http://www.blackwellmunksgaard.com

REVIEW ARTICLE

Oral squamous cell carcinoma: overview of current understanding of aetiopathogenesis and clinical implications

C Scully¹, JV Bagan²

¹University College London, London, UK; ²Valencia University and Hospital General Universitario de Valencia, Valencia, Spain

MEDLINE contains well over 14 000 papers revealed by a search using keywords 'oral squamous cell cancer' or 'oral squamous cell carcinoma', over 27 000 using 'oral carcinoma' and over 48 000 using the keywords 'oral cancer'. It is difficult to see how clinicians could keep abreast of such a subject. This paper attempts to help by providing an overview of the aetiopathogenesis of oral squamous cell carcinoma (OSCC), discussing changes in epidemiology and increasing awareness of the wide range of risk factors, emphasising the genetic background to cancer susceptibility and the genetic changes associated with progression to OSCC and highlighting clinical implications.

Oral Diseases (2009) 15, 388–399

Keywords: oral cancer; tobacco; betel; alcohol; diet; viruses; genetics

Introduction

Most oral cancer is oral squamous cell carcinoma (OSCC) – a disease found particularly in low income communities and mainly a problem of older men, 90% being in the over 45-year-age group who are exposed to the known risk factors of tobacco and/or alcohol (IARC 2004). Clinically, OSCC includes lip cancer, which accounts for the majority of OSCC and intra-oral cancer, which mainly affects the tongue (Table 1).

Oral squamous cell carcinoma is the eighth most common cancer world-wide but parts of Northern France and East Europe, particularly Hungary, and parts of South America and South East Asia have particularly high prevalences (Moore *et al*, 1999, 2000a,b).

Lip cancer is particularly a problem of older people and is especially a disease of males and particularly a lesion of the lower lip, which is largely related to the

Received 9 March 2009; accepted 10 March 2009

exposure to ultra violet irradiation from the sun. It is particularly seen in people who are exposed on a prolonged, or a repeated basis, to sunlight, and that includes fishermen, farmers, skiers and windsurfers. It is found mainly in Caucasians particularly in hot climates. There is a very good survival – up to 95% 5-year survival – probably related to the early diagnosis of a lesion, which is clinically very obvious (Scully and Moles, 2008).

Cancer within the mouth affects mainly the tongue, particularly the lateral border, especially posteriorly, in older people, in males, and often related to life-style habits, which are largely tobacco- or alcohol-related (Scully and Moles, 2008). It is a particular problem in some black and ethnic minority populations (Scully and Bedi, 2000). With around a 40% 5-year survival, it is quite different from lip cancer, with about half as good a prognosis.

There are changing patterns in both lip and intra-oral cancer. There has been a decrease in male incidence of lip cancer over about a 30-year period, but several studies show an increase in tongue cancer, particularly in younger patients, currently attributed to smoking and binge drinking amongst younger people. In males in East and Central Europe, there has been a rise in mortality rates since the 1980s along with an increase in tobacco use, and the rise is the largest for any of the common neoplasms (Bray et al, 2002; La Vecchia et al, 2004). In Western European males, there has also been a rise, but in some countries where lung cancer has decreased which suggests that it is not in those populations related so much to the classical risk factor of tobacco as to alcohol. In females, there has been a slight increase associated with more alcohol and tobacco use. OSCC remains a lethal disease for over 50% of cases diagnosed annually (Warnakulasuriya, 2008).

Aetiopathogenesis

The aetiopathogenesis of potentially malignant oral disorders and OSCC has been reviewed a number of times this millennium (e.g., Scully *et al*, 2000a,b,c; Patel *et al*, 2001; Reibel, 2003; Brinkman and Wong, 2006; Mithani *et al*, 2007; Haddad and Shin, 2008).

Correspondence: Crispian Scully, University College London, London, UK. Tel: 00442079151170 (1232), E-mail: crispian.scully@eastman.ucl.ac.uk.

Table 1 International Classification of Diseases; oral cancer

	ICD-9	ICD-10
Lip	140	C00
Tongue	141	C01-02
Gum	143	C03
Floor of mouth	144	C04
Other and unspecified mouth	145	C05-06
Salivary gland	142	C07-08
Oro-, naso-, and hypo-pharynx Other and ill-defined sites of lip, oral cavity and pharynx	146–149	C09–14

The cell of origin of OSCC is the oral keratinocyte, in which DNA mutation can be spontaneous, but mutagens increase the mutation rate. Genetic variation in the xenobiotic metabolising enzymes (XME), which influence carcinogen (cancer-causing chemical) metabolism, DNA repair mechanisms and other protective mechanisms may well help explaining differing susceptibilities to the OSCC – causing effects of the risk factors such as tobacco and alcohol (see below, and Scully *et al*, 2000a,b,c).

Risk factors

The various risk factors for OSCC appear mainly to act by increasing the rate of mutations. Lifestyle factors, especially tobacco and alcohol, appear particularly important but, in some cases, betel, sunlight exposure, ionising radiation, human papillomavirus (HPV) or other infections or immuno-incompetence are relevant (Scully and Moles, 2008). Genetics such as single nucleotide polymorphisms (SNPs) may also influence the risks, as discussed below.

Tobacco

Tobacco is by far the main risk factor for OSCC (Vallecillo Capilla et al, 2007; Hirota et al, 2008), and this applies not only to smoked but also to smokeless tobacco (Johnson, 2001; Warnakulasuriya and Ralhan, 2007), although some have suggested a somewhat lesser risk from the latter (Vigneswaran et al, 1995; Boffetta et al. 2008). Tobacco use generates carcinogens such as the TSNAs, tobacco-specific nitrosamines [N'-nitrosonornicotine,4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone] as well as free radicals, resulting in alterations in the antioxidant enzymes glutathione-S-transferase (GST), glutathione reductase, superoxide dismutase, catalase and glutathione peroxidase, as well as lipid peroxidation and total thiol, although tobacco products vary widely in their potential for producing carcinogens (Brunnemann et al, 1996; Hoffmann and Hoffmann, 1997).

There is about a 20-fold risk of OSCC with heavier tobacco smokers and a strong dose–response relationship, and the risk increases with the number of cigarettes smoked per day and the duration of smoking. Tobacco in other forms is also a risk factor: for example, data from 282 incident oral cancer cases and 1410 matched controls in Trivandrum, India analysed using multivariate conditional logistic regression models confirmed tobacco chewing to be the strongest risk factor associated with OSCC. Effects of chewing pan (betel) with or without tobacco on OSCC risk were elevated for both genders. Bidi smoking increased the risk of OSCC in men. Dose–response relations were observed for the frequency and duration of tobacco chewing and alcohol drinking, as well as in duration of bidi smoking (Muwonge *et al*, 2008).

Betel

There is also a relationship of OSCC with betel (areca nut) use, a habit of something like 20% of the world's population (Cogliano et al, 2004). The IARC long ago declared that betel was carcinogenic to humans, and that has been confirmed (Merchant et al, 2000; Jacob et al, 2004; Carpenter et al, 2005; Guh et al, 2007; Thomas et al, 2007, 2008; Reichart and Nguyen, 2008). Gene expression may be altered and arecoline, a main alkaloid in the areca nut (the main component of betel quid) may, by hypermethylation, block tumour suppressor genes (TSGs) p14, p15 and p16, and it inhibits the p53 TSG, represses DNA repair and triggers DNA damage responses in human epithelial cells (Chen et al., 2008; Park et al, 2008; Takeshima et al, 2008; Tsai et al, 2008; Cheong et al, 2009). Similar chewing habits - such as khat use – may also be implicated in OSCC in some communities (Fasanmade et al, 2007; Sawair et al, 2007). The carcinogenicity of products such as marijhuana is more controversial in the aetiopathogenesis of OSCC (Hashibe et al, 2005).

Alcohol

Alcohol (ethanol) is oxidised to acetaldehyde – a suspected carcinogen – (Boccia *et al*, 2009a) by alcohol dehydrogenases (ADHs), and acetaldehyde is then degraded to acetate (non-carcinogenic) by aldehyde dehydrogenases (ALDH). Alcohol produces about a fivefold risk for heavy drinkers, and there is a strong dose–response relationship between alcohol drinking and alcohol consumption (Pelucchi *et al*, 2008). Use of alcohol-containing mouthwashes is more controversial in the aetiopathogenesis of OSCC (McCullough and Farah, 2008; La Vecchia, 2009).

There is a greater than joint multiplicative risk for OSCC in people who are both alcohol drinkers and heavy tobacco smokers (Hashibe *et al*, 2009). Smoking increases the acetaldehyde burden following alcohol consumption, and alcohol-drinking enhances the activation of pro-carcinogens present in tobacco due to increased metabolic activation, by an induced cytochrome P450-2E1-dependent microsomal biotransformation system in the mucosa and the liver (Seitz and Cho, 2009).

Other factors

In one study of OSCC in young adults, tobacco accounted for 77%, alcohol for 52%, low vegetable consumption for 52% and all three combined for 85% of OSCC (Rodriguez *et al*, 2004). Worldwide, 25% oral cancers are attributable to tobacco usage (smoking

and/or chewing), 7–19% to alcohol drinking, 10–15% to micronutrient deficiency and more than 50% to betel quid chewing in areas of high chewing prevalence (Petti, 2008). These behaviours are widespread: 2 billion consume alcohol; 1 billion men and 250 million women smoke cigarettes; 600–1200 million people chew betel quid and an unbalanced diet is common amongst both developing and developed countries (Petti, 2008). However, a substantial proportion of OSCC cannot be attributed to tobacco or alcohol use, particularly among women and among young-onset cases (Llewellyn *et al*, 2001; Hashibe *et al*, 2009) and, in these, other factors, such as micro-organisms may thus be at play.

Viruses

There is a relationship with HPV and OSCC, particularly oropharyngeal carcinoma (D'Souza et al, 2007). A number of studies have shown DNA from HPV in OSCC, especially in oropharyngeal carcinoma. OSCC is also increased in people who have a high number of sexual partners and if they start intercourse at a young age. OSCC is increased in females who have cervical carcinoma which is related itself to HPV, and it is also increased in their partners. The possibility of sexual transmission has thus been raised (Scully, 2002, 2005). An interesting anecdotal report showed a couplehusband and wife diagnosed synchronously with OSCC: their tumours were positive for HPV16 by polymerase chain reaction (PCR) and both viral genomes were identical and closely related to the revised European prototype, HPV16R (Haddad et al, 2008). These tumours probably represented transmission between the couple (Haddad et al, 2008). Some tumours are associated with HPV and some with viruses of the herpes family, although the exact role of these viruses needs careful evaluation (Shillitoe, 2008).

Oral health

The dentition may also play a role in OSCC (Zheng et al, 1990). Head and neck and oesophageal dysplasia/cancer has been found associated with dental neglect (Dye et al, 2007; Guha et al, 2007; Abnet et al, 2008), and OSCC is less likely in people who have received dental care (Holmes et al, 2008). Periodontal disease or tooth loss has been implicated in OSCC (Meyer et al, 2008), each millimetre of alveolar bone loss being associated with a 5.23-fold increase in the risk of tongue cancer (Tezal et al, 2007). Possible mechanisms revolve around microbial interactions with dental bacterial plaque: polymicrobial supragingival plaque has a mutagenic interaction with saliva (Bloching et al, 2007), and both oral streptococci (Kurkivuori et al, 2007) and neisseria (Muto et al, 2000) may synthesise acetaldehyde from alcohol.

Host defences

Other factors implicated in OSCC may act by undermining host defences against carcinogens, or repair or defence mechanisms. These include genetic, immune and dietary defects, drugs and deprivation. Conditions associated with an increased risk of OSCC include organ transplantation, Fanconi anaemia, dyskeratosis congenita and more recently diabetes and scleroderma (Dikshit *et al*, 2006; Goutzanis *et al*, 2007). Factors that are more controversial include HIV/AIDS and various inherited cancer syndromes (e.g., Li-Fraumeni syndrome).

Diet

Diet may play a role in OSCC development as evidenced by multiple epidemiological studies worldwide (Pavia et al, 2006). Apart from the dietary risk factors for OSCC such as alcohol and other factors (Zain, 2001), the most consistent dietary findings across multiple cultural settings are a protective effect of high fruit consumption (Winn, 1995) and high vegetable consumption (Lucenteforte et al, 2008), especially yellow/orange vegetables (Sapkota et al, 2008), and diets varied in vegetables and fruit (Garavello et al, 2008). Dietary antioxidants from fruit and vegetables may be protective (Suzuki *et al*, 2006). Folate in particular may be protective (Pelucchi et al, 2003) whilst, in contrast, mild iron deficiency and low glutathione (GSH) levels which are associated with increased oxidative stress appear to increase the risk of OSCC (Richie et al, 2008). Although fruits and vegetables can be protective against OSCC (Pavia et al, 2006), probably via folate and dietary antioxidants (carotene, carotenoids, flavonoids, flavanones, vitamins A, C and E and phytosterols) (Rossi et al, 2007), preserved vegetables can be a risk factor (Sapkota et al, 2008). Even in the presence of high alcohol consumption or tobacco use, a high intake of fruit and vegetables might prevent the development of around one quarter of cases of SCC in the head and neck (Boccia et al, 2008) and possibly one half OSCC (Pavia et al, 2006). The risk of oral potentially malignant disorders is also significantly reduced with higher consumption of fruits, particularly citrus fruits and juices (Maserejian et al, 2006). Case-control studies indicate that vitamins C, E, A and carotenoids in food may decrease the risk of oral potentially malignant disorders and OSCC, but clinical trials of vitamin supplements have failed to find protective effects of beta-carotene and suggest indeed, that beta carotene and vitamin E may increase the risk (Maserejian et al, 2007). This is discussed further, elsewhere (Scully, 1995; Scheer et al, 2004; Brown and Kane, 2006).

Potentially malignant oral disorders is the term adopted at a recent WHO workshop to cover the most common ones (leukoplakia and erythroplakia) that may transform to OSCC, and others such as lichen planus, oral submucous fibrosis, actinic cheilitis (van der Waal, 2008).

Carcinogenesis

DNA mutations affect a number of genes, disrupting growth control. Multiple genetic and epigenetic events include the aberrant expression and function of molecules regulating cell signalling, growth, survival, motility, angiogenesis and cell cycle control, and underlie the progressive acquisition of a malignant phenotype by the keratinocyte which progresses via a series of genetic change steps to a premalignant or a potentially malignant cell – characterised by an ability to proliferate in an uncontrolled fashion – that may result in cancer, characterised by invasion across the epithelial basement membrane and eventual metastasis. Changes in over one hundred genes have now been implicated (Roepman *et al*, 2005). The cell cycle is disturbed particularly by various oncogenes and their over-expression or over-activity (amplification), which will drive cell proliferation.

Working in the opposite more protective direction are the TSGs: for example, the more important TSGs are p16 which acts as a checkpoint in growth control and p53, which will either repair a potentially malignant cell or it will destroy it by apoptosis (see below). Protective mechanisms include TSGs and the liver – protective particularly because of its ability to metabolise carcinogens. The carcinogen metabolising enzymes vary from person to person on a genetic basis. There are also a whole series DNA repair enzymes, which can repair the mutations and, again, there are differences genetically between individuals.

The fundamental and simplified concept of the genetic basis behind cancer is the over-expression of oncogenes and/or the silencing of TSGs. Genetic analysis now involves a detailed high-resolution mapping of regions of chromosomal gain, loss and translocation using techniques such as comparative genomic hybridization and fluorescent in-situ hybridization. PCR-based techniques can identify if there is loss of genetic material, represented by complete deletion, or loss, of one allele (known as loss of heterozygosity or LOH), and it is evident that there is a relatively common pattern of DNA allelic loss as one progress from the premalignant to malignant stage. Presumably, if a TSG is in the area of allelic loss, this would make the host more susceptible to dysfunction of the gene, leading to the development of cancer. The common regions of chromosomal loss reported in OSCC prove to be at 1p, 3p, 4p, 5q, 8p, 10p, 11q, 13q and 18q, with gains at 1q, 3q, 5p, 7q, 8q, 9q, 11q, 12p, 14q and 15q. Microarray technology involves the miniaturisation of DNA sequence hybridization onto microscopic surfaces, which can then be read by laser, able to detect and interpret signals from minute fluorophores. Microarray technology has facilitated the ability to produce huge amounts of data from nearly the entire human genome, and such DNA and messenger RNA (mRNA) arrays are proving powerful in identifying key elements involved in OSCC. For example, one microarray-based gene-expression analysis found 601 genes to be significantly regulated in OSCC tissue compared to adjacent intra-individual mucosa controls, and 25 genes with differences in their regulation comparing samples from early-stage cancer with those from advanced disease (Fialka et al, 2008). Some of the mechanisms that might be implicated in

carcinogenesis are summarised elsewhere (Haddad and Shin, 2008; Bagan and Scully, 2009).

391

Tumour suppressor genes

Tumour suppressor genes are genes that normally function in growth control – by regulating the cell cycle, apoptosis, cell adhesion and DNA repair. The normal cell cycle includes programmed cell death, or apoptosis, and cells that avoid this have the potential to become cancerous. Aberrations such as deletions or mutations in TSG genes, or silencing by hypermethylation, can thus lead to cancer.

There are several putative TSGs involved in OSCC, but the most important at present appear to be p53 (*TP53*) and p16 (Angiero *et al*, 2008). *TP53*, a gene involved in apoptosis and cell cycle regulation, is silenced (most often through point mutations), leading to premalignant disease and OSCC. Furthermore, the *MDM* class of oncogenes has been shown to inhibit *TP53* in the absence of *TP53* mutation. There is also a high incidence of LOH at chromosome 9p21, where the cyclin-dependent kinase *CDKN2a* locus codes for both *p16* and *p14* (*ARF*) – two putative TSGs that respectively regulate the cell cycle and stabilise *TP53* via *MDM2*. These TSGs are probably silenced by epigenetic control, such as promoter hypermethylation (Mithani *et al*, 2007).

There could be a myriad of other TSGs silenced in OSCC, but the pattern of silencing may differ between patients.

Oncogenes

The over-expression of oncogenes – genes that promote growth, survival and spread of cells – can lead to the development of cancer. However, the identification of oncogene over-expression is not easy. Most studied has been the epidermal growth factor receptor (*EGFR*) gene, which appears to be over-expressed in most OSCC. For example, EGFR is amplified in areca-associated OSCC compared with matched adjacent oral mucosa (Chiang *et al*, 2008).

The $\Delta Np63-\alpha$ oncogene is also over-expressed and may inhibit p73, a TSG related to TP53, which is also involved in the bcl-2 mediated apoptotic pathway, and $\Delta Np63-\alpha$ may cause the accumulation of beta-catenin, a transcription factor that may play a role in cell adhesion and in the Wnt signalling pathway. Matrix metalloproteinases (MMPs) are a family of genes thought to be involved in cell adhesion, proliferation and migration, and MMPs may act as oncogenes related to infiltrative growth and lymph node involvement. Other oncogenes involved in OSCC may include 11q13, which probably influences the Fas-associated death domain. As with TSGs, the range of oncogenes being identified is quite broad, and the mechanisms by which they act appear complex (Glazer et al, 2008; Ha et al, 2008).

Mitochondrial mutations

Mitochondria play a crucial role in cellular respiration and ATP generation, and appear to have a role in carcinogenesis, possibly via the accumulation of succinate (part of the tricarboxylic acid cycle), which inhibits the degradation of hypoxia-inducible factor (*HIF*)-1 alpha, an angiogenic factor activated by tumour hypoxia (Ha *et al*, 2008; Sun *et al*, 2009).

Stromal cell derived factor (SDF-1) and chemokine (C-X-C motif) receptor (CXCR4) may be involved in invasion and metastasis via angiogenic factors such as vascular endothelial growth factors (VEGFs) (Carmeliet and Jain, 2000). Some of the range of mechanisms that might be implicated in carcinogenesis and recently reported are summarised elsewhere (Bagan and Scully, 2009).

Single nucleotide polymorphisms

Single nucleotide polymorphisms are genome areas that have altered DNA sequences which may not lead to an amino acid alteration, or altered DNA sequences that do not seem to have any adverse effect in 'normal' individuals but may be markers for disease predisposition, or may be used to genetically identify patients, as they tend to cluster with ethnic background. SNPs in TSGs may play a role in cancer development (Drummond *et al*, 2002; Izzo *et al*, 2003): for example, SNPs in cell-cycle control pathway genes such as the CCND1 splice variant P241P may contribute to the risk of potentially malignant lesions (Ye *et al*, 2008).

Single nucleotide polymorphisms have also been detected in enzymes responsible for detoxifying environmental toxins and carcinogens (XMEs) and for DNA repair (DNA repair enzymes), and some may be markers for OSCC development (Duarte *et al*, 2008). However, this has not been confirmed in all studies (Losi-Guembarovski *et al*, 2008).

Xenometabolising enzymes

Xenometabolising enzymes have been fairly extensively studied. Alcohol (ethanol) is oxidised to acetaldehyde (the suspected carcinogenic agent in alcohol) by ADHs and cytochrome P-4502E1 (CYP2E1), both of which exhibit great inter-individual variability in activity. Individuals with a high production rate of acetaldehyde from ethanol have an increased cancer risk when they drink chronically. These include individuals with a genetically determined increased acetaldehyde production caused by ADH polymorphism (Yokoyama *et al*, 2007a, 2007b), as well as those with a decreased detoxification of acetaldehyde to acetate due to ALDH mutation.

Alcohol dehydrogenase-3 (now ADH1C) (ADH3*2 allele) may predispose to OSCC (Bouchardy *et al*, 2000; Schwartz *et al*, 2001; Zavras *et al*, 2002). ADH-2 (now ADH1B) homozygotes avoid alcohol because of adverse reactions (and are at low cancer risk), but heterozygotes (who can drink alcohol) are liable to OSCC (Seitz and Stickel, 2007). ADH-1B and ADH-7 are independently and strongly associated with protection against OSCC and aero-digestive cancers (Hashibe *et al*, 2008).

Aldehyde dehydrogenases genotypes may influence cancer susceptibility, presumably as acetaldehyde can accumulate. Some head and neck carcinomas appear to be ALDH2-associated, possibly by increases in acetaldehyde-derived DNA adducts (Yokoyama and Omori, 2003; Matsuda *et al*, 2006; Asakage *et al*, 2007; Yokoyama *et al*, 2008). The less-active homozygous ADH-1B (ADH1B*1/*1) and inactive heterozygous ALDH-2 (ALDH2*1/*2) increase the risk of upper aero-digestive tract cancer in Japanese alcoholics (Yokoyama *et al*, 2007a,b).

Alcohol dehydrogenase and methylenetetrahydrofolate reductase (MTHFR) appear to influence cancer risk as the ADH1C*2/*2/MTHFR 677TT genotype combination appears to be more susceptible to OSCC, with a 20-fold increase in risk in heavy drinkers and a 5.9- and 2.8-fold increase in risk, respectively, in moderate drinkers and light drinkers (Solomon *et al*, 2008). Others have suggested that MTHFR SNPs (C677T and A1298C) influence head and neck cancer (Boccia *et al*, 2009b), further implicating folate as a protective factor.

N-acetyl transferases (NATs) catalyse both the *N*-acetylation (usually deactivation) and *O*-acetylation (usually activation) of aromatic and heterocyclic amine carcinogens. Epidemiological studies suggest that the isozymes *NAT1* and *NAT2* polymorphisms via acetylation modify the risk of developing head and neck, and some other, cancers. Fast NAT2 acetylation appears to be a risk factor for OSCC (Buch *et al*, 2008). Ethnic differences in *NAT1* and *NAT2* genotype frequencies may also be a factor in cancer susceptibility (Hein *et al*, 2000).

The enzyme, GST, catalyses conjugation of reduced glutathione via the sulphydryl group, activity which is useful in the detoxification of endogenous compounds such as peroxidised lipids. GSTM1 (glutathione S-transferase mu 1) null genotype significantly increases susceptibility to OSCC in Asians (Patel et al, 2008; Zhuo et al, 2009) but not in Caucasians (Zhuo et al, 2009). Studies on OSCC susceptibility of genetic polymorphisms at GSTM1, GSTT1 and GSTP1 gene loci have not generally supported the hypothesis of an increased risk of GSTP1 G/G, GSTM1 or GSTT1 null genotypes for developing OSCC: in contrast, the GSTM1 A/B genotype appears protective (Hatagima et al, 2008). The isolated or combined null genotype of GSTM1 and GSTT1 appear associated with oral leukoplakia development, and the null GSTT1 genotype shows an increased risk of p53 over-expression, in oral leukoplakia (Duarte et al, 2008). There may be joint effect for GSTM1 homozygous deletion and the CYP1A1 m1m2 variant (Varela-Lema et al, 2008). Polymorphisms at mitochondrial (mt) loci alone and in combination with the risk genotype at GSTP1 also increase the risk of OSCC (Datta et al, 2007).

DNA repair genes

Functional DNA repair genes are essential to protection against carcinogenesis, and SNPs may influence development of potentially malignant oral disorders and OSCC. Nucleotide excision repair enzymes *xeroderma pigmentosum complementation group A* (XPA [A23G]), XPC (Ala499Val) and XPD (Asp312Asn) appear to predict risk of developing potentially malignant oral disorders (Wang *et al*, 2007). Study of ERCC6 (excision

392

repair cross-complementing rodent repair deficiency complementation group 6) suggests that the heterozygous and homozygous A allele of the ERCC6 codon 399 may be associated with OSCC (Chiu *et al*, 2008a,b). However, gene–environment interactions with tobacco smoking and betel quid chewing, but not alcohol drinking, were significant (Chiu *et al*, 2008a,b).

Study of the DNA double strand break repair gene XRCC4 (X-ray repair complementing defective repair in Chinese hamster cells 4) showed that patients with heterozygous del/ns at XRCC4 intron three had an increased risk of OSCC compared to those with ins/ins, and there were significant gene–environment interactions with tobacco smoking and betel quid chewing, but not with alcoholism (Chiu *et al*, 2008a,b). Interactions of various of these SNPs may increase the risk of potentially malignant oral disorders and OSCC (Marques *et al*, 2006; Majumder *et al*, 2007).

Other genes

Polymorphisms affecting gene expression of cytokines such as interleukins IL-4, -6, -8, -10 as well as tumour necrosis factor-alpha (TNF- α) appear to be strongly associated with an increased risk for OSCC (Serefoglou *et al*, 2008). Smokeless tobacco has been shown to induce TNF- α , which, along with its receptors, is over-expressed in OSCC. SNPs in TNF- α and TNF receptor (TNFR) genes may affect their expression and may be a potential determinant of susceptibility to tobacco-related OSCC: TNF- α –308G/A may be related to susceptibility, whereas SNPs involving –609TT TNFR1 and 1690 C/T TNFR2 may be protective (Gupta *et al*, 2008).

The inducible cyclo-oxygenase (COX)-2 enzyme, which plays an important role in inflammation, but also in carcinogenesis, is over-expressed in oral potentially malignant disorders and OSCC, and SNPs in the COX-2 gene may modify the risk of developing potentially malignant disorders (Pu *et al*, 2009) and the risk of OSCC, where COX-2–765G > C polymorphisms: COX-2–765C allele *vs* –765G/G genotype may be protective (Lin *et al*, 2008).

Clinical implications

Clinical diagnostic aids

A variety of commercial diagnostic aids and adjunctive techniques are on the market to potentially assist in the screening of patients for evidence of otherwise occult cancerous change or to assess the potential of clinically abnormal lesions, but none has rigorously yet been confirmed to be superior to clinical examination (Lingen *et al*, 2008; Trullenque-Eriksson *et al*, 2009). Toluidine blue staining has been shown to be related to the genetic changes (allelic loss or LOH) associated with the progression of potentially malignant lesions to OSCC (Guo *et al*, 2001; Zhang *et al*, 2005). Furthermore, a longitudinal study showed that toluidine blue identified LOH-positive lesions that progressed to OSCC (Zhang *et al*, 2005).

There have been enumerable studies on markers that might be of diagnostic or prognostic value.

Genetic characteristics such as described above can be found in OSCC tissue and offer the potential for use in diagnosis and prognostication, especially in the light of the limitations in histopathology discussed elsewhere (Abbey *et al*, 1995; Karabulut *et al*, 1995; Fischer *et al*, 2004, 2005).

Genetic changes are found in the oral mucosa in tobacco users and, in those with OSCC, are also found outwith the tumour (Boldrup et al, 2005; Gabriel et al, 2006; Proia et al, 2006; Sanz-Ortega et al, 2007; Sridhar et al, 2008). This is no surprise since, over half a century ago, Slaughter showed that 11% of patients with OSCC had another cancer elsewhere outside the primary OSCC, introducing the concept of field-cancerisation (Slaughter et al, 1953). Others have confirmed that histologically dysplastic epithelium may be found far removed from sites of OSCC (Thomson, 2002; Thomson and Hamadah, 2007). Furthermore, in patients who smoke tobacco, genetic changes not only affect the whole of the aero-digestive mucosa but also persist for a long period of time, even if the patient stops smoking. Second primary tumours are found in the upper aerodigestive tract in about 20% (Lippman and Hong, 1989).

Further, in many potentially malignant lesions, there can be a mixture of potentially malignant cells, malignant cells that have yet to invade, cells that have invaded and normal cells (Califano et al, 2000; Braakhuis et al, 2003, 2004, 2005a,b; Tabor et al, 2004). Thus, in a clinical potentially malignant lesion, there may be a range of cells present of different malignant potential, including some that traverse the epithelial basement membrane - defining the lesion as cancer. The histopathological interpretation of a biopsy from such a lesion could thus vary from benign to malignant. Thus, in patients with potentially malignant disorders, biopsy may not truly represent the worse components in a lesion. Indeed, as long as 25 or more years, an Italian study showed that leukoplakias, which were nondysplastic upon incisional biopsy, on excision contained OSCC in around 10% (Chiesa et al, 1986). Similar findings were reported more recently (Holmstrup *et al*, 2007).

Genetic markers

Developments in genetic markers may have major implications in clinical practice, some of which are discussed elsewhere (van Houten *et al*, 2000a,b). Examination of tissue for TSG silencing might be of utility. p53 (chromosome 17) expression above basal layer in potentially malignant oral disorders is predictive of cancer (Cruz *et al*, 1998). p53 is mutated and inactivated, with LOH at 17p13, but p53 changes alone appear unsuitable for cancer prediction (Warnakulasuriya, 2000). P53 changes are seen at tumour surgical margins that appeared histologically clear of cancer (Brennan *et al*, 1995; Bilde *et al*, 2009), and this may help predict recurrence (Ball *et al*, 1997). There may also be p16 changes at surgical margins (Bilde *et al*, 2009), and there have been early reports on its utility and that of RARbeta, E-cadherin, cyclin A1 and cytoglobin (Shaw *et al*, 2006, 2007). NF-kappaB and COX-2 have also been studied at tumour surgical margins (Santhi *et al*, 2006). In lymph node biopsies, p53 changes can detect unsuspected nodal metastases (Brennan *et al*, 1995; Tjebbes *et al*, 1999; Cortesina *et al*, 2000).

Telomerase (which is not expressed in normal epithelia) is expressed in potentially malignant oral disorders such as leukoplakia, and in epithelial dysplasia, OSCC and adjacent to carcinomas (Mutirangura *et al*, 1996; Kannan *et al*, 1997; Thongprasom *et al*, 1998; Sumida *et al*, 1999; Kim *et al*, 2001; Luzar *et al*, 2004, 2004; Yajima *et al*, 2004).

Other markers such as those of cell proliferation (Ki-67 antigen) and apoptosis (Bax, Bcl-2) may play a role in diagnosis: apoptotic Bcl-2 expression decreases significantly in dysplastic and early invasive lesions and then increases in consequent stages, while Ki-67 expression increases sharply in early OSCC, but significantly decreases later (Derka *et al*, 2006). A more aggressive tumour behaviour and worse prognosis may be signified by changes in a range of biomarkers – such as reduced E-cadherin expression (Hamidi *et al*, 2000; Diniz-Freitas *et al*, 2006), laminin (LN) γ 2 chain expression (Kuratomi *et al*, 2006) and decreased tumour cell transmembrane proteoglycan syndecan-1 (Máthé *et al*, 2006).

However, the most predictive of the available tissue molecular markers reported thus far and assessed in OSCC development (Lee *et al*, 2000) include chromosomal polysomy, p53 protein expression and LOH in chromosomes 3p or 9p (probably due to changes in p16). It would seem probable that the use of these markers as an adjunct to routine histopathological examination may help prognostication and effective management of the lesions, but sadly, routine use still appears to be low, perhaps hampered by the test cost, complexity, lack of facilities in some laboratories and limited outcome studies to date (Scully *et al*, 2003).

Salivary markers

Cancer-specific variations have also been demonstrated in body fluids. Saliva may contain p53 mutations, microsatellite alterations, increased mitochondrial DNA content and TSG promoter hypermethylation, and the concept of a saliva test to diagnose OSCC is very appealing (Hu *et al*, 2006, 2007, 2008; Wong, 2006; Park *et al*, 2007; Viet *et al*, 2007; Zimmermann *et al*, 2007; Xie *et al*, 2008).

RNAs in saliva have been tested in over 300 saliva samples from OSCC patients and healthy people, and the signature was always present in higher levels in the saliva of OSCC patients than in saliva from healthy people, with an overall accuracy rate of about 85% (Wang *et al*, 2006). CD44, a multistructural and multifunctional cell surface transmembrane glycoprotein molecule is also detectable in saliva. CD44 is involved in cell proliferation, cell differentiation, cell migration, angiogenesis, presentation of cytokines, chemokines and growth factors to the corresponding receptors, and docking of proteases at the cell membrane, as well as in signalling for cell survival. CD44 isoforms containing the variant 3 (v3) exon include a growth factor binding site and may be involved in OSCC progression (Franzmann *et al*, 2001; Reategui *et al*, 2007). Salivary soluble CD44 (solCD44) levels were significantly raised in head and neck cancer patients compared with normal controls, but solCD44 levels did not vary significantly with tumour size, stage, recurrence, history of radiation treatment or tobacco and alcohol risk factors (Franzmann *et al*, 2005).

Free radicals (Bahar *et al*, 2007), ALDH (Giebutowicz *et al*, 2008), endothelin (Pickering *et al*, 2007), Cyfra 21-1, the soluble fragment of cytokeratin 19 (CK19) (Zhong *et al*, 2007), insulin-like growth factor (IGF), MMP-2 and MMP-9 (Shpitzer *et al*, 2007) and p53 (Boyle *et al*, 1994; Tavassoli *et al*, 1998) are among the other substances present in saliva that are being examined for potential diagnostic value.

Serum markers

Serum could possibly also be helpful in identifying common mutations, promoter hypermethylation and LOH (Glazer *et al*, 2008; Molinolo *et al*, 2008), but not all studies have given promising results. For example, p53 antibodies in serum show no correlation with prognosis (Friedrich *et al*, 1997; Maass *et al*, 1997; Tavassoli *et al*, 1998; Gottschlich *et al*, 1999). Multicentre studies in large populations at risk of OSCC and those at low risk are needed.

Summary

Genetic changes can be found in OSCC tissue and body fluids and offer the potential for use in diagnosis and prognostication, especially in the light of the limitations in conventional histopathology.

Author contributions

Drs Scully and Bagan contributed equally in writing this paper.

References

- Abbey LM, Kaugars GE, Gunsolley JC *et al* (1995). Intraexaminer and interexaminer reliability in the diagnosis of oral epithelial dysplasia. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* **80**: 188–191.
- Abnet CC, Kamangar F, Islami F *et al* (2008). Tooth loss and lack of regular oral hygiene are associated with higher risk of esophageal squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev* **17:** 3062–3068.
- Angiero F, Berenzi A, Benetti A *et al* (2008). Expression of p16, p53 and Ki-67 proteins in the progression of epithelial dysplasia of the oral cavity. *Anticancer Res* **28**: 2535–2539.
- Asakage T, Yokoyama A, Haneda T *et al* (2007). Genetic polymorphisms of alcohol and aldehyde dehydrogenases, and drinking, smoking and diet in Japanese men with oral and pharyngeal squamous cell carcinoma. *Carcinogenesis* **28**: 865–874.
- Bagan JV, Scully C (2009). Recent advances in Oral Oncology 2008; squamous cell carcinoma aetiopathogenesis and experimental studies. *Oral Oncol.* Feb 2. [Epub ahead of print].

394

- Bahar G, Feinmesser R, Shpitzer T *et al* (2007). Salivary analysis in oral cancer patients: DNA and protein oxidation, reactive nitrogen species, and antioxidant profile. *Cancer* **109:** 54–59.
- Ball VA, Righi PD, Tejada E *et al* (1997). p53 immunostaining of surgical margins as a predictor of local recurrence in squamous cell carcinoma of the oral cavity and oropharynx. *Ear Nose Throat J* **76:** 818–823.
- Bilde A, von Buchwald C, Dabelsteen E *et al* (2009). Molecular markers in the surgical margin of oral carcinomas. *J Oral Pathol Med* **38**: 72–78.
- Bloching M, Reich W, Schubert J *et al* (2007). The influence of oral hygiene on salivary quality in the Ames Test, as a marker for genotoxic effects. *Oral Oncol* **43**: 933–939.
- Boccia S, Cadoni G, Sayed-Tabatabaei FA *et al* (2008). CYP1A1, CYP2E1, GSTM1, GSTT1, EPHX1 exons 3 and 4, and NAT2 polymorphisms, smoking, consumption of alcohol and fruit and vegetables and risk of head and neck cancer. *J Cancer Res Clin Oncol* **134**: 93–100.
- Boccia S, Hashibe M, Gallì P *et al* (2009a). Aldehyde dehydrogenase 2 and head and neck cancer: a meta-analysis implementing a Mendelian randomization approach. *Cancer Epidemiol Biomarkers Prev* **18**: 248–254.
- Boccia S, Boffetta P, Brennan P *et al* (2009b). Meta-analyses of the methylenetetrahydrofolate reductase C677T and A1298C polymorphisms and risk of head and neck and lung cancer. *Cancer Lett* **273:** 55–61.
- Boffetta P, Hecht S, Gray N et al (2008). Smokeless tobacco and cancer. Lancet Oncol 9: 667–675.
- Boldrup L, Coates PJ, Hedberg Y *et al* (2005). Expression of p63, COX-2, EGFR and beta-catenin in smokers and patients with squamous cell carcinoma of the head and neck reveal variations in non-neoplastic tissue and no obvious changes in smokers. *Int J Oncol* **27**: 1661–1667.
- Bouchardy C, Hirvonen A, Coutelle C et al (2000). Role of alcohol dehydrogenase 3 and cytochrome P-4502E1 genotypes in susceptibility to cancers of the upper aerodigestive tract. Int J Cancer 87: 734–740.
- Boyle JO, Mao L, Brennan JA *et al* (1994). Gene mutations in saliva as molecular markers for head and neck squamous cell carcinomas. *Am J Surg* **168**: 429–432.
- Braakhuis BJ, Tabor MP, Kummer JA *et al* (2003). A genetic explanation of Slaughter's concept of field cancerization: evidence and clinical implications. *Cancer Res* **63**: 1727–1730.
- Braakhuis BJ, Leemans CR, Brakenhoff RH (2004). A genetic progression model of oral cancer: current evidence and clinical implications. *J Oral Pathol Med* **33:** 317–322.
- Braakhuis BJ, Brakenhoff RH, Leemans CR (2005a). Head and neck cancer: molecular carcinogenesis. *Ann Oncol* **16**(Suppl 2): ii249–ii250.
- Braakhuis BJ, Leemans CR, Brakenhoff RH (2005b). Expanding fields of genetically altered cells in head and neck squamous carcinogenesis. *Semin Cancer Biol* **15**: 113– 120.
- Bray F, Sankila R, Ferlay J *et al* (2002). Estimates of cancer incidence and mortality in Europe in 1995. *Eur J Cancer* 38: 99–166.
- Brennan JA, Mao L, Hruban RH *et al* (1995). Molecular assessment of histopathological staging in squamous-cell carcinoma of the head and neck. *N Engl J Med* **332**: 429–435.
- Brinkman BM, Wong DT (2006). Disease mechanism and biomarkers of oral squamous cell carcinoma. *Curr Opin Oncol* 18: 228–233.
- Brown KS, Kane MA (2006). Chemoprevention of squamous cell carcinoma of the oral cavity. *Otolaryngol Clin North Am* **39:** 349–363.

- Brunnemann KD, Prokopczyk B, Djordjevic MV *et al* (1996). Formation and analysis of tobacco-specific *N*-nitrosamines. *Crit Rev Toxicol* **26:** 121–137.
- Buch SC, Nazar-Stewart V, Weissfeld JL *et al* (2008). Case– control study of oral and oropharyngeal cancer in whites and genetic variation in eight metabolic enzymes. *Head Neck* **30**: 1139–1147.
- Califano J, Westra WH, Meininger G *et al* (2000). Genetic progression and clonal relationship of recurrent premalignant head and neck lesions. *Clin Cancer Res* **6**: 347–352.
- Carmeliet P, Jain RK (2000). Angiogenesis in cancer and other diseases. *Nature* 407: 249–257.
- Carpenter JM, Syms MJ, Sniezek JC (2005). Oral carcinoma associated with betel nut chewing in the Pacific: an impending crisis? *Pac Health Dialog* **12**: 158–162.
- Chen YJ, Chang JT, Liao CT *et al* (2008). Head and neck cancer in the betel quid chewing area: recent advances in molecular carcinogenesis. *Cancer Sci* **99**: 1507–1514.
- Cheong SC, Chandramouli GV, Saleh A *et al* (2009). Gene expression in human oral squamous cell carcinoma is influenced by risk factor exposure. *Oral Oncol.* Jan 13. [Epub ahead of print].
- Chiang WF, Liu SY, Yen CY *et al* (2008). Association of epidermal growth factor receptor (EGFR) gene copy number amplification with neck lymph node metastasis in arecaassociated oral carcinomas. *Oral Oncol* **44**: 270–276.
- Chiesa F, Sala L, Costa L *et al* (1986). Excision of oral leukoplakias by CO2 laser on an out-patient basis: a useful procedure for prevention and early detection of oral carcinomas. *Tumori* **72**: 307–312.
- Chiu CF, Tsai MH, Tseng HC *et al* (2008a). A novel single nucleotide polymorphism in ERCC6 gene is associated with oral cancer susceptibility in Taiwanese patients. *Oral Oncol* **44:** 582–586.
- Chiu CF, Tsai MH, Tseng HC *et al* (2008b). A novel single nucleotide polymorphism in XRCC4 gene is associated with oral cancer susceptibility in Taiwanese patients. *Oral Oncol* **44:** 898–902.
- Cogliano V, Straif K, Baan R *et al* (2004). Smokeless tobacco and tobacco-related nitrosamines. *Lancet Oncol* **5**: 708.
- Cortesina G, Martone T, Galeazzi E *et al* (2000). Staging of head and neck squamous cell carcinoma using the MET oncogene product as marker of tumor cells in lymph node metastases. *Int J Cancer* **89**: 286–292.
- Cruz IB, Snijders PJ, Meijer CJ *et al* (1998). p53 expression above the basal cell layer in oral mucosa is an early event of malignant transformation and has predictive value for developing oral squamous cell carcinoma. *J Pathol* **184**: 360–368.
- D'Souza G, Kreimer AR, Viscidi R *et al* (2007). Case–control study of human papillomavirus and oropharyngeal cancer. *N Engl J Med* **356**: 1944–1956.
- Datta S, Majumder M, Biswas NK *et al* (2007). Increased risk of oral cancer in relation to common Indian mitochondrial polymorphisms and Autosomal GSTP1 locus. *Cancer* **110**: 1991–1999.
- Derka S, Vairaktaris E, Papakosta V *et al* (2006). Cell proliferation and apoptosis culminate in early stages of oral oncogenesis. *Oral Oncol* **42:** 540–550.
- Dikshit RP, Ramadas K, Hashibe M *et al* (2006). Association between diabetes mellitus and pre-malignant oral diseases: a cross sectional study in Kerala, India. *Int J Cancer* **118**: 453–457.
- Diniz-Freitas M, García-Caballero T, Antúnez-López J *et al* (2006). Reduced E-cadherin expression is an indicator of unfavourable prognosis in oral squamous cell carcinoma. *Oral Oncol* **42**: 190–200.

- Drummond SN, De Marco L, Pordeus Ide A *et al* (2002). TP53 codon 72 polymorphism in oral squamous cell carcinoma. *Anticancer Res* **22**: 3379–3381.
- Duarte EC, Ribeiro DC, Gomez MV *et al* (2008). Genetic polymorphisms of carcinogen metabolizing enzymes are associated with oral leukoplakia development and p53 overexpression. *Anticancer Res* **28**: 1101–1106.
- Dye BA, Wang R, Lashley R *et al* (2007). Using NHANES oral health examination protocols as part of an esophageal cancer screening study conducted in a high-risk region of China. *BMC Oral Health* **17:** 7–10.
- Fasanmade A, Kwok E, Newman L (2007). Oral squamous cell carcinoma associated with khat chewing. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* **104:** e53–e55.
- Fialka F, Gruber RM, Hitt R *et al* (2008). CPA6, FMO2, LGI1, SIAT1 and TNC are differentially expressed in earlyand late-stage oral squamous cell carcinoma – a pilot study. *Oral Oncol* **44**: 941–948.
- Fischer DJ, Epstein JB, Morton TH *et al* (2004). Interobserver reliability in the histopathologic diagnosis of oral pre-malignant and malignant lesions. *J Oral Pathol Med* **33**: 65–70.
- Fischer DJ, Epstein JB, Morton TH Jr *et al* (2005). Reliability of histologic diagnosis of clinically normal intraoral tissue adjacent to clinically suspicious lesions in former upper aerodigestive tract cancer patients. *Oral Oncol* **41**: 489–496.
- Franzmann EJ, Weed DT, Civantos FJ *et al* (2001). A novel CD44 v3 isoform is involved in head and neck squamous cell carcinoma progression. *Otolaryngol Head Neck Surg* **124**: 426–432.
- Franzmann EJ, Reategui EP, Carraway KL *et al* (2005). Salivary soluble CD44: a potential molecular marker for head and neck cancer. *Cancer Epidemiol Biomarkers Prev* **14**: 735–739.
- Friedrich RE, Bartel-Friedrich S, Plambeck K *et al* (1997). P53 auto-antibodies in the sera of patients with oral squamous cell carcinoma. *Anticancer Res* **17:** 3183–3184.
- Gabriel HE, Crott JW, Ghandour H *et al* (2006). Chronic cigarette smoking is associated with diminished folate status, altered folate form distribution, and increased genetic damage in the buccal mucosa of healthy adults. *Am J Clin Nutr* **83**: 835–841.
- Garavello W, Giordano L, Bosetti C *et al* (2008). Diet diversity and the risk of oral and pharyngeal cancer. *Eur J Nutr* **47**: 280–284.
- Giebutowicz J, Wroczyński P, Piekarczyk J *et al* (2008). Fluorimetric detection of aldehyde dehydrogenase activity in human tissues in diagnostic of cancers of oral cavity. *Acta Pol Pharm* **65:** 81–84.
- Glazer CA, Chang SS, Ha PK *et al* (2008). Applying the molecular biology and epigenetics of head and neck cancer in everyday clinical practice. *Oral Oncol.* Jul 30. [Epub ahead of print].
- Gottschlich S, Folz BJ, Goeroegh T *et al* (1999). A new prognostic indicator for head and neck cancer p53 serum antibodies? *Anticancer Res* **19:** 2703–2705.
- Goutzanis L, Vairaktaris E, Yapijakis C *et al* (2007). Diabetes may increase risk for oral cancer through the insulin receptor substrate-1 and focal adhesion kinase pathway. *Oral Oncol* **43:** 165–173.
- Guh JY, Chen HC, Tsai JF *et al* (2007). Betel-quid use is associated with heart disease in women. *Am J Clin Nutr* **85**: 1229–1235.
- Guha N, Boffetta P, Wünsch Filho V *et al* (2007). Oral health and risk of squamous cell carcinoma of the head and neck and esophagus: results of two multicentric case–control studies. *Am J Epidemiol* **166**: 1159–1173.

- Guo Z, Yamaguchi K, Sanchez-Cespedes M *et al* (2001). Allelic losses in OraTest-directed biopsies of patients with prior upper aerodigestive tract malignancy. *Clin Cancer Res* **7:** 1963–1968.
- Gupta R, Sharma SC, Das SN (2008). Association of TNFalpha and TNFR1 promoters and 3' UTR region of TNFR2 gene polymorphisms with genetic susceptibility to tobaccorelated oral carcinoma in Asian Indians. *Oral Oncol* **44:** 455– 463.
- Ha PK, Chang SS, Glazer CA *et al* (2008). Molecular techniques and genetic alterations in head and neck cancer. *Oral Oncol.* Jul 30. [Epub ahead of print].
- Haddad RI, Shin DM (2008). Recent advances in head and neck cancer. *New Engl J Med* **359**: 1143–1154.
- Haddad R, Crum C, Chen Z *et al* (2008). HPV16 transmission between a couple with HPV-related head and neck cancer. *Oral Oncol* **44**: 812–815.
- Hamidi S, Salo T, Kainulainen T *et al* (2000). Expression of alpha(v)beta6 integrin in oral leukoplakia. *Br J Cancer* 82: 1433–1440.
- Hashibe M, Straif K, Tashkin DP *et al* (2005). Epidemiologic review of marijuana use and cancer risk. *Alcohol* **35**: 265–275.
- Hashibe M, McKay JD, Curado MP *et al* (2008). Multiple ADH genes are associated with upper aerodigestive cancers. *Nat Genet* **40**: 707–709.
- Hashibe M, Brennan P, Chuang SC *et al* (2009). Interaction between tobacco and alcohol use and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. *Cancer Epidemiol Biomarkers Prev* 18: 541–550.
- Hatagima A, Costa EC, Marques CF *et al* (2008). Glutathione S-transferase polymorphisms and oral cancer: a case–control study in Rio de Janeiro, Brazil. *Oral Oncol* **44:** 200–207.
- Hein DW, Doll MA, Fretland AJ *et al* (2000). Molecular genetics and epidemiology of the NAT1 and NAT2 acetylation polymorphisms. *Cancer Epidemiol Biomarkers Prev* **9**: 29–42.
- Hirota SK, Braga FP, Penha SS *et al* (2008). Risk factors for oral squamous cell carcinoma in young and older Brazilian patients: a comparative analysis. *Med Oral Patol Oral Cir Bucal* **13**: E227–E231.
- Hoffmann D, Hoffmann I (1997). The changing cigarette, 1950–1995. J Toxicol Environ Health **50:** 307–364.
- Holmes L Jr, Desvignes-Kendrick M, Slomka J et al (2008). Is dental care utilization associated with oral cavity cancer in a large sample of community-based United States residents? *Community Dent Oral Epidemiol*. Nov 19. [Epubahead of print].
- Holmstrup P, Vedtofte P, Reibel J *et al* (2007). Oral premalignant lesions: is a biopsy reliable? *J Oral Pathol Med* **36**: 262–266.
- van Houten VM, Tabor MP, van den Brekel MW et al (2000a). Molecular assays for the diagnosis of minimal residual head-and-neck cancer: methods, reliability, pitfalls, and solutions. *Clin Cancer Res* **6**: 3803–3816.
- van Houten VM, van den Brekel MW, Denkers F et al (2000b). Molecular diagnosis of head and neck cancer. Recent Results Cancer Res 157: 90–106.
- Hu S, Li Y, Wang J, Xie Y *et al* (2006). Human saliva proteome and transcriptome. *J Dent Res* **85**: 1129–1133.
- Hu S, Loo JA, Wong DT (2007). Human saliva proteome analysis. *Ann N Y Acad Sci* **1098**: 323–329.
- Hu S, Arellano M, Boontheung P *et al* (2008). Salivary proteomics for oral cancer biomarker discovery. *Clin Cancer Res* **14**: 6246–6252.
- IARC (2004). GLOBOCAN 2002. Cancer Incidence, Mortality and Prevalence Worldwide (2002 estimates).

- Izzo JG, Papadimitrakopoulou VA, Liu DD *et al* (2003). Cyclin D1 genotype, response to biochemoprevention, and progression rate to upper aerodigestive tract cancer. *J Natl Cancer Inst* **95**: 198–205.
- Jacob BJ, Straif K, Thomas G *et al* (2004). Betel quid without tobacco as a risk factor for oral precancers. *Oral Oncol* **40**: 697–704.
- Johnson N (2001). Tobacco use and oral cancer: a global perspective. J Dent Educ **65**: 328–339.
- Kannan S, Tahara H, Yokozaki H et al (1997). Telomerase activity in premalignant and malignant lesions of human oral mucosa. *Cancer Epidemiol Biomarkers Prev* 6: 413– 420.
- Karabulut A, Reibel J, Therkildsen MH *et al* (1995). Observer variability in the histologic assessment of oral premalignant lesions. *J Oral Pathol Med* **24:** 198–200.
- Kim HR, Christensen R, Park NH *et al* (2001). Elevated expression of hTERT is associated with dysplastic cell transformation during human oral carcinogenesis in situ. *Clin Cancer Res* **7:** 3079–3086.
- Kuratomi Y, Kumamoto M, Kidera K *et al* (2006). Diffuse expression of laminin gamma2 chain in disseminating and infiltrating cancer cells indicates a highly malignant state in advanced tongue cancer. *Oral Oncol* **42**: 73–76.
- Kurkivuori J, Salaspuro V, Kaihovaara P *et al* (2007). Acetaldehyde production from ethanol by oral streptococci. *Oral Oncol* **43**: 181–186.
- La Vecchia C (2009). Mouthwash and oral cancer risk: an update. *Oral Oncol* **45:** 198–200.
- La Vecchia C, Lucchini F, Negri E *et al* (2004). Trends in oral cancer mortality in Europe. *Oral Oncol* **40**: 433–439.
- Lee JJ, Hong WK, Hittelman WN *et al* (2000). Predicting cancer development in oral leukoplakia: ten years of translational research. *Clin Cancer Res* **6**: 1702–1710.
- Lin YC, Huang HI, Wang LH *et al* (2008). Polymorphisms of COX-2–765G > C and p53 codon 72 and risks of oral squamous cell carcinoma in a Taiwan population. *Oral Oncol* 44: 798–804.
- Lingen MW, Kalmar JR, Karrison T *et al* (2008). Critical evaluation of diagnostic aids for the detection of oral cancer. *Oral Oncol* **44**: 10–22.
- Lippman SM, Hong WK (1989). Second malignant tumors in head and neck squamous cell carcinoma: the overshadowing threat for patients with early-stage disease. *Int J Radiat Oncol Biol Phys* **17:** 691–694.
- Llewellyn CD, Johnson NW, Warnakulasuriya KA (2001). Risk factors for squamous cell carcinoma of the oral cavity in young people – a comprehensive literature review. *Oral Oncol* 37: 401–418.
- Losi-Guembarovski R, Cólus IM, De Menezes RP *et al* (2008). Lack of association among polymorphic xenobiotic-metabolizing enzyme genotypes and the occurrence and progression of oral carcinoma in a Brazilian population. *Anticancer Res* **28**: 1023–1028.
- Lucenteforte E, Garavello W, Bosetti C *et al* (2008). Dietary factors and oral and pharyngeal cancer risk. *Oral Oncol*. Nov 4. [Epub ahead of print].
- Luzar B, Poljak M, Marin IJ *et al* (2004). Human telomerase catalytic subunit gene re-expression is an early event in oral carcinogenesis. *Histopathology* **45**: 13–19.
- Maass JD, Gottschlich S, Goeroegh T *et al* (1997). Head and neck cancer and p53-immunogenicity. *Anticancer Res* 17: 2873–2874.
- Majumder M, Sikdar N, Ghosh S *et al* (2007). Polymorphisms at XPD and XRCC1 DNA repair loci and increased risk of oral leukoplakia and cancer among NAT2 slow acetylators. *Int J Cancer* **120**: 2148–2156.

- Marques CF, Koifman S, Koifman RJ *et al* (2006). Influence of CYP1A1, CYP2E1, GSTM3 and NAT2 genetic polymorphisms in oral cancer susceptibility: results from a case– control study in Rio de Janeiro. *Oral Oncol* **42**: 632–637.
- Maserejian NN, Giovannucci E, Rosner B et al (2006). Prospective study of fruits and vegetables and risk of oral premalignant lesions in men. Am J Epidemiol 164: 556–566.
- Maserejian NN, Giovannucci E, Rosner B *et al* (2007). Prospective study of vitamins C, E, and A and carotenoids and risk of oral premalignant lesions in men. *Int J Cancer* **120:** 970–977.
- Máthé M, Suba Z, Németh Z *et al* (2006). Stromal syndecan-1 expression is an adverse prognostic factor in oral carcinomas. *Oral Oncol* **42**: 493–500.
- Matsuda T, Yabushita H, Kanaly RA *et al* (2006). Increased DNA damage in ALDH2-deficient alcoholics. *Chem Res Toxicol* **19**: 1374–1378.
- McCullough MJ, Farah CS (2008). The role of alcohol in oral carcinogenesis with particular reference to alcohol-containing mouthwashes. *Aust Dent J* **53**: 302–305.
- Merchant A, Husain SS, Hosain M *et al* (2000). Paan without tobacco: an independent risk factor for oral cancer. *Int J Cancer* **86**: 128–131.
- Meyer MS, Joshipura K, Giovannucci E *et al* (2008). A review of the relationship between tooth loss, periodontal disease, and cancer. *Cancer Causes Control* **19:** 895–907.
- Mithani SK, Mydlarz WK, Grumbine FL *et al* (2007). Molecular genetics of premalignant oral lesions. *Oral Dis* **13**: 126–133.
- Molinolo AA, Amornphimoltham P, Squarize CH *et al* (2008). Dysregulated molecular networks in head and neck carcinogenesis. *Oral Oncol.* Sep 18. [Epub ahead of print].
- Moore S, Johnson N, Pierce A *et al* (1999). The epidemiology of lip cancer: a review of global incidence and aetiology. *Oral Dis* **5**: 185–195.
- Moore SR, Johnson NW, Pierce AM *et al* (2000a). The epidemiology of tongue cancer: a review of global incidence. *Oral Dis* **6**: 75–84.
- Moore SR, Johnson NW, Pierce AM *et al* (2000b). The epidemiology of mouth cancer: a review of global incidence. *Oral Dis* **6**: 65–74.
- Mutirangura A, Supiyaphun P, Trirekapan S *et al* (1996). Telomerase activity in oral leukoplakia and head and neck squamous cell carcinoma. *Cancer Res* **56:** 3530–3533.
- Muto M, Hitomi Y, Ohtsu A *et al* (2000). cetaldehyde production by non-pathogenic Neisseria in human oral microflora: implications for carcinogenesis in upper aerodigestive tract. *Int J Cancer* **88**: 342–350.
- Muwonge R, Ramadas K, Sankila R *et al* (2008). Role of tobacco smoking, chewing and alcohol drinking in the risk of oral cancer in Trivandrum, India: a nested case–control design using incident cancer cases. *Oral Oncol* **44**: 446–454.
- Park NJ, Zhou X, Yu T *et al* (2007). Characterization of salivary RNA by cDNA library analysis. *Arch Oral Biol* 52: 30–35.
- Park S, Bae J, Nam BH *et al* (2008). Aetiology of cancer in Asia. *Asian Pac J Cancer Prev* **9**: 371–380.
- Patel V, Leethanakul C, Gutkind JS (2001). New approaches to the understanding of the molecular basis of oral cancer. *Crit Rev Oral Biol Med* **12:** 55–63.
- Patel BP, Rawal UM, Rawal RM *et al* (2008). Tobacco, antioxidant enzymes, oxidative stress, and genetic susceptibility in oral cancer. *Am J Clin Oncol* **31**: 454–459.
- Pavia M, Pileggi C, Nobile CG *et al* (2006). ssociation between fruit and vegetable consumption and oral cancer: a metaanalysis of observational studies. *Am J Clin Nutr* **83**: 1126–1134.

- Pelucchi C, Talamini R, Negri E *et al* (2003). Folate intake and risk of oral and pharyngeal cancer. *Ann Oncol* **14**: 1677–1681.
- Pelucchi C, Gallus S, Garavello W *et al* (2008). Alcohol and tobacco use, and cancer risk for upper aerodigestive tract and liver. *Eur J Cancer Prev* **17:** 340–344.
- Petti S (2008). Lifestyle risk factors for oral cancer. *Oral Oncol* [Epub ahead of print].
- Pickering V, Jordan RC, Schmidt BL (2007). Elevated salivary endothelin levels in oral cancer patients – a pilot study. *Oral Oncol* **43**: 37–41.
- Proia NK, Paszkiewicz GM, Nasca MA *et al* (2006). Smoking and smokeless tobacco-associated human buccal cell mutations and their association with oral cancer – a review. *Cancer Epidemiol Biomarkers Prev* **15**: 1061–1077.
- Pu X, Lippman SM, Yang H *et al* (2009). Cyclooxygenase-2 gene polymorphisms reduce the risk of oral premalignant lesions. *Cancer* 2009 Feb 5. [Epub ahead of print].
- Reategui EP, de Mayolo AA, Das PM *et al* (2007). Characterization of CD44v3-containing isoforms in head and neck cancer. *Cancer Biol Ther* **5**: 1163–1168.
- Reibel J (2003). Prognosis of oral pre-malignant lesions: significance of clinical, histopathological, and molecular biological characteristics. *Crit Rev Oral Biol Med* 14: 47–62.
- Reichart PA, Nguyen XH (2008). Betel quid chewing, oral cancer and other oral mucosal diseases in Vietnam: a review. *J Oral Pathol Med* 37: 511–514.
- Richie JP Jr, Kleinman W, Marina P *et al* (2008). Blood iron, glutathione, and micronutrient levels and the risk of oral cancer. *Nutr Cancer* **60**: 474–482.
- Rodriguez T, Altieri A, Chatenoud L *et al* (2004). Risk factors for oral and pharyngeal cancer in young adults. *Oral Oncol* **40**: 207–213.
- Roepman P, Wessels LF, Kettelarij N *et al* (2005). An expression profile for diagnosis of lymph node metastases from primary head and neck squamous cell carcinomas. *Nat Genet* **37**: 182–186.
- Rossi M, Garavello W, Talamini R *et al* (2007). Flavonoids and the risk of oral and pharyngeal cancer: a case–control study from Italy. *Cancer Epidemiol Biomarkers Prev* 16: 1621–1625.
- Santhi WS, Sebastian P, Varghese BT *et al* (2006). NF-kappaB and COX-2 during oral tumorigenesis and in assessment of minimal residual disease in surgical margins. *Exp Mol Pathol* 81: 123–130.
- Sanz-Ortega J, Roig F, Al-Mousa MM *et al* (2007). 17p13 (p53 locus), 5q21 (APC locus) and 9p21 (p16 locus) allelic deletions are frequently found in oral exfoliative cytology cells from smoker patients with non-small-cell lung cancer. *Histol Histopathol* **22**: 541–545.
- Sapkota A, Hsu CC, Zaridze D *et al* (2008). Dietary risk factors for squamous cell carcinoma of the upper aerodigestive tract in central and eastern Europe. *Cancer Causes Control* **19:** 1161–1170.
- Sawair FA, Al-Mutwakel A, Al-Eryani K *et al* (2007). High relative frequency of oral squamous cell carcinoma in Yemen: qat and tobacco chewing as its aetiological background. *Int J Environ Health Res* **17:** 185–195.
- Scheer M, Kuebler AC, Zöller JE (2004). Chemoprevention of oral squamous cell carcinomas. *Onkologie* 27: 187–193.
- Schwartz SM, Doody DR, Fitzgibbons ED *et al* (2001). Oral squamous cell cancer risk in relation to alcohol consumption and alcohol dehydrogenase-3 genotypes. *Cancer Epidemiol Biomarkers Prev* **10**: 1137–1144.
- Scully C (1995). Oral precancer: preventive and medical approaches to management. *Eur J Cancer B Oral Oncol* **31B:** 16–26.

- Scully C (2002). Oral squamous cell carcinoma; from an hypothesis about a virus, to concern about possible sexual transmission. *Oral Oncol* **38**: 227–234.
- Scully C (2005). Oral cancer; the evidence for sexual transmission. Br Dent J 199: 203–207.
- Scully C, Bedi R (2000). Ethnicity and oral cancer. *Lancet* Oncol 1: 37–42.
- Scully C, Moles D (2008). Oral cancer. In: Heggenhougen KH, Quah S, eds *Encyclopedia of public health*. Vol 4. San Diego, CA: Academic Press, pp. 668–677.
- Scully C, Field JK, Tanzawa H (2000a). Genetic aberrations in oral or head and neck squamous cell carcinoma (SCCHN):
 1. Carcinogen metabolism, DNA repair and cell cycle control. *Oral Oncol* 36: 256–263.
- Scully C, Field JK, Tanzawa H (2000b). Genetic aberrations in oral or head and neck squamous cell carcinoma 2: chromosomal aberrations. *Oral Oncol* 36: 311–327.
- Scully C, Field JK, Tanzawa H (2000c). Genetic aberrations in oral or head and neck squamous cell carcinoma 3: clinicopathological applications. *Oral Oncol* 36: 404–413.
- Scully C, Sudbø J, Speight PM (2003). Progress in determining the malignant potential of oral lesions. J Oral Pathol Med 32: 251–256.
- Seitz HK, Cho CH (2009). Contribution of alcohol and tobacco use in gastrointestinal cancer development. *Methods Mol Biol* **472**: 217–241.
- Seitz HK, Stickel F (2007). Molecular mechanisms of alcohol-mediated carcinogenesis. *Nat Rev Cancer* 7: 599–612.
- Serefoglou Z, Yapijakis C, Nkenke E *et al* (2008). Genetic association of cytokine DNA polymorphisms with head and neck cancer. *Oral Oncol* **44**: 1093–1099.
- Shaw RJ, Liloglou T, Rogers SN *et al* (2006). Promoter methylation of P16, RARbeta, E-cadherin, cyclin A1 and cytoglobin in oral cancer: quantitative evaluation using pyrosequencing. *Br J Cancer* **94**: 561–568.
- Shaw RJ, Hall GL, Woolgar JA *et al* (2007). Quantitative methylation analysis of resection margins and lymph nodes in oral squamous cell carcinoma. *Br J Oral Maxillofac Surg* **45:** 617–622.
- Shillitoe EJ (2008). The role of viruses in squamous cell carcinoma of the oropharyngeal mucosa. *Oral Oncol.* Oct 24. [Epub ahead of print].
- Shpitzer T, Bahar G, Feinmesser R *et al* (2007). A comprehensive salivary analysis for oral cancer diagnosis. *J Cancer Res Clin Oncol* **133**: 613–617.
- Slaughter DP, Southwick HW, Smejkal W (1953). Field cancerization in oral stratified squamous epithelium; clinical implications of multicentric origin. *Cancer* **6**: 963–968.
- Solomon PR, Selvam GS, Shanmugam G (2008). Polymorphism in ADH and MTHFR genes in oral squamous cell carcinoma of Indians. *Oral Dis* 14: 633–639.
- Sridhar S, Schembri F, Zeskind J *et al* (2008). Smoking-induced gene expression changes in the bronchial airway are reflected in nasal and buccal epithelium. *BMC Genomics* **9**: 259.
- Sumida T, Hamakawa H, Sogawa K *et al* (1999). Telomerase components as a diagnostic tool in human oral lesions. *Int J Cancer* **80**: 1–4.
- Sun W, Zhou S, Chang SS *et al* (2009). Mitochondrial mutations contribute to HIF1alpha accumulation via increased reactive oxygen species and up-regulated pyruvate dehydrogenase kinase 2 in head and neck squamous cell carcinoma. *Clin Cancer Res* **15**: 476–484.
- Suzuki T, Wakai K, Matsuo K *et al* (2006). Effect of dietary antioxidants and risk of oral, pharyngeal and laryngeal squamous cell carcinoma according to smoking and drinking habits. *Cancer Sci* **97**: 760–767.

398

- Tabor MP, Brakenhoff RH, Ruijter-Schippers HJ *et al* (2004). Genetically altered fields as origin of locally recurrent head and neck cancer: a retrospective study. *Clin Cancer Res* **10**: 3607–3613.
- Takeshima M, Saitoh M, Kusano K *et al* (2008). High frequency of hypermethylation of p14, p15 and p16 in oral pre-cancerous lesions associated with betel-quid chewing in Sri Lanka. *J Oral Pathol Med* **37**: 475–479.
- Tavassoli M, Brunel N, Maher R *et al* (1998). p53 antibodies in the saliva of patients with squamous cell carcinoma of the oral cavity. *Int J Cancer* **78**: 390–391.
- Tezal M, Sullivan MA, Reid ME et al (2007). Chronic periodontitis and the risk of tongue cancer. Arch Otolaryngol Head Neck Surg 133: 450–454.
- Thomas SJ, Bain CJ, Battistutta D *et al* (2007). Betel quid not containing tobacco and oral cancer: a report on a case– control study in Papua New Guinea and a meta-analysis of current evidence. *Int J Cancer* **120**: 1318–1323.
- Thomas SJ, Harris R, Ness AR *et al* (2008). Betel quid not containing tobacco and oral leukoplakia: a report on a cross-sectional study in Papua New Guinea and a meta-analysis of current evidence. *Int J Cancer* **123**: 1871–1876.
- Thomson PJ (2002). Field change and oral cancer: new evidence for widespread carcinogenesis? *Int J Oral Maxillofac Surg* **31:** 262–266.
- Thomson PJ, Hamadah O (2007). Cancerisation within the oral cavity: the use of 'field mapping biopsies' in clinical management. *Oral Oncol* **43**: 20–26.
- Thongprasom K, Mutirangura A, Cheerat S (1998). Telomerase activity in oral lichen planus. J Oral Pathol Med 27: 395– 398.
- Tjebbes GW, Leppers vd Straat FG, Tilanus MG *et al* (1999). p53 tumor suppressor gene as a clonal marker in head and neck squamous cell carcinoma: p53 mutations in primary tumor and matched lymph node metastases. *Oral Oncol* **35**: 384–389.
- Trullenque-Eriksson A, Muñoz-Corcuera M, Campo-Trapero J et al (2009). Analysis of new diagnostic methods in suspicious lesions of the oral mucosa. *Med Oral Patol Oral Cir Bucal*. Feb 16. [Epub ahead of print].
- Tsai YS, Lee KW, Huang JL *et al* (2008). Arecoline, a major alkaloid of areca nut, inhibits p53, represses DNA repair, and triggers DNA damage response in human epithelial cells. *Toxicology* **249**: 230–237.
- Vallecillo Capilla M, Romero Olid MN, Olmedo Gaya MV et al (2007). Factors related to survival from oral cancer in an Andalusian population sample (Spain). *Med Oral Patol Oral Cir Bucal* **12:** E518–E523.
- Varela-Lema L, Taioli E, Ruano-Ravina A *et al* (2008). Metaanalysis and pooled analysis of GSTM1 and CYP1A1 polymorphisms and oral and pharyngeal cancers: a HuGE-GSEC review. *Genet Med* **10**: 369–384.
- Viet CT, Jordan RC, Schmidt BL (2007). DNA promoter hypermethylation in saliva for the early diagnosis of oral cancer. *J Calif Dent Assoc* **35**: 844–849.
- Vigneswaran N, Tilashalski K, Rodu B et al (1995). Tobacco use and cancer. A reappraisal. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 80: 178–182.
- van der Waal I (2008). Potentially malignant disorders of the oral and oropharyngeal mucosa; terminology, classification and present concepts of management. *Oral Oncol.* 2008 Jul 30. [Epub ahead of print].
- Wang J, Henry S, Yu T et al (2006). Salivary oral cancer transcriptome biomarkers (SOCTB) for clinical detection; 35th Annual Meeting of the American Association for Dental Research. Abstract #218.

- Wang Y, Spitz MR, Lee JJ *et al* (2007). Nucleotide excision repair pathway genes and oral premalignant lesions. *Clin Cancer Res* **13**: 3753–3758.
- Warnakulasuriya S (2000). Lack of molecular markers to predict malignant potential of oral precancer. *J Pathol* **190**: 407–409.
- Warnakulasuriya S (2008). Global epidemiology of oral and oropharyngeal cancer. *Oral Oncol.* 2008 Sep 17. [Epub ahead of print].
- Warnakulasuriya KA, Ralhan R (2007). Clinical, pathological, cellular and molecular lesions caused by oral smokeless tobacco – a review. J Oral Pathol Med 36: 63– 77.
- Winn DM (1995). Diet and nutrition in the etiology of oral cancer. *Am J Clin Nutr* **61:** 437S–445S.
- Wong DT (2006). Salivary diagnostics for oral cancer. J Calif Dent Assoc 34: 303–308.
- Xie H, Onsongo G, Popko J *et al* (2008). Proteomics analysis of cells in whole saliva from oral cancer patients via value-added three-dimensional peptide fractionation and tandem mass spectrometry. *Mol Cell Proteomics* 7: 486–498.
- Yajima Y, Noma H, Furuya Y *et al* (2004). Quantification of telomerase activity of regions unstained with iodine solution that surround oral squamous cell carcinoma. *Oral Oncol* 40: 314–320.
- Ye Y, Lippman SM, Lee JJ *et al* (2008). Genetic variations in cell-cycle pathway and the risk of oral premalignant lesions. *Cancer* **113**: 2488–2495.
- Yokoyama A, Omori T (2003). Genetic polymorphisms of alcohol and aldehyde dehydrogenases and risk for esophageal and head and neck cancers. *Jpn J Clin Oncol* **33:** 111–121.
- Yokoyama A, Tsutsumi E, Imazeki H *et al* (2007a). Contribution of the alcohol dehydrogenase-1B genotype and oral microorganisms to high salivary acetaldehyde concentrations in Japanese alcoholic men. *Int J Cancer* **121**: 1047–1054.
- Yokoyama A, Tsutsumi E, Imazeki H *et al* (2008). Salivary acetaldehyde concentration according to alcoholic beverage consumed and aldehyde dehydrogenase-2 genotype. *Alcohol Clin Exp Res* **32:** 1607–1614.
- Zain RB (2001). Cultural and dietary risk factors of oral cancer and precancer a brief overview. *Oral Oncol* **37:** 205–210.
- Zavras AI, Wu T, Laskaris G *et al* (2002). Interaction between a single nucleotide polymorphism in the alcohol dehydrogenase 3 gene, alcohol consumption and oral cancer risk. *Int J Cancer* **97:** 526–530.
- Zhang L, Williams M, Poh CF *et al* (2005). Toluidine blue staining identifies high-risk primary oral premalignant lesions with poor outcome. *Cancer Res* **65**: 8017–8021.
- Zheng TZ, Boyle P, Hu HF *et al* (1990). Dentition, oral hygiene, and risk of oral cancer: a case–control study in Beijing, People's Republic of China. *Cancer Causes Control* **1**: 235–241.
- Zhong LP, Zhang CP, Zheng JW *et al* (2007). Increased Cyfra 21-1 concentration in saliva from primary oral squamous cell carcinoma patients. *Arch Oral Biol* **52**: 1079–1087.
- Zhuo W, Wang Y, Zhuo X *et al* (2009). CYP1A1 and GSTM1 polymorphisms and oral cancer risk: association studies via evidence-based meta-analyses. *Cancer Invest* 27: 86–95.
- Zimmermann BG, Park NJ, Wong DT (2007). Genomic targets in saliva. *Ann N Y Acad Sci* **1098**: 184–191.

Copyright of Oral Diseases is the property of Blackwell Publishing Limited and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.