations with epithelial activation. Whether this activation results from intrinsic capacity of the epithelial cell (including a possible infectious agent) or is dictated by the extrinsic autoimmune reaction is an open question. The answer is probably both. SGEC from SS patients, even after longterm cultures *in vitro*, keep a constitutively high expression of the molecules mentioned above, indicative of intrinsic activation. On the other hand, various factors seem to control their expression; e.g. cytokines such as IFN- γ consistently upregulate both adhesion and co-stimulatory molecules.

Chemokines and germinal center formation

As discussed elsewhere, salivary gland dysfunction is related to the ongoing accumulation of lymphocytes and destruction of glandular architecture. In approximately 20% of patients (Xanthou *et al*, 1999; Salomonsson *et al*, 2003) the perpetuation of the lesion involves the formation of ectopic GC-like structures with evidence of clonal B-cell proliferation (Moutsopoulos *et al*, 1990). Chemokines are key regulators in these processes.

In brief, CXCL13 (or B-cell attractant, BCA1) attracts naïve B cells and certain T cells through the receptor CXCR5 (or BLR1) and is important for GC formation. Epithelial cells are implicated in the production of CXCL13 (Xanthou et al, 2001; Salomonsson et al, 2002, 2003) as well as endothelial cells and monocytes (Amft et al, 2001; Barone et al, 2005). Normal mechanisms such as transcytosis (capture and subsequent presentation of molecules by cells that do not synthesize them) or experimental differences may be the reason for such discrepancies. Either way, the expression of CXCR5 by infiltrating cells (Amft et al, 2001; Salomonsson et al, 2002) points out the importance of CXCL13 for cellular trafficking in LMSGs of SS patients. The same applies for CCL21 (or secondary lymphoid tissue/T-cell attractant, SLC/TCA), a chemoattractant for T cells and dendritic cells highly expressed by high-endothelial venules (HEVs). Whether epithelial (Xanthou et al, 2001; Salomonsson et al, 2003) or endothelial (Barone et al, 2005) cells are the source, CCL21 seems to lead to a higher degree of organization within the aggregate through the formation of peripheral lymph node addressin (PNAd)-positive HEVs (Barone et al, 2005). In addition, three other potent T-cell chemoattractants have been studied in SS lesions; CXCL9 (or monokine induced by INF- γ , Mig), CXCL10 (or IFN- γ inducible 10 kDa protein, IP10) and CXCL12 (or stromal-cell derived factor, SDF1). Epithelial cells from SS patients produce CXCL9 and CXCL10 while most of the $CD3^+$ lymphocytes in periductal foci express CXCR3 (common receptor for CXCL9 and CXCL10) (Ogawa et al, 2002). On the other hand, CXCL12 is constitutively expressed by both SS and controls, whereas all three chemokines are consistently upregulated by IFN-y (Amft et al, 2001; Ogawa et al, 2002). Finally, B-cell activating factor (BAFF) is a cytokine that plays an important role in various developmental stages of B cells and seems to be especially important for the survival of autoreactive B cells (Thien et al, 2004) and autoantibody production (Pers et al, 2005). BAFF is present in salivary glands of patients with SS and although unclear, epithelial cells seem to participate in its production (Groom et al, 2002).

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The initiation and perpetuation of the autoimmune lesion in SS, the organization in ectopic GCs and probably the progression to lymphoma is largely attributed to factors that favor lymphocytic homing in LMSG. Epithelial production of chemokines is crucial in these processes.

Epithelial apoptosis and exosomes

Detection in the patients' sera of antibodies against the autoantigens Ro/SS-A and La/SS-B is one of the criteria for SS diagnosis (Vitali *et al*, 2002). The Ro/La ribonucleoprotein complexes are protein-RNA complexes formed by the association of the Ro52 kDa, Ro60 kDa and La proteins with small cytoplasmic RNA (hyRNA) (Slobbe *et al*, 1992; Fabini *et al*, 2000). The Ro/La proteins are intracellular and therefore not accessible to the immune system. It takes specific mechanisms, such as apoptosis and exosomes in order for them to be disclosed and become immunogenic.

Apoptosis is a mechanism of utmost importance during embryogenesis and leads to elimination of damaged cells throughout life. The initial stimulus may be either intrinsic to the cell (e.g. uncorrectable DNA damage) or delivered extrinsically by other cells. The most important pathways of the second category are the Fas (CD95)/Fas ligand (FasL) and that of perforin and granzynes (Figure 4). Elevated apoptotic death of the epithelia from SS patients compared with the controls has been repeatedly reported, indicating that these pathways are functional and lead to epithelial destruction (Polihronis et al, 1998; Manganelli and Fietta, 2003). Fas and FasL is expressed by acinar and ductal epithelial cells, as well as by the infiltrating mononuclear cells of LMSGs and lacrimal glands from SS patients (Kong et al, 1997; Polihronis et al, 1998; Matsumura et al, 2000; Abu Helu et al, 2001; Tsubota et al, 2003). Perforin and granzyme B expression was observed in the mononuclear cell infiltrates of SS patients, but not in biopsies from control individuals



Figure 4 Apoptosis and exosomes. An initial, probably pauci-immune phase of epithelial destruction (A) is followed by a lymphocyte-dependent autoimmune aggression involving Fas/FasL and perforin/granzymes (B). This leads to typical apoptotic characteristics including bleb formation, inside-out flip of the membrane and translocation of intracellular antigens to the surface (C). Exosomes also may be vectors of autoantigens (D)

(Polihronis *et al*, 1998). Furthermore, IFN- γ induces apoptosis by at least three ways: first, upregulates Fas expression by epithelial cells (Kong et al, 1997; Matsumura et al. 2000: Abu Helu et al. 2001), secondly, upregulates CD40 expression which in turn downregulates c-FLIP (anti-apoptotic molecule downstream of Fas) (Ping et al, 2005) and thirdly, through STAT-1 mediated upregulation of caspase-8 expression (Fulda and Debatin, 2002). The study of intracellular components of the apoptotic pathway points to the same direction. The balance between apoptotic (e.g. Bax) and anti-apoptotic (e.g. Bcl-2, Bcl-xL) molecules dictates cellular fate, and several studies show Bax predominance in epithelial cells. In contrast, anti-apoptotic molecules are mainly expressed by infiltrating lymphocytes partly explaining the chronicity of the disease (Kong et al, 1998; Humphreys-Beher et al, 1999).

Key characteristics of apoptosis include bleb formation, inside-out flip of the membrane and translocation of intracellular components to the membrane (Martin *et al*, 1995). Indeed, it has been clearly shown that induction of apoptosis leads to the redistribution of La from the nucleus to the cytoplasm, while both Ro and La fill apoptotic blebs and then are exposed on the surface of the cell (McArthur *et al*, 2002; Ohlsson *et al*, 2002b). Therefore, apoptotic death not only diminishes the total number of epithelia and the functional capacity of the gland but also leads to presentation of intracellular antigens to the immune system evoking an autoimmune reaction.

Exosomes are small (30-100 nm) membrane vesicles. which result from the fusion of endosomes/lysosomes with the membrane. Dendritic cells, cytotoxic T and B lymphocytes, platelets, reticulocytes and neoplastic intestinal epithelial cells produce exosomes. Largely unclear, the biological functions of exosomes include the exclusion of obsolete proteins and membranes, while recent studies indicate that they may have immunologic functions, such as the stimulation or inactivation of T cells, directly or indirectly by the transfer of antigens to the dendritic cells (Thery et al, 2002). Recently, our group has shown that, in long-term cultures, nonneoplastic SGEC constitutively release exosomes in vitro. Notably, the exosomes secreted by SGEC contain the autoantigens Ro/SS-A, La/SS-B and Sm that are major targets of immune responses in several autoimmune disorders (Kapsogeorgou et al, 2005) and therefore seem to be a novel cell-free antigen presentation mechanism in SS (Figure 4D).

The viral hypothesis

The pathogenesis of autoimmune diseases is multifactorial and includes genetic, hormonal and environmental factors. On the other hand, a link between autoimmunity and infectious agents, namely a virus, has been speculated for a long time but unquestionable data are still lacking (Zinkernagel, 2002). HIV, HTLV-1 and HCV infections in a percentage of patients present with symptoms resembling SS (Talal *et al*, 1990; Haddad *et al*, 1992; Ohyama *et al*, 1998). None of these agents, however, can reproduce the full spectrum of

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clinical and laboratory manifestations of SS, while the vast majority of confirmed SS patients have no other evidence of HIV or HTLV-1 infection. Chronic HCV infection warrants special reference because sialadenitis is accompanied by lymphocytic infiltrate of the LMSG, highly similar to that of SS. HCV RNA is present in salivary epithelial cells in patients with chronic HCV infection (Arrieta *et al*, 2001). Moreover, mixed cryoglobulinemia and B-cell lymphoma are probable sequelae of both entities (Ramos-Casals *et al*, 2005). Still, several features differentiate chronic HCV infection and SS making an etiologic linkage less probable (Scott *et al*, 1997).

Recently, our group published data that link SS with the presence of coxsackie virus sequences in salivary gland tissues. RT-PCR for the conserved non-coding region of enteroviral RNA and subsequent sequencing, revealed 97–99% similarity with the coxsackie B4 and A13 strains (Triantafyllopoulou *et al*, 2004). These data, along with the viral ability to establish persistent noncytolytic infections *in vitro*, render coxsackie virus a probable pathogenetic factor of SS.

Summary

The target of the present paper is to point out the major role of the epithelial cell in the autoimmune lesions of SS. The exocrine manifestations of SS result mainly from the immune attack of the various organs' epithelial component. On the other hand, at the cellular/molecular level, it is clear that key pathogenetic mechanisms that lead to the initiation and perpetuation of the lesion involve the epithelial cell. We propose, therefore, the following theory that unifies most of the data, center on the role of the epithelium and justifies the name 'autoimmune epithelitis'.

In genetically liable individuals, a certain event such as a psychological and/or endocrine stress, initiates a sequel that leads to the activation of a silent offending factor (most probably an epitheliotropic virus), which results in the activation of the exocrine glands' epithelium. This leads to the recruitment, activation and expansion of lymphocytes in the lesion that in some individuals form well-organized structures resembling GCs; in patients with certain adverse prognostic factors this may progress to MALT lymphoma. Apoptosis of the epithelial cell spreads the autoimmune reaction to various antigenic determinants and as it is a normal cellturnover mechanism, may account for the immune attack on the epithelia of other organs by sensitized lymphocytes. This insult is mostly subclinical but in a minority of patients may progress to include nonepithelial structures, and therefore becomes clinically overt. Further studies aimed at the target cell will certainly clarify these issues.

References

Abraham S, Begum S, Isenberg D (2004). Hepatic manifestations of autoimmune rheumatic diseases. *Ann Rheum Dis* 63: 123–129.

- Abu Helu RF, Dimitriou ID, Kapsogeorgou EK, Moutsopoulos HM, Manoussakis MN (2001). Induction of salivary gland epithelial cell injury in Sjögren's syndrome: in vitro assessment of T cell-derived cytokines and Fas protein expression. J Autoimmun 17: 141–153.
- Aichinger G, Fill H, Wick G (1985). In situ immune complexes, lymphocyte subpopulations, and HLA-DR-positive epithelial cells in Hashimoto thyroiditis. *Lab Invest* **52**: 132–140.
- Al-Hashimi I, Khuder S, Haghighat N, Zipp M (2001). Frequency and predictive value of the clinical manifestations in Sjögren's syndrome. *J Oral Pathol Med* **30**: 1–6.
- Amft N, Curnow SJ, Scheel-Toellner D *et al* (2001). Ectopic expression of the B cell-attracting chemokine BCA-1 (CXCL13) on endothelial cells and within lymphoid follicles contributes to the establishment of germinal center-like structures in Sjögren's syndrome. *Arthritis Rheum* **44**: 2633– 2641.
- Arrieta JJ, Rodriguez-Inigo E, Ortiz-Movilla N *et al* (2001). In situ detection of hepatitis C virus RNA in salivary glands. *Am J Pathol* **158**: 259–264.
- Barone F, Bombardieri M, Manzo A *et al* (2005). Association of CXCL13 and CCL21 expression with the progressive organization of lymphoid-like structures in Sjögren's syndrome. *Arthritis Rheum* **52**: 1773–1784.
- Bertram U, Halberg P (1965). Organ antibodies in Sjögren's syndrome. *Acta Allergol* **20**: 472–483.
- Bossini N, Savoldi S, Franceschini F *et al* (2001). Clinical and morphological features of kidney involvement in primary Sjögren's syndrome. *Nephrol Dial Transplant* **16**: 2328–2336.
- Boumba D, Skopouli FN, Moutsopoulos HM (1995). Cytokine mRNA expression in the labial salivary gland tissues from patients with primary Sjögren's syndrome. *Br J Rheumatol* **34:** 326–333.
- Cain HC, Noble PW, Matthay RA (1998). Pulmonary manifestations of Sjögren's syndrome. *Clin Chest Med* **19**: 687–699, viii.
- Coll J, Anglada J, Tomas S *et al* (1997). High prevalence of subclinical Sjögren's syndrome features in patients with autoimmune thyroid disease. *J Rheumatol* **24**: 1719–1724.
- Constantopoulos SH, Moutsopoulos HM (1986). Respiratory involvement in patients with primary Sjögren's syndrome. Is it a problem? *Scand J Rheumatol* **61**(Suppl.): 146–150.
- Constantopoulos SH, Papadimitriou CS, Moutsopoulos HM (1985). Respiratory manifestations in primary Sjögren's syndrome. A clinical, functional, and histologic study. *Chest* **88**: 226–229.
- D'Arbonneau F, Ansart S, Le Berre R, Dueymes M, Youinou P, Pennec YL (2003). Thyroid dysfunction in primary Sjögren's syndrome: a long-term follow up study. *Arthritis Rheum* **49**: 804–809.
- Dalavanga YA, Constantopoulos SH, Galanopoulou V, Zerva L, Moutsopoulos HM (1991). Alveolitis correlates with clinical pulmonary involvement in primary Sjögren's syndrome. *Chest* **99:** 1394–1397.
- Davidson BK, Kelly CA, Griffiths ID (2000). Ten year of follow up of pulmonary function in patients with primary Sjögren's Syndrome. *Ann Rheum Dis* **59**: 709–712.
- Deheinzelin D, Capelozzi VL, Kairalla RA, Barbas Filho JV, Saldiva PH, de Carvalho CR (1996). Interstitial lung disease in primary Sjögren's syndrome. Clinical-pathological evaluation and response to treatment. *Am J Respir Crit Care Med* **154**: 794–799.

- Dimitriou ID, Kapsogeorgou EK, Moutsopoulos HM, Manoussakis MN (2002). CD40 on salivary gland epithelial cells: high constitutive expression by cultured cells from Sjögren's syndrome patients indicating their intrinsic activation. *Clin Exp Immunol* **127:** 386–392.
- Fabini G, Rutjes SA, Zimmermann C, Pruijn GJ, Steiner G (2000). Analysis of the molecular composition of Ro ribonucleoprotein complexes. Identification of novel Y RNA binding proteins. *Eur J Biochem* 267: 2778–2789.
- Fulda S, Debatin KM (2002). IFN-γ sensitizes for apoptosis by upregulating caspase-8 expression through the Stat1 pathway. *Oncogene* **21**: 2295–2308.
- Goules A, Masouridi S, Tzioufas AG, Ioannidis JP, Skopouli FN, Moutsopoulos HM (2000). Clinically significant and biopsy-documented renal involvement in primary Sjogren syndrome. *Medicine (Baltimore)* **79:** 241–249.
- Grewal IS, Flavell RA (1998). CD40 and CD154 in cellmediated immunity. *Annu Rev Immunol* 16: 111–135.
- Groom J, Kalled SL, Cutler AH *et al* (2002). Association of BAFF/BLyS over expression and altered B cell differentiation with Sjögren's syndrome. *J Clin Invest* **109**: 59–68.
- Haddad J, Deny P, Munz-Gotheil C *et al* (1992). Lymphocytic sialadenitis of Sjögren's syndrome associated with chronic hepatitis C virus liver disease. *Lancet* **339**: 321–323.
- Hanafusa T, Pujol-Borrell R, Chiovato L, Russell RC, Doniach D, Bottazzo GF (1983). Aberrant expression of HLA-DR antigens on thyrocytes in Graves' disease: relevance for autoimmunity. *Lancet* **2**: 1111–1115.
- Hansen BU, Lindgren S, Eriksson S *et al* (1988). Clinical and immunological features of Sjögren's syndrome in patients with primary biliary cirrhosis with emphasis on focal sialadenitis. *Acta Med Scand* **224:** 611–619.
- Hansen BU, Ericsson UB, Henricsson V, Larsson E, Manthorpe R, Warfvinge G (1991). Autoimmune thyroiditis and primary Sjögren's syndrome: clinical and laboratory evidence of the coexistence of the two diseases. *Clin Exp Rheumatol* **9**: 137–141.
- Hatron PY, Wallaert B, Gosset D *et al* (1987). Subclinical lung inflammation in primary Sjögren's syndrome. Relationship between bronchoalveolar lavage cellular analysis findings and characteristics of the disease. *Arthritis Rheum* **30**: 1226–1231.
- Humphreys-Beher MG, Peck AB, Dang H, Talal N (1999). The role of apoptosis in the initiation of the autoimmune response in Sjögren's syndrome. *Clin Exp Immunol* **116**: 383–387.
- Ito I, Nagai S, Kitaichi M *et al* (2005). Pulmonary manifestations of primary Sjögren's syndrome: a clinical, radiologic, and pathologic study. *Am J Respir Crit Care Med* **171:** 632– 638.
- Kapsogeorgou EK, Moutsopoulos HM, Manoussakis MN (2001a). Functional expression of a costimulatory B7.2 (CD86) protein on human salivary gland epithelial cells that interacts with CD28 receptor, but has reduced binding to CTLA4. *J Immunol* **166**: 3107–3113.
- Kapsogeorgou EK, Dimitriou ID, Abu-Helu RF, Moutsopoulos HM, Manoussakis MN (2001b). Activation of epithelial and myoepithelial cells in the salivary glands of patients with Sjögren's syndrome: high expression of intercellular adhesion molecule-1 (ICAM.1) in biopsy specimens and cultured cells. *Clin Exp Immunol* **124**: 126–133.
- Kapsogeorgou EK, Abu-Helu RF, Moutsopoulos HM, Manoussakis MN (2005). Salivary gland epithelial cell exosomes: a source of autoantigenic ribonucleoproteins. *Arthritis Rheum* **52:** 1517–1521.

- Karsh J, Pavlidis N, Weintraub BD, Moutsopoulos HM (1980). Thyroid disease in Sjögren's syndrome. *Arthtritis Rheum* 23: 1326–1329.
- Kaserer K, Exner M, Mosberger I, Penner E, Wrba F (1998). Characterization of the inflammatory infiltrate in autoimmune cholangitis. A morphological and immunhistochemical study. *Virchows Arch* **432**: 217–222.
- Kassan SS, Moutsopoulos HM (2004). Clinical manifestations and early diagnosis of Sjögren's syndrome. *Arch Intern Med* **164:** 1275–1284.
- Kong L, Ogawa N, Nakabayashi T *et al* (1997). Fas and Fas ligand expression in the salivary glands of patients with primary Sjögren's syndrome. *Arthritis Rheum* **40**: 87–97.
- Kong L, Ogawa N, McGuff HS *et al* (1998). Bcl-2 family expression in salivary glands from patients with primary Sjögren's syndrome: involvement of Bax in salivary gland destruction. *Clin Immunol Immunopathol* **88:** 133–141.
- Manganelli P, Fietta P (2003). Apoptosis and Sjogren syndrome. *Semin Arthritis Rheum* **33:** 49–65.
- Manoussakis MN, Dimitriou ID, Kapsogeorgou EK *et al* (1999). Expression of B7 costimulatory molecules by salivary gland epithelial cells in patients with Sjögren's syndrome. *Arthritis Rheum* **42:** 229–239.
- Martin SJ, Reutelingsperger CP, McGahon AJ *et al* (1995). Early redistribution of plasma membrane phosphatidylserine is a general feature of apoptosis regardless of the initiating stimulus: inhibition by overexpression of Bcl-2 and Abl. *J Exp Med* **182**: 1545–1556.
- Matsumura R, Umemiya K, Kagami M *et al* (1998a). Glandular and extraglandular expression of the Fas-Fas ligand and apoptosis in patients with Sjögren's syndrome. *Clin Exp Rheumatol* **16**: 561–568.
- Matsumura R, Umemiya K, Nakazawa T *et al* (1998b). Expression of cell adhesion molecules in tubulointerstitial nephritis associated with Sjögren's syndrome. *Clin Nephrol* **49**: 74–81.
- Matsumura R, Umemiya K, Goto T *et al* (2000). Interferon gamma and tumor necrosis factor alpha induce Fas expression and anti-Fas mediated apoptosis in a salivary ductal cell line. *Clin Exp Rheumatol* **18**: 311–318.
- Matsumura R, Umemiya K, Goto T *et al* (2001). Glandular and extraglandular expression of costimulatory molecules in patients with Sjögren's syndrome. *Ann Rheum Dis* **60**: 473– 482.
- Matsuyama N, Ashizawa K, Okimoto T, Kadota J, Amano H, Hayashi K (2003). Pulmonary lesions associated with Sjögren's syndrome: radiographic and CT findings. *Br J Radiol* **76:** 880–884.
- McArthur C, Wang Y, Veno P, Zhang J, Fiorella R (2002). Intracellular trafficking and surface expression of SS-A (Ro), SS-B (La), poly(ADP-ribose) polymerase and alphafodrin autoantigens during apoptosis in human salivary gland cells induced by tumour necrosis factor-alpha. *Arch Oral Biol* **47**: 443–448.
- Montoya MC, Sancho D, Vicente-Manzanares M, Sanchez-Madrid F (2002). Cell adhesion and polarity during immune interactions. *Immunol Rev* **186:** 68–82.
- Moutsopoulos HM (1994). Sjögren's syndrome: autoimmune epithelitis. *Clin Immunol Immunopathol* **72:** 162–165.
- Moutsopoulos HM, Balow JE, Lawley TJ, Stahl NI, Antonovych TT, Chused TM (1978). Immune complex glomerulonephritis in sicca syndrome. *Am J Med* **64**: 955– 960.
- Moutsopoulos HM, Hooks JJ, Chan CC, Dalavanga YA, Skopouli FN, Detrick B (1986). HLA-DR expression by labial minor salivary gland tissues in Sjögren's syndrome. *Ann Rheum Dis* **45:** 677–683.

- Moutsopoulos HM, Tzioufas AG, Bai MK, Papadopoulos NM, Papadimitriou CS (1990). Association of serum IgM kappa monoclonicity in patients with Sjögren's syndrome with an increased proportion of kappa positive plasma cells infiltrating the labial minor salivary glands. *Ann Rheum Dis* **49**: 929–931.
- Ogawa N, Ping L, Zhenjun L, Takada Y, Sugai S (2002). Involvement of the interferon- γ -induced T cell-attracting chemokines, interferon- γ -inducible 10-kd protein (CXCL10) and monokine induced by interferon- γ (CXCL9), in the salivary gland lesions of patients with Sjögren's syndrome. *Arthritis Rheum* **46**: 2730–2741.
- Ohlsson M, Szodoray P, Loro LL, Johannessen AC, Jonsson R (2002a). CD40, CD154, Bax and Bcl-2 expression in Sjögren's syndrome salivary glands: a putative anti-apoptotic role during its effector phases. *Scand J Immunol* **56**: 561–571.
- Ohlsson M, Jonsson R, Brokstad KA (2002b). Subcellular redistribution and surface exposure of the Ro52, Ro60 and La48 autoantigens during apoptosis in human ductal epithelial cells: a possible mechanism in the pathogenesis of Sjögren's syndrome. *Scand J Immunol* **56**: 456–469.
- Ohyama Y, Nakamura S, Hara H *et al* (1998). Accumulation of human T lymphotropic virus type I-infected T cells in the salivary glands of patients with human T lymphotropic virus type I-associated Sjögren's syndrome. *Arthritis Rheum* **41**: 1972–1978.
- Papiris SA, Saetta M, Turato G *et al* (1997). CD4-positive Tlymphocytes infiltrate the bronchial mucosa of patients with Sjögren's syndrome. *Am J Respir Crit Care Med* **156**(2 Pt 1): 637–641.
- Papiris SA, Maniati M, Constantopoulos SH, Roussos C, Moutsopoulos HM, Skopouli FN (1999). Lung involvement in primary Sjögren's syndrome is mainly related to the small airway disease. Ann Rheum Dis 58: 61–64.
- Parkin B, Chew JB, White VA, Garcia-Briones G, Chhanabhai M, Rootman J (2005). Lymphocytic infiltration and enlargement of the lacrimal glands: a new subtype of primary Sjögren's syndrome? *Ophthalmology* **112**: 2040–2047.
- Perez B, Kraus A, Lopez G, Cifuentes M, Alarcon-Segovia D (1995). Autoimmune thyroid disease in primary Sjögren's syndrome. Am J Med 99: 480–484.
- Pers JO, Daridon C, Devauchelle V *et al* (2005). BAFF overexpression is associated with autoantibody production in autoimmune diseases. *Ann N Y Acad Sci* **1050**: 34–39.
- Ping L, Ogawa N, Sugai S (2005). Novel role of CD40 in Fasdependent apoptosis of cultured salivary epithelial cells from patients with Sjögren's syndrome. *Arthritis Rheum* 52: 573– 581.
- Polihronis M, Tapinos NI, Theocharis SE, Economou A, Kittas C, Moutsopoulos HM (1998). Modes of epithelial cell death and repair in Sjögren's syndrome (SS). *Clin Exp Immunol* 114: 485–490.
- Ramos-Casals M, Garcia-Carrasco M, Cervera R *et al* (2000). Thyroid disease in primary Sjogren syndrome. Study in a series of 160 patients. *Medicine (Baltimore)* **79:** 103–108.
- Ramos-Casals M, DeVita S, Tzioufas AG (2005). Hepatitis C virus, Sjögren's syndrome and B-cell lymphoma: linking infection, autoimmunity and cancer. *Autoimmun Rev* **4**: 8–15.
- Ravald N, List T (1998). Caries and periodontal conditions in patients with primary Sjögren's syndrome. *Swed Dent J* 22: 97–103.
- Rosenberg ME, Schendel PB, McCurdy FA, Platt JL (1988). Characterization of immune cells in kidneys from patients with Sjögren's syndrome. *Am J Kidney Dis* **11**: 20–22.

- Royer B, Cazals-Hatem D, Sibilia J *et al* (1997). Lymphomas in patients with Sjögren's syndrome are marginal zone B-cell neoplasms, arise in diverse extranodal and nodal sites, and are not associated with viruses. *Blood* **90**: 766–775.
- Salomonsson S, Larsson P, Tengner P, Mellquist E, Hjelmstrom P, Wahren-Herlenius M (2002). Expression of the B cell-attracting chemokine CXCL13 in the target organ and autoantibody production in ectopic lymphoid tissue in the chronic inflammatory disease Sjögren's syndrome. *Scand J Immunol* **55**: 336–342.
- Salomonsson S, Jonsson MV, Skarstein K *et al* (2003). Cellular basis of ectopic germinal center formation and autoantibody production in the target organ of patients with Sjögren's syndrome. *Arthritis Rheum* **48:** 3187–3201.
- Scott CA, Avellini C, Desinan L et al (1997). Chronic lymphocytic sialoadenitis in HCV-related chronic liver disease: comparison of Sjögren's syndrome. *Histopathology* **30:** 41–48.
- Skopouli FN (2001). Kidney injury in Sjögren's syndrome. *Nephrol Dial Transplant* **16**(Suppl. 6): 63–64.
- Skopouli FN, Barbatis C, Moutsopoulos HM (1994). Liver involvement in primary Sjögren's syndrome. Br J Rheumatol 33: 745–748.
- Skopouli FN, Dafni U, Ioannidis JP, Moutsopoulos HM (2000). Clinical evolution, and morbidity and mortality of primary Sjögren's syndrome. *Semin Arthritis Rheum* 29: 296–304.
- Slobbe RL, Pluk W, van Venrooij WJ, Pruijn GJ (1992). Ro ribonucleoprotein assembly in vitro. Identification of RNA– protein and protein–protein interactions. J Mol Biol 227: 361–366.
- Spachidou MP, Bourazopoulou E, Kapsogeorgou EK *et al* (2005). Functional Toll-like receptors (TLRs) are expressed by cultured salivary gland epithelial cells: increased mRNA expression of TLR-1, -2 and -4 in cells derived from patients with primary Sjögren's syndrome. *Arthritis Rheum* **52**: 278 (abstract 684).
- Takeda K, Kaisho T, Akira S (2003). Toll-like receptors. Annu Rev Immunol 21: 335–376.
- Talal N, Dauphinee MJ, Dang H, Alexander SS, Hart DJ, Garry RF (1990). Detection of serum antibodies to retroviral proteins in patients with primary Sjögren's syndrome (autoimmune exocrinopathy). Arthritis Rheum 33: 774–781.
- Tandon N, Makgoba MW, Gahmberg CG, Weetman AP (1992). The expression and role in T cell adhesion of LFA-3 and ICAM-2 on human thyroid cells. *Clin Immunol Immunopathol* **64**: 30–35.
- Tektonidou MG, Anapliotou M, Vlachoyiannopoulos P, Moutsopoulos HM (2004). Presence of systemic autoimmune disorders in patients with autoimmune thyroid diseases. *Ann Rheum Dis* 63: 1159–1161.
- Thery C, Zitvogel L, Amigorena S (2002). Exosomes: composition, biogenesis and function. *Nat Rev Immunol* **2:** 569–579.
- Thieblemont C, Mayer A, Dumontet C *et al* (2002). Primary thyroid lymphoma is a heterogeneous disease. *J Clin Endocrinol Metab* 87: 105–111.
- Thien M, Phan TG, Gardam S *et al* (2004). Excess BAFF rescues self-reactive B cells from peripheral deletion and allows them to enter forbidden follicular and marginal zone niches. *Immunity* **20**: 785–798.
- Triantafyllopoulou A, Tapinos N, Moutsopoulos HM (2004). Evidence for coxsackievirus infection in primary Sjögren's syndrome. *Arthritis Rheum* **50**: 2897–2902.
- Tsubota K, Fujita H, Tsuzaka K, Takeuchi T (2003). Quantitative analysis of lacrimal gland function, apoptotic figures, Fas and Fas ligand expression of lacrimal glands in dry eye patients. *Exp Eye Res* **76**: 233–240.

- Tsunawaki S, Nakamura S, Ohyama Y *et al* (2002). Possible function of salivary gland epithelial cells as non-professional antigen-presenting cells in the development of Sjögren's syndrome. *J Rheumatol* **29**: 1884–1896.
- Vitali C, Bombardieri S, Jonsson R *et al* (2002). Classification criteria for Sjögren's syndrome: a revised version of the European criteria proposed by the American-European Consensus Group. *Ann Rheum Dis* **61**: 554–558.
- Voulgarelis M, Dafni UG, Isenberg DA, Moutsopoulos HM (1999). Malignant lymphoma in primary Sjögren's syndrome: a multicenter, retrospective, clinical study by the European Concerted Action on Sjögren's Syndrome. *Arthritis Rheum* **42:** 1765–1772.
- Wallaert B, Prin L, Hatron PY, Ramon P, Tonnel AB, Voisin C (1987). Lymphocyte subpopulations in bronchoalveolar lavage in Sjögren's syndrome. Evidence for an expansion of cytotoxic/suppressor subset in patients with alveolar neutrophilia. Chest 92: 1025–1031.
- Xanthou G, Tapinos NI, Polihronis M, Nezis IP, Margaritis LH, Moutsopoulos HM (1999). CD4 cytotoxic and dendritic cells in the immunopathologic lesion of Sjögren's syndrome. *Clin Exp Immunol* **118**: 154–163.

- Xanthou G, Polihronis M, Tzioufas AG, Paikos S, Sideras P, Moutsopoulos HM (2001). "Lymphoid" chemokine messenger RNA expression by epithelial cells in the chronic inflammatory lesion of the salivary glands of Sjögren's syndrome patients: possible participation in lymphoid structure formation. *Arthritis Rheum* **44**: 408–418.
- Yamadori I, Fujita J, Bandoh S *et al* (2002). Nonspecific interstitial pneumonia as pulmonary involvement of primary Sjögren's syndrome. *Rheumatol Int* **22**: 89–92.
- Youssef WI, Tavill AS (2002). Connective tissue diseases and the liver. J Clin Gastroenterol **35**: 345–349.
- Zinkernagel RM (2002). Anti-infection immunity and autoimmunity. Ann N Y Acad Sci 958: 3–6.
- Zintzaras E, Voulgarelis M, Moutsopoulos HM (2005). The risk of lymphoma development in autoimmune diseases: a meta-analysis. *Arch Intern Med* **165**: 2337–2344.

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