http://www.blackwellmunksgaard.com

## HOT TOPIC

# **Probiotics: contributions to oral health**

JH Meurman<sup>1</sup>, I Stamatova<sup>2</sup>

<sup>1</sup>Institute of Dentistry, University of Helsinki and Department of Oral and Maxillofacial Diseases, Helsinki University Central Hospital, Finland; <sup>2</sup>Department of Operative Dentistry and Endodontics, Faculty of Dentistry, Medical University, Plovdiv, Bulgaria

Probiotics have been extensively studied for their healthpromoting effects. The main field of research has been in the gastrointestinal tract. However, in the past few years probiotics have also been investigated in the oral health perspective, which is the topic of the present review. We discuss the mechanisms of bacterial adhesion, potential of probiotics in oral cavity colonization, interspecies interactions, and possible effects on immunomodulation, and means of probiotic administration. We suggest that probiotic treatment of diseases other than dental caries and periodontal disease should also be systematically investigated. In general, hardly any randomized controlled trials have been conducted in this area and the studies on probiotics vs oral health are still in their cradle. Hence, much more investigations are called for before any evidence-based conclusions can be drawn: if or not probiotic therapy can be recommended for oral health purposes.

Oral Diseases (2007) 13, 443–451

**Keywords:** probiotics; lactobacilli; bifidobacteria; W. *cibaria*; oral health; *Candida* 

### Introduction

Over the past 5 or 6 years there has been a nearexponential increase in publications on probiotics. A Medline search indicates that for the period since January 2006 more than 360 articles addressing probiotics have been published. The term 'probiotics' has undergone several definitions arriving at the final one, officially adopted by the International Scientific Association for Probiotics and Prebiotics term, outlining the breadth and scope of probiotics as they are known today: 'Live microorganisms, which when administered in adequate amounts, confer a health benefit on the host' (Guarner et al, 2005). The idea of probiotics dates back to the first decade of 1900 when the Ukrainian bacteriologist and Nobel Laureate Ilya Metchnikof (1908) studying the flora of the human intestine developed a theory that senility is caused by poisoning of the body by the products of some of these bacteria. To prevent the multiplication of these organisms he proposed a diet containing milk fermented by lactobacilli which produce large amounts of lactic acid and for a time this diet became widely popular. Probiotic organisms are thought to act through a variety of mechanisms including the competition with potential pathogens for nutrients or enterocyte adhesion sites, including degradation of toxins, production of antimicrobial substances, and local and systemic immunomodulation (Silva et al, 1987; Lewis and Freedman, 1998; Isolauri et al, 2001). The latter definition is better because it does not restrict the application of the term only to probiotics with intestinal outcomes. Table 1 presents some of the main fields of activity of probiotics in general medicine. Discussing these investigations in detail, however, is beyond the scope of the present review.

There are a number of different organisms that can be classified as probiotics. The most common probiotic strains belong to the genera *Lactobacillus* and *Bifidobacterium*. *Lactobacillus* species from which probiotic strains have been isolated include *L. acidophilus*, *L. johnsonii*, *L. casei*, *L. rhamnosus*, *L. gasseri*, and *L. reuteri*. *Bifidobacterium* strains include *B. bifidum*, *B. longum*, and *B. infantis*.

Within dentistry, studies with *L. rhamnosous* GG (Meurman *et al*, 1994; Näse *et al*, 2001; Ahola *et al*, 2002), *L. reuteri* (Nikawa *et al*, 2004) have defined their potential in interacting with *S. mutans* by reducing the number of this caries pathogen, thus suggesting a role of probiotics in caries prophylaxis. Similarly, our group has recently observed that probiotic administration reduced oral *Candida* counts in the elderly – a finding that might offer a new strategy for controlling oral yeast infections (Hatakka *et al*, 2007).Yet, there is a paucity of information regarding the contributions of probiotics to oral health. The present article aims at summarizing the literature published in the past few years with respect

Correspondence: Prof. JH Meurman, PB 41, FI00014 University of Helsinki, Helsinki, Finland. Tel: +358 9 19127 272, Fax: +358 9 19127 517, E-mail: jukka.meurman@helsinki.fi Received 16 October 2006; accepted 16 January 2007

Table 1	Examples	of recent	studies	where	probiotics	have	been	investigated	

Scopes of activity	Reference	Result
Cancer risk reduction	El-Nezami et al (2006)	Possible reduction of liver cancer
	Mego et al (2005)	Interaction with pathogenic gastrointestinal bacteria leading to decreased cancer risk
	Commane et al (2005)	Multifactorial anticarcinogenic activity
Gastrointestinal health	Brzozowski et al (2006)	Attenuation of the adverse effects of <i>Helicobacter pylori</i>
	Bergonzelli et al (2005)	General review over the issue defining the positive effect of probiotics
Urinary tract health	Reid <i>et al</i> (2001)	Coaggregation with uropathogens
	Falagas et al (2006)	Prevent urogenital infections in women
Immune response induction	Christensen et al (2006)	Insufficient data to prove the positive probiotic effect
*	Rinne et al (2005)	Elevated numbers of IgM, IgA, IgG-secreting cells
Antimicrobial potential	Olivares et al (2006)	Antibacterial activity against different pathogenic bacteria
Ĩ	Hutt et al (2006)	Strain-dependent antibacterial effect
Cardiovascular system	Aihara et al (2005)	High blood pressure reduction
<b>9</b>	Jauhiainen et al (2005)	Blood pressure lowering effect

 Table 2 Test strains considered probiotics in the oral cavity

Test strain	Reference	Type of experiment	Feature tested	Result
S. salivarius	Burton et al (2006a)	In vivo	Reduction of VSC	Reduced VSC levels
L. rhamnosus GG	Busscher et al (1999)	In vitro	Inhibition of S. mutans	Positive correlation to S. mutans inhibition
L. acidophilus		In vivo		
L. casei	C. 1. (1(2000))	<b>r</b> .		
L. reuteri	Çaglar <i>et al</i> (2006)	In vivo	Inhibition of <i>S. mutans</i>	Reduced S. mutans levels
Bifidobacterium DN-173 010	Çaglar et al (2005a)	In vivo	Inhibition of S. mutans	Reduces levels of caries pathogens
L. rhamnosus GG	Hatakka et al (2007)	In vivo	Inhibition of C. albicans	Reduce high yeast counts
Propionibacterium				
freudenreichii ssp.				
shermanii JS				
L. rhamnosus	Haukioja et al (2006a)	In vitro	Adherence	Better adherence than bifidobacteria
L. paracasei			Survival in saliva	
L. johnsonii		<b>r</b> .		
L. rhamnosus GG	Haukioja et al (2006b)	In vitro	Inhibition of S. mutans	Inhibit S. mutans adhesion to salivary pellicle
L. casei				
L. reuteri W. cibaria	$V_{app,\alpha}$ at $al(2005)$	In vitro	Adherence	S protoin positively effects adhesion
	Kang <i>et al</i> (2005)			S-protein positively affects adhesion
W. cibaria	Kang <i>et al</i> (2006)	In vivo	Reduction of VSC	Inhibited production of VSC
L. casei Shirota L. acidophilus	Lima <i>et al</i> (2005)	In vitro	Adhesion	Different pattern of adhesion according to the test strain
L. rhamnosus GG	Yli-Knuuttila et al (2006)	In vivo	Adherence	Only temporary colonization in oral cavity

VSC, volatile sulfur compounds.

to the possible role of probiotics on oral and dental health. Table 2 presents the possible probiotic strains in the oral cavity. In 2005 two reviews on probiotics in the oral health perspective were published (Çaglar *et al*, 2005a,b; Meurman, 2005). The reader of the current text is advised to refer to these recent reviews for earlier data.

## Probiotic strains in the oral cavity

Figure 1 presents the ultrastructure of one of the most studied probiotic bacterium, *Lactobacillus rhamnosus* GG. An essential requirement for a microorganism to be 'an oral probiotic' is its ability to adhere to and colonize surfaces in the oral cavity. Microorganisms generally considered as probiotics may not have oral cavity as their inherent habitat and, subsequently, their possibility to confer benefit on oral health is then questionable. Paster *et al* (2001) in an attempt to determine bacterial diversity in the human subgingival plaque by using

culture-independent molecular methods have estimated that the total species diversity in the oral cavity ranges between 500 and 600 species. This number was further extended by Kazor *et al* (2003), who detected 200 additional unknown species on the dorsum of the tongue, making the number of species in the mouth to reach 700. Lactobacilli make approximately 1% of the cultivable oral microflora (Marsh and Martin, 1999).

The most common lactobacilli species recovered from saliva in a study by Teanpaisan and Dahlen (2006) were *L. fermentum, L. rhamnosus, L. salivarius, L. casei, L. acidophilus* and *L. plantarum.* Three of them are probiotic strains used in dairy products. A similar diversity in the oral lactobacilli flora was observed by Colloca *et al* (2000) who found *L. fermentum, L. plantarum, L. salivarius* and *L. rhamnosus* to be the predominant species in healthy human mouth. Kõll-Klais *et al* (2005) found no differences in salivary lactobacilli counts between chronic periodontitis and

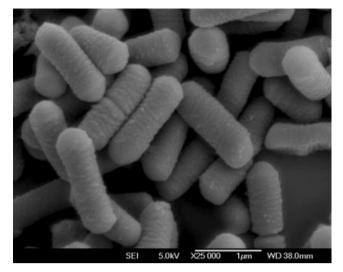


Figure 1 Scanning electron micrograph showing the typical form of probiotic strain *Lactobacillus rhamnosus* GG successfully tested in oral health trials (photograph: Kari Lounatmaa)

healthy patients, L. gasseri and L. fermentum being the predominant species among other isolates: L. oris, L. plantarum, L. paracasei, L. rhamnosus, L. gasseri, L. acidophilus and L. cispatus. However, in a later study the same authors observed a higher prevalence of homofermentative lactobacilli in healthy mouths compared to samples from patients with chronic periodontitis (Kõll-Klais et al, 2006). These findings indicate that lactobacilli as members of resident oral microflora could play an important role in the microecological balance in the oral cavity. These studies further demonstrated that lactobacilli strains with probiotic properties may indeed be found in the oral cavity. Yet there is no evidence whether these lactobacilli strains were detected due to the frequent consumption of dairy products leading to temporary colonization only, or if the oral environment is their permanent habitat. There are no long-term follow-up studies published to answer this question.

One mechanism of action of probiotics is suggested to be their modulation of host immune response. Immune inductive sites in the oral cavity are within the diffuse lymphoid aggregates of the Waldeyer's ring. Lingual and pharyngeal tonsils and adenoids contain most of the lymphatic tissue. The role of these anatomic structures as inductive sites of mucosal immunity has been shown by intranasally delivered vaccines (Wu *et al*, 1997). Dendritic cells scattered in mucosal surfaces are pivotal in the front-line bacterial recognition (antigen presentation) and in activating T-cell responses. Depending on the signals from dendritic cells either immune tolerance or active immune response toward a specific antigen may occur (Banchereau and Steinman, 1998). A marked production of interleukin-10 by dendritic cells in gut mucosa has been registered after administration of a probiotic mixture (Hart *et al*, 2004). However, more studies on activation of the oral immune inductive sites after probiotic administration are needed before further conclusions can be drawn. Such investigations might even cast light on probiotic effects in general and substantiate their specific applications in the future.

## Probiotic activity in the oral cavity

Attachment, adhesion, and oral colonization of probiotics The mechanism of adhesion to oral surfaces is an issue of importance for the long-term probiotic effect of the microorganisms. Among the different assays available to study the adhesion phenomenon, two model systems predominate: systems using saliva-coated hydroxylapatite, and hydroxylapatite coated with buffers, proteins, and other substances (Ostengo and Nader-Macias, 2004). The pattern of adhesion of different probiotic strains to oral epithelial cells has been tested as well. Most of the experiments on adhesion have been carried out with strains broadly used as probiotics in dairy products such as yogurt and cheese (Table 3).

Yli-Knuuttila *et al* (2006) assessed colonization of *L. rhamnosus* GG (LGG) in the oral cavity of healthy students. After the 14-day trial period, the occurrence of LGG in the oral cavity decreased gradually, indicating that no permanent colonization had occurred and that the oral persistence of LGG was only temporary. However, further colonization studies with larger materials and in different patient groups are still needed.

A relatively new strain and a potential candidate for a probiotic, *Weissella cibaria*, isolated from humans and animals worldwide, as well as from fermented foods, was tested for co-aggregation ability with *Fusobacterium nucleatum* and their attachment to epithelial cells (Kang *et al*, 2005). *Fusobacterium nucleatum* plays an important role as a bridge-organism that facilitates the colonization of other bacteria by co-aggregation

Vehicle	Strain	Outcome	Reference
Lozenge	S. salivarius	Reduces oral VSC levels	Burton et al (2005)
Straw, tablet	L. reuteri ATCC 55 730	S. mutans level reduction	Caglar et al (2006)
Yoghurt	Bifidobacterium DN-173 010	Reduction of salivary S. mutans	Caglar et al (2005b)
Cheese	L. rhamnosus GG; Prorionibacterium JS	Reduced risk of high yeast counts and hyposalivation	Hatakka et al (2007)
Rinse solution	W. cibaria	Reduction of VSC	Kang et al (2006)
Capsule, liquid	L. sporogenes, L. bifidum, L. bulgaricus, L. thermophilus, L. acidophilus, L. casei, L. rhamnosus	Increased salivary counts of lactobacilli without significant decrease in S. mutans counts	Montalto et al (2004)
Yogurt drink	L. rhamnosus GG	Temporary oral cavity colonization	Yli-Knuuttila et al (2006

(Kolenbrander, 2000). Many authors have reported that the co-aggregation abilities of lactobacilli species might enable them to form a barrier that prevents colonization of pathogenic bacteria (Reid et al. 1988: Boris et al. 1997), due to the production of a microenvironment around these pathogens in which inhibiting substances were generated by Lactobacillus species. Kang et al (2005) reported that W. cibaria efficiently co-aggregated with F. nucleatum. Pronase treatment led to additional reduction in co-aggregation between both species, thus indicating the proteinaceus character of the interspecies interaction. Heat-resistant components firmly attached to the cell surface of W. cibaria were responsible for the co-aggregation with F. nucleatum. The results of this study clearly showed that the S-layer proteins of the bacterial cell wall may play an important role in the adherence of W. cibaria to the epithelial cells.

In a study addressing the survival of bacteria in saliva and their adherence to oral surfaces, Haukioja et al (2006a.(b)) tested the colonization potential of different commercially available probiotics and Lactobacillus and Bifidobacterium strains obtained from the dairy industry. The results cast light on several controversial points reflecting mechanisms of colonization in the oral cavity. All test strains demonstrated 24-h survival rates in saliva but with great variations among the strains in their binding capacity to the saliva-coated surfaces. Lactobacilli showed better adherence than bifidobacteria. Thus, lactobacilli may compete for the same binding sites on saliva coated hydroxylapatite with F. nucleatum which explains their lower colonization capacity. This phenomenon indicates that probiotics might affect the formation of oral biofilms and modify resident microflora. Haukioja et al (2006a,b) defined a novel mechanism whereby lactobacilli and B. lactis Bb12 affected the composition of salivary pellicle on hydroxyapatite and thereby inhibited S. mutans adherence in vitro.

## Studies on probiotics and dental caries

The impact of oral administration of probiotics on dental caries has been studied in several experiments utilizing different test strains. Lactobacillus rhannosus GG (Meurman et al, 1994; Näse et al, 2001; Ahola et al, 2002) and L. casei (Busscher et al, 1999) have proved their potential to hamper growth of these oral streptococci. Çaglar et al (2006) registered definite S. mutanscount reduction after a 2-week consumption of yoghurt containing L. reuteri. A temporary reduction in S. mutans was observed during the period of yogurt intake and few days after cessation of consumption, indicating the necessity of continual administration of the probiotic in order to achieve an effect.

Little information is available about the relationship between probiotic bifidobacteria and counts of *S. mutans.* The only study addressing this study question tested *Bifidobacterium* DN-173 010 (Çaglar *et al*, 2005a,b). A statistically significant reduction in salivary mutans streptococci was observed. Due to the limitations of the study protocol with bifidobacteria, however, further investigations are needed for drawing final conclusions. Considering the growing body of evidence about the role of probiotics on caries pathogens, however, it has been suggested that the operative approach in caries treatment might be challenged by probiotic implementation with subsequent less invasive intervention in clinical dentistry (Anderson and Shi, 2006). However, we strongly feel that more studies are definitely needed before this goal could be achieved. Most of the studies cited above do not meet the criteria of investigations for evidence-based medicine.

## Probiotics and periodontal disease

Another issue in oral pathology, chronic periodontitis, could also benefit from orally administered probiotics. The presence of periodontal pathogens could be regulated by means of antagonistic interactions. A decrease in gum bleeding and reduced gingivitis has been observed by Krasse *et al* (2006) with the application of *L. reuteri*. Kõll-Klais *et al* (2006) reported that resident lactobacilli flora inhibits the growth of *Porphyromonas gingivalis* and *Prevotella intermedia* in 82% and 65%, respectively.

Probiotic strains included in periodontal dressings at optimal concentration of  $10^8$  CFU ml<sup>-1</sup> were shown to diminish the number of most frequently isolated periodontal pathogens: *Bacteroides* sp., *Actinomyces* sp. and *S. intermedius*, and also *C. albicans* (Volozhin *et al*, 2004). These authors registered a 10- to 12-month remission period after periodontal treatment by application of the periodontal dressing that comprised collagen and *L. casei*. Nevertheless, similar to the case with dental caries, however, there is not yet any true evidence on the effect of probiotic therapy on periodontal disease.

## Probiotics and imbalanced oral ecosystem

Halitosis, the oral malodor, is a condition normally ascribed to disturbed commensal microflora equilibrium. It has recently been positively affected by regular administration of probiotics. Kang et al (2006) have shown a definite inhibitory effect on the production of volatile sulfur compounds (VSC) by F. nucleatum after ingestion of Weissella cibaria both in vitro and in vivo. In children, a marked reduction in the levels of H<sub>2</sub>S and CH<sub>3</sub>SH by approximately 48.2% (*P* < 0.01) and 59.4%(P < 0.05), respectively, was registered after gargling with W. cibaria containing rinse. The possible mechanism in the VSC reduction is the hydrogen peroxide generated by W. cibaria that inhibits the proliferation of F. nucleatum. Streptococcus salivarius, also a possible candidate for an oral probiotic, has demonstrated inhibitory effect on VSC by competing for colonization sites with species causing an increase in levels of VSC (Burton et al, 2005, 2006a,b). Burton et al (2006a,b) further reported that S. salivarius strain K12 produced two lantibiotic bacteriocins, compounds that are inhibitory to strains of several species of gram-positive bacteria implicated in halitosis. However, the few studies published on the role of probiotics in the treatment of halitosis do not entitle any evidence-based conclusions. Nevertheless, we think that this might be an

area where probiotic therapy indeed could bring something new if the preliminary observations on the 'balancing' effect of probiotics on VSC-generating microflora are confirmed. Randomized, blinded, and placebo-controlled studies with large enough patient materials are also needed in this area.

### Probiotics and yeasts

Oral cavity with its variety of functions and complex structures is a specific site with its inherent pathology and diseases although the mouth is of course closely related to other parts and systems of the body. Candida albicans is among the most common infectious agents in the oral cavity. The incidence of yeast infections is higher at older age and under conditions of impaired immunity. Testing the pattern of colonization of L. acidophilus and L. fermentum, Elahi et al (2005) showed a rapid decline in C. albicans in mice after the intake of probiotic strains. Continuous consumption of probiotics led to almost undetectable numbers of fungi in the oral cavity, maintaining the protective effect for a prolonged period after cessation of application. The capacity of different lactobacilli to stimulate cellular and humoral factors of mucosal protection varies particularly in terms of salivary nitrous oxide and  $\gamma$ -interferon levels. Elahi et al (2005) have observed a correlation between the highest peak of interleukin-4 secretion and complete eradication of C. albicans. The results obtained in animal studies, however, require further testing of the effect of the strains on cases with clinically manifested C. albicans infection in humans.

A reduction in the prevalence of *C. albicans* in the elderly after consumption of probiotic cheese containing *L. rhamnosus* GG and *Propionibacterium freudenreichii* ssp. *shermanii* JS has been registered by Hatakka *et al* (2007) which was as an interesting observation in this randomized placebo-controlled trial. A concomitant feature of the probiotic activity observed in this study was the diminished risk of hyposalivation and the feeling of dry mouth of the subjects. The authors had no explanation to this and the finding certainly needs to be confirmed in further investigations.

It could be hypothesized that extending research on oral pathology, such as yeast infections, with respect to probiotics, and analyzing the molecular mechanisms of probiotic activity, might further broaden the field of their potential applications.

### Administration of probiotics

Appropriate forms of administration of probiotic strains have been discussed in several articles. Dairy products supplemented with probiotics are a natural means of oral administration and easily adopted in dietary regime. However, for the purposes of prevention or treatment of oral diseases, specifically targeted applications, formulas, devices, or carriers with slow release of probiotics might be needed.

Montalto *et al* (2004) administered probiotic mix both in capsules and in liquid form without observing statistically significant difference, however, in the *S. mutans* counts between the two test groups. A specially designed straw with a reservoir containing probiotics has also been presented by Çaglar *et al* (2006) who compared the effect of two non-dairy delivery methods, a Life top straw (BioGaia AB, Stockholm, Sweden) and a lozenge on the effectiveness of *L. reuteri* to reduce the number of *S. mutans*. Both means of administration showed significant reduction in salivary *S. mutans* levels in half of the patients when compared with subjects who received placebo.

A recent invention for caries prophylaxis is a chewing gum containing *L. reuteri Prodentis*. Consumed twice daily this was marketed to regulate *S. mutans* counts in the oral cavity (http://www.biogaia.se). The average content of *L. reuteri* was  $10^8$  CFU ml<sup>-1</sup>. However, we conclude that the most suitable means of delivery and dosages of probiotics for various oral health purposes have not been defined. Table 2 summarizes the variety of vehicles used so far for the administration of probiotics for oral health purposes.

## Safety aspects

The issue of safety is of special concern during the past few years due to the increased probiotic supplementation of different food products. From the safety point of view, the putative probiotic microorganisms should not be pathogenic, should not have any growth-stimulating effects on bacteria causing diarrhea, and should not have an ability to transfer antibiotic resistance genes. The probiotics should rather be able to maintain genetic stability in oral microflora (Grajek *et al*, 2005).

The increased probiotic consumption inevitably leads to increased concentrations of these species in the host organism. Lactobacillus bacteremia is a rare entity, and data on its clinical significance are mainly found through case reports. For the last 30 years there have been approximately 180 reported cases (Boriello et al, 2003). Clinical characteristics of Lactobacillus bacteremia are highly variable, ranging from asymptomatic to septic shock-like symptoms. Any viable microorganism is capable of causing bacteremia, however, especially in patients with severe underlying diseases or in immunocompromised state. Nevertheless, the present literature supports the conclusion that the incidence of Lactobacillus bacteremia is unsubstantial and that all the cases where it has been registered are individuals with other systemic diseases such as diabetes, cardiovascular diseases, gastrointestinal disorders, malignancies, or organ transplant patients (Husni et al, 1997; Cannon et al, 2005). However, it is evident that careful monitoring is needed in this regard in the future.

Several studies have been carried out in immunocompromised patients. In a controlled study exposing 35 HIV-positive patients to *L. reuteri*, no clinically significant side effects were noted (Wolf *et al*, 1998). Salminen *et al* (2002) found no increase in *Lactobacillus* bacteria in blood culture samples when screening the Finnish population for the period 1990–2000. Specifically, their study showed no increase related to the increasing probiotic use of LGG-containing commercial dairy products during that period. Further, Salminen

(2006) has recently reported no adverse effects caused by LGG ingestion, or LGG treatment in general, on HIVpositive patients. CD4<sup>+</sup> cell counts or viral load levels were analyzed and all these patients received highly active antiretroviral therapy. Consequently, LGG-containing products are not likely to exert any major health risks among HIV-positive patients (Salminen, 2006).

An indirect proof of safety might be the results of studies investigating lactobacilli species as live vectors in delivery of antigens at mucosal sites. Animal experiments have shown that L. lactis, L. casei, L. plantarum, L. helveticus and recombinant L. plantarum are capable of inducing both systemic and mucosal immune response against S. pneumoniae antigens and tetanus toxin, respectively, delivered by an intranasal route (Grangette et al. 2001; Oliveira et al. 2006).

The absence of acquired antibiotic resistances is another safety criterion to be tested in potential probiotic candidates. Some probiotics are closely related to opportunistic bacteria and this may also cause transferral of antimicrobial resistance genes in between microorganisms (Lester et al, 2006). Several results from antibiotic susceptibility tests claim that the tet(W) and tet(S) genes in some probiotic lactobacilli and bifidobacteria strains are responsible for gentamycin, sulfamethoxazole, polymyxin B, and tetracycline resistance (Huys et al, 2006; Masco et al, 2006). These investigations emphasize the need for a minimal safety evaluation during the selection of strains for probiotic use. However, further studies are also needed in this area because the increasing number of species that develop resistance to commonly used antimicrobial drugs is of great global concern. Hence, before any recommendations can be given for probiotic therapy in preventing and/or treating microbial infections instead of using antibiotic or antifungal drugs, transferral of resistance genes needs to be carefully investigated.

## Conclusions and recommendations for future research

The present review briefly outlines the potential for probiotic strains in improving oral and dental health. Similar to their better known actions in the gastrointestinal tract, probiotics exert their effects in many ways also in the oral cavity. The mechanisms of probiotic action in the mouth are anticipated to be similar to that observed with gastrointestinal indications. However, data on 'oral probiotics' are yet insufficient, and it is not known whether the putative probiotic strains could modulate, for example, immune response in the oral cavity as has been suggested to take place in the gut mucosa. The epithelial structure and chemical composition of excretions in the gastrointestinal tract differ from those in the mouth mucosa and saliva. The resident microbiota is also different in these anatomic

Problem	Recommendation/comment	<b>Table 4</b> Problems and recommendationsin oral probiotic research	
Complex microbiology of the oral cavity	Systematic screening for potential resident probiotic strains. Interactions between		
	microorganisms of the mouth are poorly understood		
Different microbial attachment sites	Investigating microbial (probiotic) attachment separately on the teeth, and on keratinized and non-keratinized epithelium. Probably different probiotics are needed for therapy in dental and oral mucosal diseases		
Saliva	Salivary defense mechanisms, both specific and nonspecific, should be investigated in relation to potential probiotics. Data from gastrointestinal studies are not directly applicable in saliva parameters		
Safety	Strains that readily ferment dietary carbohydrates and decrease pH in the mouth are not suitable probiotics for oral health purposes. In addition, general safety aspects such as those related to potential invasiveness and antibiotic resistance genes must be screened		
Means of administration and dosage	Slow-release approach should be investigated. It appears that probiotic therapy in order to be effective needs to be continuously administered. Optimal dosages of probiotics in oral health indications need to be assessed		
Trials	Randomized controlled trails are needed with patients materials based on proper power calculations. Probiotic intervention should be tested in the clinical setting using potential strains for specific oral health purposes		
Genetically modified microorganisms	Whether or not potentially probiotic microbial strains can or should be genetically modified in order to strengthen their beneficial potential or characteristics needs to be investigated. In the first hand, this calls for extensive studies on the mechanisms of probiotic action		

sites. Consequently, results from studies conducted in patients with gastrointestinal diseases cannot be directly adopted in oral medicine and dentistry.

The oral cavity with its diversity of microbial species has been shown to harbor strains also distinguished as probiotics as such. In this regard further studies identifying resident probiotics of the mouth, clarifying the mechanism of their colonization, and the eventual effect on the oral environment are needed. Studies of the probiotic effect on the balance of the oral ecosystem would also be needed. Of particular interest might be studies on the combined effect of different probiotics applied simultaneously, thus testing the possible additive, cumulative, or competitive modes of action in the oral environment.

The studies quoted here for the safety of probiotics may be regarded as a starting point for further and more thorough research. We recommend focusing both on novel strains and conducting studies where currently known probiotic microorganisms would be investigated in the oral health perspective. For example, many starter cultures have been studied for probiotic effect with noncontributory results but this area is still far from proper coverage. Consequently, more studies are needed here, too. Further, the basic criteria a strain should meet in order to be considered as an 'oral probiotic' might vary depending on the specific indications for which it is anticipated. When compared with the criteria appropriate for respective strains in other parts of the gastrointestinal tract, oral applications may need modification. In other words, for an 'oral probiotic' different criteria may be needed than for those of other health indications. Systematic screening and discovery of 'latent' or 'resident' probiotic microorganisms is needed to identify the best candidate probiotics for oral and dental diseases.

Understanding the mechanisms whereby probiotic species modulate oral immunity is important, and the role of probiotic therapy in the treatment of oral manifestations of other diseases such as cutaneous diseases should also be investigated. There are no data as to whether probiotics exert any effect on oral manifestations of autoimmune diseases. In this regard it might be interesting to conduct studies on patients with lichen planus, pemphigus vulgaris, cicatricial pemphigoid, or aphthous stomatitis.

So far the vehicles for administration of probiotics have mainly been dairy products, most of which are produced by lactic acid fermentation. Species that ferment sugar and lower oral pH are detrimental to the teeth. Hence, systematic studies and randomized controlled trials are called for to find out the best probiotic strains and means of their administration in different oral health indications. Finally, possibilities to genetically modify or engineer potential probiotic strains may offer totally new visions and need to be studied. Table 4 briefly summarizes some of the aspects we have discussed here.

### Acknowledgement

JHM was supported by a grant from the Juho Vainio Foundation, Helsinki, Finland.

#### References

- Ahola AJ, Yli-Knuuttila H, Suomalainen T *et al* (2002). Shortterm consumption of probiotic-containing cheese and its effect on dental caries risk factors. *Arch Oral Biol* **47**: 799–804.
- Aihara K, Kajimoto O, Hirata H, Takahashi R, Nakamura Y (2005). Effect of powdered fermented milk with *Lactobacillus helveticus* on subjects with high-normal blood pressure or mild hypertension. J Am Coll Nutr 24: 257–265.
- Anderson MH, Shi W (2006). A probiotic approach to caries management. *Pediatr Dent* 28: 151–153.
- Banchereau J, Steinman RM (1998). Dendritic cells and the control of immunity. *Nature* **392:** 245–252.
- Bergonzelli GE, Blum S, Brussow H, Corthesy-Theulaz I (2005). Probiotics as a treatment strategy for gastrointestinal diseases? *Digestion* 72: 57–68.
- Boriello SP, Hammes WP, Holzapfel W *et al* (2003). Safety of probiotics that contain lactobacilli or bifidobacteria. *Clin Infect Dis* **36**: 775–780.
- Boris S, Suarez JE, Barbes C (1997). Characterization of the aggregation promoting factor from *Lacobacillus gasseri*, a vaginal isolate. *J Appl Microbiol* **83:** 413–420.
- Brzozowski T, Konturek PC, Mierzwa M *et al* (2006). Effect of probiotics and triple eradication therapy on the cyclooxygenase (COX)-2 expression, apoptosis, and functional gastric mucosal impairment in *Helicobacter pylori*-infected Mongolian gerbils. *Helicobacter* **11**: 10–20.
- Burton JO, Chilcott CN, Tagg JR (2005). The rationale and potential for the reduction of oral malodour using *Streptococcus salivarius* probiotics. *Oral Dis* **11**(Suppl. I): 29–31.
- Burton JP, Wescombe PA, Moore CJ, Chilcott CN, Tagg JR (2006a). Safety assessment of the oral cavity probiotic *Streptococcus salivarius* K12. *Appl Environ Microbiol* **72**: 3050–3053.
- Burton JP, Chilcott CN, Moore CJ, Speiser G, Tagg JR (2006b). A preliminary study of the effect of probiotic Streptococcus salivarius K12 on oral malodour parameters. *J Appl Microbiol* **100**: 754–764.
- Busscher HJ, Mulder AF, van der Mei CH (1999). In vitro adhesion to enamel and in vivo colonization of tooth surfaces by lactobacilli from a bio-yogurt. *Caries Res* 33: 403–404.
- Çaglar E, Kargul B, Tanboga I (2005a). Bacteriotherapy and probiotics' role on oral health. *Oral Dis* **11:** 131–137.
- Çaglar E, Sandalli N, Twetman S, Kavaloglu S, Ergeneli S, Selvi S (2005b). Effect of yogurt with *Bifidobacterium* DN-173 010 on salivary mutans streptococci and lactobacilli in young adults. *Acta Odontol Scand* 63: 317–320.
- Çaglar E, Cilder SK, Ergeneli S, Sandalli N, Twetman S (2006). Salivary mutans streptococci and lactobacilli levels after ingestion of the probiotic bacterium *Lactobacillus reuteri* ATCC 55739 by straws or tablets. *Acta Odontol Scand* **64**: 314–318.
- Cannon JP, Lee TA, Bolanos JT, Danzinger LH (2005). Pathogenic relevance of *Lactobacillus*: a retrospective review of over 200 cases. *Eur J Clin Microbiol Infect Dis* **24**: 31–40.
- Christensen HR, Larsen CN, Kaestel P et al (2006). Immunomodulating potential of supplementation with probiotics: a dose-response study in healthy young adults. *FEMS Immunol Med Microbiol* **47:** 380–390.
- Colloca ME, Ahumada MC, Lopez ME, Nader-Macias ME (2000). Surface properties of lactobacilli isolated from healthy subjects. *Oral Dis* **6**: 227–233.
- Commane D, Hughes R, Shortt C, Rowland I (2005). The potential mechanisms involved in the anti-carcinogenic action of probiotics. *Mutat Res* **11**: 276–289.

- Elahi S, Pang G, Clancy A, Clancy R (2005). Enhanced clearance of *Candida albicans* from the oral cavities of mice following oral administration of *Lactobacillus acidophilus*. *Clin Exp Immunol* **141**: 29–36.
- El-Nezami HS, Polychronaki NN, Ma J *et al* (2006). Probiotic supplementation reduces a biomarker for increased risk of liver cancer in young men from Southern China. *Am J Clin Nutr* **83**: 1199–1203.
- Falagas ME, Betsi GI, Tokas T, Athanasiou S (2006). Probiotics for prevention of recurrent urinary tract infections in women: a review of the evidence from microbiological and clinical studies. *Drugs* 66: 1253–1261.
- Grajek W, Olejnik A, Sip A (2005). Probiotics, prebiotics and antioxidants as functional foods. *Acta Biochim Pol* **52**: 665–671.
- Grangette C, Müller-Alouf H, Goudercourt D, Geoffroy M-C, Turneer M, Mereenier A (2001). Mucosal immune responses and protection against tetanus toxin after intranasal immunization with recombinant *L. plantarum*. *Infect Immun* 69: 1547–1553.
- Guarner F, Perdigon G, Coerthier G, Salminen S, Koletzko B, Morelli L (2005). Should yoghurt cultures be considered probiotic? *Br J Nutr* **93**: 783–786.
- Hart AL, Lammers K, Brigidi P *et al* (2004). Modulation of human dendritic cell phenotype and function by probiotic bacteria. *Gut* **53**: 1602–1609.
- Hatakka K, Ahola AJ, Yli-Knuuttila H, Richardson M, Poussa T, Meurman JK (2007). Probiotics reduce the prevalence of oral Candida in the elderly – a randomized controlled trial. *J Dent Res* **86**: 125–130.
- Haukioja A, Yli-Knuuttila H, Liomaranta V *et al* (2006a). Oral adhesion and survival of probiotic and other lactobacilli and bifidobacteria in vitro. *Oral Micorbiol Immunol* **21**: 326–332.
- Haukioja A, Loimarant V, Tenovuo J (2006b). Effect of lactobacilli and bifidobacterium on streptococcal adherence. IADR Congress, Dublin, 13–16 September (Abstract 0702).
- Husni RN, Gordon SM, Washington JA, Longworth DL (1997). Lactobacillus bacteremia and endocarditis review of 45 cases. *Clin Infect Dis* **25**: 1048–1055.
- Hutt P, Shchepetova J, Loivukene K, Kullisaar T, Mikelsaar M (2006). Antagonistic activity of probiotic lactobacilli and bifidobacteria against entero- and uropathogens. *J Appl Microbiol* **100**: 1324–1332.
- Huys G, D'Haene K, Swings J (2006). Genetic basis of tetracycline and minocycline resistance in potentially probiotic *Lactobacillus plantarum* strain CCUG 43738. *Antimicrob Agents Chemother* **50**: 1550–1551.
- Isolauri E, Sutas Y, Kankaanpaa P, Arvilommi H, Salminen S (2001). Probiotics: effects on immunity. *Am J Clin Nutr* 73(Suppl. 2): S444–S450.
- Jauhiainen T, Vapaatalo H, Poussa T, Kyronpalo S, Rasmussen M, Korpela R (2005). *Lactobacillus helveticus* fermented milk lowers blood pressure in hypertensive subjects in 24-h ambulatory blood pressure measurement. *Am J Hypertens* 18: 1600–1605.
- Kang MS, Na HS, Oh LS (2005). Coaggregation ability of *Weissella cibaria* isolates with *Fusobacterium nucleatum* and their adhesiveness to epithelial cells. *FEMS Microbiol Lett* **253**: 323–329.
- Kang M-S, Kim B-G, Lee H-C, Oh J-S (2006). Inhibitory effect of *Weissella cibaria* isolates on the production of volatile sulphur compounds. J Clin Periodontol 33: 226–232.
- Kazor CE, Mitchell PM, Lee AM *et al* (2003). Diversity of bacterial populations on the tongue dorsa of patients with halitosis and healthy patients. *J Clin Microbiol* **41**: 558–563.

- Kolenbrander PE (2000). Oral microbial communities: biofilms, interactions, and genetic systems. *Annu Rev Microbiol* **54**: 413–437.
- Kõll-Klais P, Mändar R, Leibur E, Marcotte H, Hammarström L, Mikelsaar M (2005). Oral lactobacilli in chronic periodontitis and periodontal health: species composition and antimicrobial activity. *Oral Microbiol Immunol* 20: 354–361.
- Kõll-Klais P, Mändar R, Leibur E, Marcotte H, Hammarström L, Mikelsaar M (2006). Oral lactobacilli in chronic periodontitis: species composition and antimicrobial activity. IADR Congress, Dublin, 13–16 September (Abstract 0081).
- Krasse P, Carlsson B, Dahl C, Paulsson A, Nilsson A, Sinkiewicz G (2006). Decreased gum bleeding and reduced gingivitis by the probiotic *Lactobacillus reuteri*. Swed Dent J **30**: 55–60.
- Lester CH, Frimodt-Moller N, Sorensen TL, Monnet DL, Hammerun AM (2006). In vivo transfer of the vanA resistance gene from an *Enterococcus faecium* isolate of animal origin to an *E. faecium* isolate of human origin in the intestines of human volunteers. *Antimicrob Agents Chemother* **50**: 596–599.
- Lewis SJ, Freedman AR (1998). The use of biotherapeutic agents in the prevention and treatment of gastrointestinal disease. *Aliment Pharmacol Ther* **12:** 807–822.
- Lima LM, Motisuki C, Spolidorio DM, Santos-Pintol L (2005). *In vitro* evaluation of probiotics microorganisms adhesion to an artificial caries model. *Eur J Clin Nutr* **59**: 884–886.
- Marsh P, Martin MV (1999). Oral Microbiology. 4th edn. Wright: Oxford.
- Masco L, Van Hoorde K, De Brandt E, Swings J, Huys G (2006). Antimicrobial susceptibility of *Bifidobacterium* strains from humans, animals and probiotic products. *J Antimicrob Chemother* **58**: 85–94.
- Mego M, Majek J, Koncekova R *et al* (2005). Intramucosal bacteria in colon cancer and their elimination by probiotic strain *Enterococcus faecium* M-74 with organic selenium. *Folia Microbiol (Praha)* **50:** 443–7.
- Meurman JH (2005). Probiotics: do they have a role in oral medicine and dentistry? *Eur J Oral Sci* **113**: 188–196.
- Meurman JH, Antila H, Salminen S (1994). Recovery of *Lactobacillus* strain GG (ATCC 53103) from saliva of healthy volunteers after consumption of yoghurt prepared with the bacterium. *Microb Ecol Health Dis* **7**: 295–298.
- Montalto M, Vastola M, Marigo L *et al* (2004). Probiotics treatment increases salivary counts of lactobacilli: a double-blind, randomized, controlled study. *Digestion* **69**: 53–56.
- Näse L, Hatakka K, Savilahti E *et al* (2001). Effect of longterm consumption of a probiotic bacterium, *Lactobacillus rhamnosus* GG, in milk on dental caries and caries risk children. *Caries Res* **35**: 412–420.
- Nikawa H, Makihira S, Fukushima H, Nishimura H, Ozaki Y, Ishida K (2004). *Lactobacillus reuteri* in bovine milk fermented decreases the oral carriage of mutans streptococci. *Int J Food Microbiol* **95:** 219–223.
- Olivares M, Diaz-Ropero MP, Martin R, Rodriguez JM, Xaus J (2006). Antimicrobial potential of four *Lactobacillus* strains isolated from breast milk. *J Appl Microbiol* **101**: 72–79.
- Oliveira ML, Areas AP, Campos IB *et al* (2006). Induction of systemic and mucosal immune response and decrease in Streptococcus pneumoniae colonization by nasal inoculation of mice with recombinant lactic acid bacteria expressing pneumococcal surface antigen A. *Microbes Infect* **8**: 1016–1024.

450

- Ostengo MC, Nader-Macias EM (2004). Hydroxylapatite beads as an experimental model to study adhesion of lactic acid bacteria from the oral cavity to hard tissues. *Methods Mol Biol* **268**: 447–452.
- Paster BJ, Boches SK, Galvin JL *et al* (2001). Bacterial diversity in human subgingival plaque. *J Bacteriol* **183**: 3770–3783.
- Reid G, McGroarty JA, Angotti R, Cook RL (1988). Lactobacillus inhibitor production against Escherichia coli and coaggregation ability with uropathogens. Can J Microbiol 34: 344–351.
- Reid G, Bruce AW, Fraser N, Heinemann C, Owen J, Henning B (2001). Oral probiotics can resolve urogenital infections. *FEMS Immunol Med Microbiol* **30**: 49–52.
- Rinne M, Kalliomaki M, Arvilommi H, Salminen S, Isolauri E (2005). Effect of probiotics and breastfeeding on the bifidobacterium and lactobacillus/enterococcus microbiota and humoral immune responses. *J Pediatr* **147**: 186–191.
- Salminen M (2006). Lactobacillus bacteremia, with special focus on the safety of probiotic Lactobacillus rhamnosus GG. Dissertation Thesis, University of Helsinki: Helsinki.
- Salminen MK, Tynkkynen S, Hilpi R et al (2002). Lactobacillus bacteremia during a rapid increase in probiotic use of Lactobacillus rhamnosus GG in Finland. Clin Infect Dis 35: 1155–1156.

- Silva M, Jacobus NV, Deneke C, Gorbach SL (1987). Antimicrobial substance from a human lactobacillus strain. *Antimicrob Agents Chemother* **31**: 1231–1233.
- Teanpaisan R, Dahlen G (2006). Use of polymerase chain reaction techniques and sodium dodecyl sulphate-polyacrylamide get electrophoresis for differentiation of oral *Lactobacillus* species. *Oral Microbiol Immunol* **21**: 79–83.
- Volozhin AI, Il'in VK, Maksimovskii IM *et al* (2004). Development and use of periodontal dressing of collagen and *Lactobacillus casei* 37 cell suspension in combined treatment of periodontal disease of inflammatory origin (a microbiological study). *Stomatologiia* (Mosk) 83: 6–8.
- Wolf BW, Wheeler D, Ataya DG, Garleb KA (1998). Safety and tolerance of *Lactobacillus reuteri* supplementation to a population infected with the human immunodeficiency virus. *Food Chem Toxicol* **36**: 1085–1094.
- Wu H-Y, Nguyen HH, Russell MW (1997). Nasal lymphoid tissue (NALT) as a mucosal immune inductive site. *Scand J Immunol* 46: 506–513.
- Yli-Knuuttila H, Snäll J, Kari K, Meurman JH (2006). Colonization of *Lactobacillus rhamnosus* GG in the oral cavity. *Oral Microbiol Immunol* 21: 129–131.

Copyright of Oral Diseases is the property of Blackwell Publishing Limited and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.