Oral Diseases (2011) 17, 355–361 doi:10.1111/j.1601-0825.2010.01763.x © 2010 John Wiley & Sons A/S All rights reserved

www.wiley.com

REVIEW ARTICLE

Anoikis mediators in oral squamous cell carcinoma

J Bunek*, P Kamarajan*, YL Kapila

Department of Periodontics and Oral Medicine, School of Dentistry, University of Michigan, Ann Arbor, MI, USA

Anoikis - apoptotic cell death triggered by loss of extracellular matrix (ECM) contacts - is dysregulated in many chronic debilitating and fatal diseases. Mechanisms rendering tumor cells resistant to anoikis, although not completely understood, possess significant therapeutic promise. In death receptor-mediated anoikis mechanisms, focal adhesion kinase (FAK) and receptor-interacting protein (RIP) dissociate, leading to association of RIP with Fas, formation of the death-inducing signaling complex (DISC), activation of caspase-3, and propagation of anoikis. In contrast, anoikis resistance is accomplished through constitutive activation of survival pathways that include integrin-dependent activation of FAK and extracellularsignal-regulated kinase (ERK). In addition, FAK and RIP association confers anoikis resistance by inhibiting the association of RIP with Fas and formation of the death signaling complex, which allows cells to escape anoikis. Up-regulation of CD44 also contributes to survival signals and promotes anoikis resistance. This review will focus on the roles of death receptors, prosurvival pathways, and the molecular players involved in anoikis escalation and resistance in oral squamous cell carcinoma. Oral Diseases (2011) 17, 355-361

Keywords: anoikis; oral squamous cell carcinoma; extracellular matrix; focal adhesion kinase; Fas/CD95; receptor interacting protein

Introduction

During development and normal physiologic processes, unwanted cells die by a process known as apoptosis. Tissue homeostasis is achieved when the rate of mitosis is in balance with apoptosis. When unbalanced, cells either divide too rapidly and resist normal apoptosis, leading to excessive cell accumulation as in cancer, or undergo premature apoptosis, leading to a net cell loss,

*These authors contributed equally.

as in the autoimmune diseases of pemphigus and pemphigoid (Gniadecki, 1998).

Apoptosis resulting from loss of cell adhesion to the extracellular matrix (ECM), or inappropriate adhesion is defined as 'anoikis' (Figure 1) (Frisch and Francis, 1994). Malignant tumors are not self-limiting in their growth, and capable of invading adjacent tissues and organs, as well as metastasizing to lymph nodes and other distant sites. Human tumor cell lines derived from metastatic lesions are more resistant to anoikis than tumors derived from primary oral squamous cell carcinomas (OSCC), and those tumor cell lines are more anoikis resistant than primary oral keratinocytes (Swan *et al*, 2003).

Oral squamous cell carcinoma, the most common malignant neoplasm of the oral cavity, is responsible for most deaths related to oral cancer and has a poor 5-year survival rate that has not changed in decades (Jemal *et al*, 2008). Patients with OSCC are commonly treated by radical surgical excision, resulting in significant loss of function and disfigurement, leading to a decrease in overall quality of life. As anoikis resistance leads to a more aggressive phenotype in cancers in general and in OSCC (Swan *et al*, 2003) understanding the processes that regulate anoikis will be paramount to advancing novel therapeutics to improve these stagnant statistics.

Anoikis and OSCC

Although anoikis resistance is implicated in different human malignancies, including ovarian (Tang et al, 2010), lung (Balsara and Testa, 2002), breast (Streuli and Gilmore, 1999), colon (Shanmugathasan and Jothy, 2000), and head and neck (Bockmuhl et al. 1997) cancers, novel findings are emerging about the role of anoikis in OSCC. When a tumor is confined to an organ or tissue, surgical removal and radiation therapy have proven effective; however, there are limited curative treatment options currently available for patients with metastatic disease (Garrison and Kyprianou, 2004). Tumor cells that acquire malignant potential have developed mechanisms to resist anoikis, allowing survival of cancer cells in systemic circulation, thereby facilitating secondary tumor formation in distant organ sites (Eble and Haier, 2006).

Correspondence: Yvonne L. Kapila, Department of Periodontics and Oral Medicine, School of Dentistry, University of Michigan, Ann Arbor, MI 48109-1078, USA. Tel: +1 734 615 2295, Fax: +1 734 763 5503, E-mail: ykapila@umich.edu

Received 3 June 2010; revised 13 August 2010; accepted 10 September 2010

Anoikis mediators in oral squamous cell carcinoma | Bunek et al

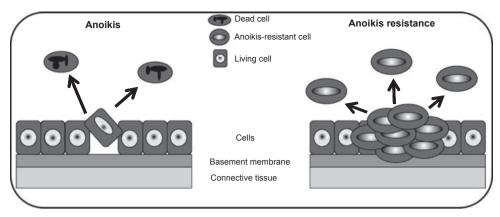


Figure 1 Schematic representation of anoikis vs anoikis resistance

Emerging data indicate that multiple pathways regulate anoikis, whereby involvement of integrin receptors, death receptors, adhesion molecules, and complex signaling cascades can lead to anoikis resistance and spread of metastatic cancer cells (Ishida *et al*, 2007; Kamarajan and Kapila, 2007; Kupferman *et al*, 2007; Chiarugi, 2008; Neiva *et al*, 2009). Anoikis-resistant OSCC cell lines exhibit a unique karyotypic and genotypic fingerprint that differs from that of anoikissensitive OSCC cells (Kupferman *et al*, 2007). Knowledge of these specific anoikis-resistance processes may help in developing therapies that explicitly target metastatic spread, new opportunities for less invasive and more successful treatments, and potential prognostic markers for aggressive OSCC.

Anoikis mediators

Anoikis mediators, and thus potential anoikis targets for therapy of aggressive OSCC, include ECM molecules, receptors that regulate cell survival pathways, such as integrins and proteoglycans, and cell death pathways, such as Fas/CD95, TNFR1, TNFR2, DR4, and DR5, and their cognate signaling molecules, plus molecules that engage in crosstalk between these two pathways.

ECM

The ECM is comprised of collagenous and non-collagenous proteins, proteoglycans and glycoproteins, including laminin and fibronectin, and growth factors, and alteration of their composition may promote cancer development and progression (Stetler-Stevenson, 1996). The ECM supports tissue homeostasis, is a physical scaffold for support and anchorage of cells, and it regulates the behavior of cells, influencing their survival, apoptosis, anoikis, differentiation, development, migration, proliferation, shape, and function.

Of the ECM proteins, type I collagen is more effective than fibronectin in delaying anoikis in some cell types (Bozzo *et al*, 2006). Furthermore, collagens are disrupted in invasive SCC, yet normal in carcinoma *in situ* or oral premalignant lesions (Giannelli *et al*, 2001). Matrix metalloproteinases (MMPs) can regulate OSCC tumor cell migration, invasion, survival, and apoptosis or anoikis presumably by degrading ECM proteins including collagens (Kurihara *et al*, 2009). Thus, altered collagen metabolism may play an important role in anoikis regulation in OSCC.

The other major adhesive ECM proteins, laminin, fibronectin, and tenascin have been closely linked to anoikis regulation (Ziober et al, 2001; Zhang et al, 2004b; Dai et al, 2005; Boisvert-Adamo and Aplin, 2006; Kamarajan and Kapila, 2007; Lange et al, 2007; Ordonez et al. 2007). Laminin-5, which is present in normal epithelial tissues and the primary component of the basement membrane of epithelium, is increased in SCC (Ziober et al, 2001). Overexpression of laminin-5 correlates with increased tumor invasiveness and motility of OSCC, and this increased motility is thought to be controlled and enhanced by $\alpha 2\beta 1$ and $\alpha 3\beta 1$ integrins (Decline and Rousselle, 2001). Fibronectin is a multiadhesive matrix protein with domains that bind collagen/gelatin, fibrin, and heparin and regions that undergo alternatively splicing, which bind integrin and proteoglycan receptors. Fragments of fibronectin containing an alternatively spliced V region and function-perturbing point mutations in the high-affinity heparin-binding domain disrupt cell adhesion to the ECM and mediate invasion, motility and anoikis of OSCC cells (Kapila et al, 1997; Kamarajan and Kapila, 2007). Laminin and fibronectin are key ECM adhesion glycoprotein components involved in OSCC anoikis regulation.

Integrin receptors

Integrins are alpha and beta heterodimeric receptor complexes that connect ECM components to intracellular signaling pathways. Integrins regulate cell adhesion, migration, survival, and anoikis resistance by activating complex signaling networks (Mitra *et al*, 2005). Integrin suppression leads to anoikis and apoptosis in human mammary epithelial cells (Haenssen *et al*, 2010). Overexpression of integrin α 4 increases phosphorylation of its downstream mediator, focal adhesion kinase (FAK), and suppresses anoikis in primary fibroblasts (Joo *et al*, 2008). In addition, inhibiting the β 1 integrin induces anoikis in epithelial cells (Bouchard *et al*, 2007). Several integrins recognize fibronectin and are present on the surface of OSCC cells, including $\alpha 4\beta 1$, $\alpha 5\beta 1$, and $\alpha \nu \beta 1$ (Zhang *et al*, 2004a). Suppression of integrin $\alpha \nu$ and its downstream phosphorylation of FAK and ERK induces anoikis in human OSCC cells (Zhang *et al*, 2004b; Kamarajan and Kapila, 2007). Integrin switching from $\alpha \nu \beta 5$ to $\alpha \nu \beta 6$ protects OSCC cells from anoikis (Janes and Watt, 2004). Furthermore, integrin $\alpha \nu \beta 6$ and the enzyme cyclooxygenase-2 are implicated in OSCC progression and have been suggested as possible therapeutic targets (Nystrom *et al*, 2006).

Focal adhesion kinase, a key regulator of integrin signaling, interacts with p130cas, paxillin, talin, Src, phosphoinositide 3-kinase, Shc, ERK, and p53 to regulate cell survival and anoikis resistance (Kapila et al, 1999; Janes and Watt, 2004; Shiraki et al, 2005; Kamarajan and Kapila, 2007; Koschny et al, 2007; Sakamoto et al, 2010). Disrupting FAK signaling and genetic deletion of FAK induce anoikis and apoptosis in cells (Taylor et al. 2008: Schwock et al. 2009). Furthermore, anoikis is associated with reduced levels of FAK phosphorylation in multiple cell types including OSCC (Frisch et al, 1996; Valentinis et al, 1998; Zhang et al, 2004b; Masuda et al, 2005; Kamarajan and Kapila, 2007; Johnson et al, 2008; Wendt et al, 2008; Noda et al, 2009) and some cell types seem to undergo FAKindependent anoikis (Wei et al, 2004; Diaz-Montero et al, 2006). Conversely, FAK overexpression is present in numerous human malignancies, including SCC, with the degree of overexpression correlating with the magnitude of aggressiveness; demonstrating a protective role for FAK in apoptosis (Aronsohn et al, 2003; Jiang et al, 2010). FAK activates multiple signaling pathways that may fine tune cell-type-specific phenotypes and cell survival (Zoug et al, 2009). For example, treatment of OSCC cells with safingol, a protein kinase C (PKC) inhibitor, leads to a rapid decrease in FAK phosphorvlation, a subsequent decrease in FAK protein levels, and anoikis; suggesting that FAK suppression via PKC inhibition contributes to safingol-induced anoikis (Noda et al, 2009). Emerging data from early-phase cancer clinical trials with orally available small-molecule inhibitors of FAK are promising (Schultze and Fiedler, 2010). Integrin receptors and their downstream signaling pathways, including FAK, show promise as anoikis regulatory targets.

Proteoglycan receptors

Proteoglycan receptors also regulate cell survival and apoptosis, and alterations in these receptors are associated with tumorigenesis (Misra *et al*, 2008). CD44 is a multifunctional protein involved in cell adhesion, migration, apoptosis, and drug resistance. CD44 is a primary receptor for hyaluronan (HA), a major component of the ECM, and it plays a critical role in cell signaling and cell–ECM interactions in cancer (Ponta *et al*, 2003; Hauptschein *et al*, 2005; Hao *et al*, 2010). CD44 undergoes alternative splicing and several of its spliced variants are associated with tumorigenesis. In fact, expression of various CD44 protein variants

correlates with aggressive human cancers, including head and neck SCC and breast cancer (To et al, 2010). Overexpression of CD44 is associated with apoptotic resistance and down regulation of CD44 is associated with anoikis, as is the case in head and neck SCC (Roehlecke et al, 2000; Harper et al, 2010). CD44 is also believed to be a stem cell marker in head and neck tumors as in other cancers (Ailles and Prince, 2009; Harper et al, 2010). Inhibition of the proteoglycan receptor CD44s inhibits breast cancer cell adhesion, motility, and invasion (Afify et al, 2009). Downregulation of CD44 is associated with tumor metastasis in OSCC cells and treatment with an anti-CD44 antibody enhances the invasive potential of OSCC cell lines (Sato et al. 2004). Furthermore, OSCC patients demonstrating irregular staining and expression of CD44 exhibited more advanced disease and shortened survival (Kosunen et al, 2007). Monoclonal antibodies (BIWA 4, bivatuzumab) against specific CD44 isoforms have been tested in patients with HNSCCs for potential imaging or targeting therapy against tumors (Lyons and Jones, 2007). A clinical trial using anti-CD44v6 antibodies to treat head and neck cancer showed promise but the death of a patient suspended the trial (Orian-Rousseau, 2010). CD44 expression plays an important role in the behavior of malignant tumors, and its variants may be future targets for OSCC anoikis regulation and treatment

Another family of proteoglycan receptors of potential significance in anoikis regulation are the syndecans as they modulate cancer cell apoptosis and survival (Choi et al, 2009; Shimada et al, 2009). Syndecans interact with a number of ECM ligands including fibroblast growth factor, vascular endothelial growth factor, transforming growth factor-beta, epithelial growth factor, and fibronectin (Tkachenko et al, 2005). In many cell models, overexpression of syndecans enhances cell adhesion and migration in normal and neoplastic cells, whereas interference with syndecan function decreases cell migration, consequently enhancing cell spreading (Choi et al, 2009; Khotskaya et al, 2009). Alterations in syndecan-2 levels can also regulate cancer cell apoptosis (Orosco et al, 2007). Like CD44, syndecans interact with integrins and have multiple adhesion and cosignaling functions, but their mechanistic contribution to anoikis remains poorly understood. Proteoglycan receptors regulate key tumorgenic functions in OSCC progression, and are therefore promising therapeutic targets for regulation of cancer cell anoikis regulation.

Death receptors and their ligands

Apoptosis is regulated by both intrinsic and extrinsic cell death pathways. The intrinsic pathway is mediated by intracellular signals induced by DNA damage or cytokine deprivation leading to caspase activation and cell death (Green, 2000). In the extrinsic pathway, cell surface death receptors, such as Fas/CD95, TNFR1, TNFR2, DR4, and DR5, are characterized by the presence of a death domain and are activated by extracellular death ligands. Activation or oligomerization of death receptors leads

to formation of a death-inducing signaling complex (DISC) and subsequent caspase activation that mediates cell death (Sharma *et al*, 2000; Taylor *et al*, 2008).

Anoikis can be mediated by activation of the death receptor pathway and caspase activation (Frisch and Screaton, 2001). The CD95/Fas pathway has been implicated in chemotherapy-induced tumor cell death in a number of studies. Treatment with anticancer drugs trigger an increase in CD95/Fas ligand (CD95/FasL) expression, which stimulates the Fas receptor pathway in an autocrine or paracrine manner, whereby FasL binds its receptor (Fulda and Debatin, 2004). Fas receptors are expressed in both normal and tumor cells, including OSCC cells (Iwase et al, 2003; Ozoren and El-Deiry, 2003). Up-regulation of FasL and downregulation of Fas expression are early and frequent events associated with the evolution of esophageal SCC (Gratas et al, 1998). Furthermore, Fas is expressed in low quantities in OSCC and FasL expression correlates negatively with the degree of differentiation and apoptosis in OSCC (Loro et al, 1999). OSCC cells undergo apoptosis in response to treatment with FasL or Fas antibody (Moers et al, 1999). Anoikis triggered by activation of Fas pathways in OSCC could be an important defense mechanism against tumorigenesis (Kamarajan et al, 2010). However, these approaches have not been applied in OSCC clinical trails (Papenfuss et al, 2008).

Tumor necrosis factor alpha (TNF-alpha) is a multifunctional cytokine involved in apoptosis, anoikis, cell survival, inflammation, immunity, and cell signaling through two different death receptor subtypes, TNFR1 and TNFR2 (Simonitsch and Krupitza, 1998; Locksley *et al*, 2001; Engbers-Buijtenhuijs *et al*, 2005). TNFR1 forms a DISC similar to Fas, involving TRADD, FADD, and procaspase-8. Mice lacking TNFR1 and TNFR2 are resistant to death and liver injury induced by an anti-Fas antibody (Costelli *et al*, 2003). Furthermore, the cell surface receptor TNFR1 has been detected only in small numbers in OSCC (Gupta *et al*, 2008).

TNF-related apoptosis-inducing ligand (TRAIL) is a cytokine that interacts with death receptors DR4 and DR5 to facilitate the selective elimination of malignant cells through the induction of apoptosis (LeBlanc and Ashkenazi, 2003; Mahmood and Shukla, 2010). DR5, and to a lesser extent DR4, mediates anoikis in human colorectal carcinoma cell lines (Shiraki et al, 2005; Laguinge et al, 2008). Cancer cells are more susceptible than normal cells to apoptosis induction by TRAIL, and combinations of TRAIL and chemotherapeutics can act synergistically to kill tumor cells (Koschny et al, 2007). Primary cells from OSCC are sensitive to TRAIL, but metastatic cell lines of OSCC are resistant to TRAIL exposure (Noutomi et al, 2009). Therapeutic agents targeting TRAIL and its receptors have been developed for clinical application. TRAIL receptor agonists, including recombinant human TRAIL and an agonistic monoclonal antibody against DR4 and DR5, are currently being tested in phase I and II clinical trails (Bellail et al, 2009). These results suggest TRAIL resistance imparts metastatic capacity to primary OSCC tumors, and therapeutic agents targeting TRAIL hold promise for OSCC.

Death ligands are critical for control of tissue homeostasis and spread of primary OSCC, and apoptosis by death receptors is considered an important defense mechanism against metastasis that may also be important for anoikis resistance.

Cell survival and cell death receptor crosstalk

Receptor interacting protein (RIP), a kinase with homology to both serine/threonine and tyrosine kinases, interacts with both death receptors and FAK survival pathways and is found in many tissues (Meylan and Tschopp, 2005). RIP is activated and cleaved upon treatment with TNF, Fas, or TRAIL in cell death pathways (Kim et al, 2000). Once activated, RIP interacts with both the death receptors TNFR1, TRAF2, FADD, and TRADD, and epithelial growth factor receptors, subsequently playing a key role in inhibiting the activation of NF-kB, thus potentiating apoptosis (Habib et al, 2001; Meylan and Tschopp, 2005). RIP also contributes to TNF-induced JNK, ERK, and p38 MAPK kinase activation (Lee et al, 2003). Furthermore, RIP seems to play a dual role, as it can bind to FAK in human tumor cells providing a prosurvival signal, and it can interact with Fas in cell death pathways to initiate anoikis (Kurenova et al, 2004; Meylan and Tschopp, 2005). Therefore, RIP appears to participate in both prosurvival and cell death pathways, and thus, could constitute a possible target for modulating OSCC anoikis resistance. Indeed, RIP was recently found to mediate crosstalk between the Fas/CD95 death receptor and integrin/FAK signaling pathways in regulating anoikis. Under anoikis conditions, RIP forms a complex with Fas while dissociating from FAK, whereas under survival conditions, it dissociates from Fas and complexes with FAK. RIP is a key shuttling protein that communicates with both integrin/FAK survival signals and Fas/death signals in anoikis (Figure 2) (Kamarajan et al, 2010), and may thus serve as a potential target to modulate anoikis.

Summary and future directions

Oral squamous cell carcinoma treatments have shown limited success in the clinical setting in recent decades. Currently, there are many new promising potential targets for cancer therapy and OSCC prognostic markers. Regulation of anoikis and apoptosis pathways in tumor cells, including OSCC involves multiple ECM proteins, cell surface receptors and intracellular signaling targets, and these may be cell-type specific and may serve as therapeutic targets for tumorigenesis. The ability to target both death receptor pathways and prosurvival pathways concurrently seems a promising approach in anticancer therapy. Existing potential targets need to be examined to develop more successful therapeutic tools to provide vital and necessary improvements in cancer treatment.

Anoikis mediators in oral squamous cell carcinoma | Bunek et al

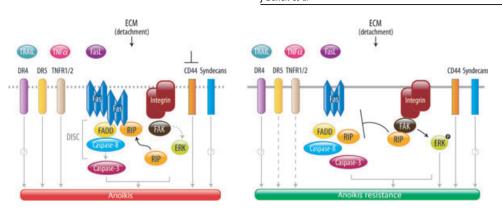


Figure 2 Signaling pathways that mediate anoikis and anoikis resistance. Loss of cell–extracellular matrix (ECM) contacts (detachment) and dissociation of integrin and focal adhesion kinase (FAK) fails to activate survival signals via molecules that include extracellular-signal-regulated kinase (ERK). In death receptor-mediated anoikis mechanisms, FAK and receptor-interacting protein (RIP) dissociate, leading to association of RIP with Fas, formation of the death-inducing signaling complex (DISC), activation of caspase-3, and propagation of anoikis. The DISC is comprised of Fas, FADD, and caspase-8. Anoikis is further activated by an increase in the cell surface expression of Fas and DR5 receptors. The role of the death receptor ligands, TRAIL, TNF- α , and FasL in death receptor activation during anoikis is still unclear. Inhibition of proteoglycan receptors, like CD44 promotes anoikis processes. The role of other proteoglycans, like Syndecans in anoikis regulation is still unclear. Upon detachment from the ECM, cells are capable of evading anoikis and acquire the ability to survive thereby becoming anoikis resistant. This mechanism is achieved through the involvement of constitutive activity of survival pathways that include integrin-dependent activation of FAK and subsequent ERK phosphorylation. In addition, FAK and RIP associate, thereby inhibiting the association of RIP with Fas and formation of the DISC, which favors cell survival. Up-regulation of CD44 also contributes to survival signals and promotes anoikis resistance. Cancer cells, including oral squamous cell carcinoma cells exhibit the unique property of anoikis resistance when detached from their ECM

Acknowledgements

We truly regret that we could not cite the seminal work of many of our colleagues owing to space limitations. This work was supported by Grant R01 DE014429 from the National Institutes of Health to YLK.

References

- Afify A, Purnell P, Nguyen L (2009). Role of CD44s and CD44v6 on human breast cancer cell adhesion, migration, and invasion. *Exp Mol Pathol* **86**: 95–100.
- Ailles L, Prince M (2009). Cancer stem cells in head and neck squamous cell carcinoma. *Methods Mol Biol* 568: 175–193.
- Aronsohn MS, Brown HM, Hauptman G, Kornberg LJ (2003). Expression of focal adhesion kinase and phosphorylated focal adhesion kinase in squamous cell carcinoma of the larynx. *Laryngoscope* **113**: 1944–1948.
- Balsara BR, Testa JR (2002). Chromosomal imbalances in human lung cancer. *Oncogene* **21**: 6877–6883.
- Bellail AC, Qi L, Mulligan P, Chhabra V, Hao C (2009). TRAIL agonists on clinical trials for cancer therapy: the promises and the challenges. *Rev Recent Clin Trials* **4**: 34–41.
- Bockmuhl U, Petersen S, Schmidt S *et al* (1997). Patterns of chromosomal alterations in metastasizing and nonmetastasizing primary head and neck carcinomas. *Cancer Res* **57**: 5213–5216.
- Boisvert-Adamo K, Aplin AE (2006). B-RAF and PI-3 kinase signaling protect melanoma cells from anoikis. *Oncogene* **25**: 4848–4856.
- Bouchard V, Demers MJ, Thibodeau S *et al* (2007). Fak/Src signaling in human intestinal epithelial cell survival and anoikis: differentiation state-specific uncoupling with the PI3-K/Akt-1 and MEK/Erk pathways. *J Cell Physiol* **212**: 717–728.
- Bozzo C, Sabbatini M, Tiberio R *et al* (2006). Activation of caspase-8 triggers anoikis in human neuroblastoma cells. *Neurosci Res* 56: 145–153.

- Chiarugi P (2008). From anchorage dependent proliferation to survival: lessons from redox signalling. *IUBMB Life* **60**: 301–307.
- Choi S, Kim Y, Park H et al (2009). Syndecan-2 overexpression regulates adhesion and migration through cooperation with integrin alpha2. Biochem Biophys Res Commun **384**: 231–235.
- Costelli P, Aoki P, Zingaro B *et al* (2003). Mice lacking TNFalpha receptors 1 and 2 are resistant to death and fulminant liver injury induced by agonistic anti-Fas anti-body. *Cell Death Differ* **10**: 997–1004.
- Dai R, Iwama A, Wang S, Kapila YL (2005). Diseaseassociated fibronectin matrix fragments trigger anoikis of human primary ligament cells: p53 and c-myc are suppressed. *Apoptosis* 10: 503–512.
- Decline F, Rousselle P (2001). Keratinocyte migration requires alpha2beta1 integrin-mediated interaction with the laminin 5 gamma2 chain. *J Cell Sci* **114**: 811–823.
- Diaz-Montero CM, Wygant JN, McIntyre BW (2006). PI3-K/Akt-mediated anoikis resistance of human osteosarcoma cells requires Src activation. *Eur J Cancer* 42: 1491–1500.
- Eble JA, Haier J (2006). Integrins in cancer treatment. Curr Cancer Drug Targets 6: 89–105.
- Engbers-Buijtenhuijs P, Kamphuis M, van der Sluijs Veer G et al (2005). A novel time resolved fluorometric assay of anoikis using Europium-labelled Annexin V in cultured adherent cells. *Apoptosis* **10:** 429–437.
- Frisch SM, Francis H (1994). Disruption of epithelial cellmatrix interactions induces apoptosis. J Cell Biol 124: 619– 626.
- Frisch SM, Screaton RA (2001). Anoikis mechanisms. Curr Opin Cell Biol 13: 555–562.
- Frisch SM, Vuori K, Ruoslahti E, Chan-Hui PY (1996). Control of adhesion-dependent cell survival by focal adhesion kinase. J Cell Biol 134: 793–799.
- Fulda S, Debatin KM (2004). Exploiting death receptor signaling pathways for tumor therapy. *Biochim Biophys Acta* **1705**: 27–41.
- Garrison JB, Kyprianou N (2004). Novel targeting of apoptosis pathways for prostate cancer therapy. *Curr Cancer Drug Targets* **4:** 85–95.

359

- Giannelli G, Milillo L, Marinosci F *et al* (2001). Altered expression of integrins and basement membrane proteins in malignant and pre-malignant lesions of oral mucosa. *J Biol Regul Homeost Agents* **15:** 375–380.
 - Gniadecki R (1998). Regulation of keratinocyte proliferation. *Gen Pharmacol* **30:** 619–622.
 - Gratas C, Tohma Y, Barnas C *et al* (1998). Up-regulation of Fas (APO-1/CD95) ligand and down-regulation of Fas expression in human esophageal cancer. *Cancer Res* **58**: 2057–2062.
 - Green DR (2000). Apoptotic pathways: paper wraps stone blunts scissors. *Cell* **102:** 1–4.
 - Gupta R, Sharma SC, Das SN (2008). Association of TNF-alpha and TNFR1 promoters and 3' UTR region of TNFR2 gene polymorphisms with genetic susceptibility to tobacco-related oral carcinoma in Asian Indians. *Oral Oncol* **44:** 455–463.
- Habib AA, Chatterjee S, Park SK *et al* (2001). The epidermal growth factor receptor engages receptor interacting protein and nuclear factor-kappa B (NF-kappa B)inducing kinase to activate NF-kappa B. Identification of a novel receptor-tyrosine kinase signalosome. *J Biol Chem* **276:** 8865–8874.
- Haenssen KK, Caldwell SA, Shahriari KS *et al* (2010). ErbB2 requires integrin alpha 5 for anoikis resistance via Src regulation of receptor activity in human mammary epithelial cells. *J Cell Sci* **123**: 1373–1382.
- Hao JL, Cozzi PJ, Khatri A, Power CA, Li Y (2010). CD147/EMMPRIN and CD44 are potential therapeutic targets for metastatic prostate cancer. *Curr Cancer Drug Targets* **10**: 287–306.
- Harper LJ, Costea DE, Gammon L *et al* (2010). Normal and malignant epithelial cells with stem-like properties have an extended G2 cell cycle phase that is associated with apoptotic resistance. *BMC Cancer* **10**: 166.
- Hauptschein RS, Sloan KE, Torella C *et al* (2005). Functional proteomic screen identifies a modulating role for CD44 in death receptor-mediated apoptosis. *Cancer Res* **65**: 1887–1896.
- Ishida H, Wada K, Masuda T *et al* (2007). Critical role of estrogen receptor on anoikis and invasion of squamous cell carcinoma. *Cancer Sci* **98:** 636–643.
- Iwase M, Watanabe H, Kondo G, Ohashi M, Nagumo M (2003). Enhanced susceptibility of oral squamous cell carcinoma cell lines to FAS-mediated apoptosis by cisplatin and 5-fluorouracil. *Int J Cancer* **106**: 619–625.
- Janes SM, Watt FM (2004). Switch from alphavbeta5 to alphavbeta6 integrin expression protects squamous cell carcinomas from anoikis. *J Cell Biol* **166**: 419–431.
- Jemal A, Siegel R, Ward E *et al* (2008). Cancer statistics, 2008. *CA Cancer J Clin* **58**: 71–96.
- Jiang H, Liu L, Ye J *et al* (2010). Focal adhesion kinase serves as a marker of cervical lymph node metastasis and is a potential therapeutic target in tongue cancer. *J Cancer Res Clin Oncol* **136**: 1295–1302.
- Johnson TR, Khandrika L, Kumar B et al (2008). Focal adhesion kinase controls aggressive phenotype of androgenindependent prostate cancer. Mol Cancer Res 6: 1639–1648.
- Joo NE, Watanabe T, Chen C *et al* (2008). NG2, a novel proapoptotic receptor, opposes integrin alpha4 to mediate anoikis through PKCalpha-dependent suppression of FAK phosphorylation. *Cell Death Differ* **15**: 899–907.
- Kamarajan P, Kapila YL (2007). An altered fibronectin matrix induces anoikis of human squamous cell carcinoma cells by suppressing integrin alpha v levels and phosphorylation of FAK and ERK. *Apoptosis* 12: 2221–2231.
- Kamarajan P, Bunek J, Lin Y, Nunez G, Kapila YL (2010). Receptor-interacting protein shuttles between cell death and survival signaling pathways. *Mol Biol Cell* 21: 481–488.

- Kapila YL, Niu J, Johnson PW (1997). The high affinity heparin-binding domain and the V region of fibronectin mediate invasion of human oral squamous cell carcinoma cells *in vitro*. *J Biol Chem* **272**: 18932–18938.
- Kapila YL, Wang S, Johnson PW (1999). Mutations in the heparin binding domain of fibronectin in cooperation with the V region induce decreases in pp125(FAK) levels plus proteoglycan-mediated apoptosis via caspases. *J Biol Chem* 274: 30906–30913.
- Khotskaya YB, Dai Y, Ritchie JP *et al* (2009). Syndecan-1 is required for robust growth, vascularization, and metastasis of myeloma tumors *in vivo*. J Biol Chem **284**: 26085–26095.
- Kim JW, Choi EJ, Joe CO (2000). Activation of deathinducing signaling complex (DISC) by pro-apoptotic C-terminal fragment of RIP. *Oncogene* **19:** 4491–4499.
- Koschny R, Ganten TM, Sykora J *et al* (2007). TRAIL/bortezomib cotreatment is potentially hepatotoxic but induces cancer-specific apoptosis within a therapeutic window. *Hepatology* **45**: 649–658.
- Kosunen A, Pirinen R, Ropponen K *et al* (2007). CD44 expression and its relationship with MMP-9, clinicopathological factors and survival in oral squamous cell carcinoma. *Oral Oncol* **43:** 51–59.
- Kupferman ME, Patel V, Sriuranpong V *et al* (2007). Molecular analysis of anoikis resistance in oral cavity squamous cell carcinoma. *Oral Oncol* **43**: 440–454.
- Kurenova E, Xu LH, Yang X *et al* (2004). Focal adhesion kinase suppresses apoptosis by binding to the death domain of receptor-interacting protein. *Mol Cell Biol* **24**: 4361–4371.
- Kurihara Y, Hatori M, Ando Y *et al* (2009). Inhibition of cyclooxygenase-2 suppresses the invasiveness of oral squamous cell carcinoma cell lines via down-regulation of matrix metalloproteinase-2 production and activation. *Clin Exp Metastasis* **26:** 425–432.
- Laguinge LM, Samara RN, Wang W *et al* (2008). DR5 receptor mediates anoikis in human colorectal carcinoma cell lines. *Cancer Res* **68**: 909–917.
- Lange K, Kammerer M, Hegi ME *et al* (2007). Endothelin receptor type B counteracts tenascin-C-induced endothelin receptor type A-dependent focal adhesion and actin stress fiber disorganization. *Cancer Res* **67**: 6163–6173.
- LeBlanc HN, Ashkenazi A (2003). Apo2L/TRAIL and its death and decoy receptors. *Cell Death Differ* **10**: 66–75.
- Lee TH, Huang Q, Oikemus S *et al* (2003). The death domain kinase RIP1 is essential for tumor necrosis factor alpha signaling to p38 mitogen-activated protein kinase. *Mol Cell Biol* **23**: 8377–8385.
- Locksley RM, Killeen N, Lenardo MJ (2001). The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell* **104:** 487–501.
- Loro LL, Vintermyr OK, Johannessen AC, Liavaag PG, Jonsson R (1999). Suppression of Fas receptor and negative correlation of Fas ligand with differentiation and apoptosis in oral squamous cell carcinoma. *J Oral Pathol Med* **28**: 82–87.
- Lyons AJ, Jones J (2007). Cell adhesion molecules, the extracellular matrix and oral squamous carcinoma. *Int J Oral Maxillofac Surg* **36**: 671–679.
- Mahmood Z, Shukla Y (2010). Death receptors: targets for cancer therapy. *Exp Cell Res* **316**: 887–899.
- Masuda T, Wada K, Nakajima A *et al* (2005). Critical role of peroxisome proliferator-activated receptor gamma on anoikis and invasion of squamous cell carcinoma. *Clin Cancer Res* **11**: 4012–4021.
- Meylan E, Tschopp J (2005). The RIP kinases: crucial integrators of cellular stress. *Trends Biochem Sci* **30**: 151–159.

360

- Misra S, Obeid LM, Hannun YA *et al* (2008). Hyaluronan constitutively regulates activation of COX-2-mediated cell survival activity in intestinal epithelial and colon carcinoma cells. *J Biol Chem* **283**: 14335–14344.
- Mitra SK, Hanson DA, Schlaepfer DD (2005). Focal adhesion kinase: in command and control of cell motility. *Nat Rev Mol Cell Biol* **6**: 56–68.
- Moers C, Warskulat U, Muschen M *et al* (1999). Regulation of CD95 (Apo-1/Fas) ligand and receptor expression in squamous-cell carcinoma by interferon-gamma and cisplatin. *Int J Cancer* **80**: 564–572.
- Neiva KG, Zhang Z, Miyazawa M *et al* (2009). Cross talk initiated by endothelial cells enhances migration and inhibits anoikis of squamous cell carcinoma cells through STA-T3/Akt/ERK signaling. *Neoplasia* **11**: 583–593.
- Noda T, Iwai S, Hamada M, Fujita Y, Yura Y (2009). Induction of apoptosis of detached oral squamous cell carcinoma cells by safingol. Possible role of Bim, focal adhesion kinase and endonuclease G. *Apoptosis* 14: 287– 297.
- Noutomi T, Itoh M, Toyota H, Takada E, Mizuguchi J (2009). Tumor necrosis factor-related apoptosis-inducing ligand induces apoptotic cell death through c-Jun NH2-terminal kinase activation in squamous cell carcinoma cells. *Oncol Rep* 22: 1169–1172.
- Nystrom ML, McCulloch D, Weinreb PH et al (2006). Cyclooxygenase-2 inhibition suppresses alphavbeta6 integrin-dependent oral squamous carcinoma invasion. Cancer Res 66: 10833–10842.
- Ordonez C, Zhai AB, Camacho-Leal P *et al* (2007). GPIanchored CEA family glycoproteins CEA and CEACAM6 mediate their biological effects through enhanced integrin alpha5beta1-fibronectin interaction. *J Cell Physiol* **210**: 757– 765.
- Orian-Rousseau V (2010). CD44, a therapeutic target for metastasising tumours. *Eur J Cancer* **46**: 1271–1277.
- Orosco A, Fromigue O, Bazille C *et al* (2007). Syndecan-2 affects the basal and chemotherapy-induced apoptosis in osteosarcoma. *Cancer Res* **67**: 3708–3715.
- Ozoren N, El-Deiry WS (2003). Cell surface death receptor signaling in normal and cancer cells. *Semin Cancer Biol* **13**: 135–147.
- Papenfuss K, Cordier SM, Walczak H (2008). Death receptors as targets for anti-cancer therapy. J Cell Mol Med 12: 2566– 2585.
- Ponta H, Sherman L, Herrlich PA (2003). CD44: from adhesion molecules to signalling regulators. *Nat Rev Mol Cell Biol* **4:** 33–45.
- Roehlecke C, Kuhnt AK, Fehrenbach H *et al* (2000). Resistance of L132 lung cell clusters to glyoxal-induced apoptosis. *Histochem Cell Biol* **114**: 283–292.
- Sakamoto S, McCann RO, Dhir R, Kyprianou N (2010). Talin1 promotes tumor invasion and metastasis via focal adhesion signaling and anoikis resistance. *Cancer Res* **70**: 1885–1895.
- Sato S, Miyauchi M, Kato M *et al* (2004). Upregulated CD44v9 expression inhibits the invasion of oral squamous cell carcinoma cells. *Pathobiology* **71**: 171–175.
- Schultze A, Fiedler W (2010). Therapeutic potential and limitations of new FAK inhibitors in the treatment of cancer. *Expert Opin Investig Drugs* **19**: 777–788.
- Schwock J, Dhani N, Cao MP *et al* (2009). Targeting focal adhesion kinase with dominant-negative FRNK or Hsp90 inhibitor 17-DMAG suppresses tumor growth and metastasis of SiHa cervical xenografts. *Cancer Res* **69**: 4750–4759.

- Shanmugathasan M, Jothy S (2000). Apoptosis, anoikis and their relevance to the pathobiology of colon cancer. *Pathol Int* **50**: 273–279.
- Sharma K, Wang RX, Zhang LY *et al* (2000). Death the Fas way: regulation and pathophysiology of CD95 and its ligand. *Pharmacol Ther* **88**: 333–347.
- Shimada Y, Ishii G, Nagai K *et al* (2009). Expression of podoplanin, CD44, and p63 in squamous cell carcinoma of the lung. *Cancer Sci* **100**: 2054–2059.
- Shiraki K, Yamanaka T, Inoue H *et al* (2005). Expression of TNF-related apoptosis-inducing ligand in human hepatocellular carcinoma. *Int J Oncol* **26**: 1273–1281.
- Simonitsch I, Krupitza G (1998). Autocrine self-elimination of cultured ovarian cancer cells by tumour necrosis factor alpha (TNF-alpha). *Br J Cancer* **78**: 862–870.
- Stetler-Stevenson WG (1996). Dynamics of matrix turnover during pathologic remodeling of the extracellular matrix. *Am J Pathol* 148: 1345–1350.
- Streuli CH, Gilmore AP (1999). Adhesion-mediated signaling in the regulation of mammary epithelial cell survival. J Mammary Gland Biol Neoplasia 4: 183–191.
- Swan EA, Jasser SA, Holsinger FC *et al* (2003). Acquisition of anoikis resistance is a critical step in the progression of oral tongue cancer. *Oral Oncol* **39:** 648–655.
- Tang MKS, Zhou HY, Yam JWP, Wong AST (2010). c-Met overexpression contributes to the acquired apoptotic resistance of nonadherent ovarian cancer cells through a cross talk mediated by phosphatidylinositol 3-kinase and extracellular signal-regulated kinase 1/2. *Neoplasia* 12: 128–138.
- Taylor RC, Cullen SP, Martin SJ (2008). Apoptosis: controlled demolition at the cellular level. *Nat Rev Mol Cell Biol* 9: 231–241.
- Tkachenko E, Rhodes JM, Simons M (2005). Syndecans: new kids on the signaling block. *Circ Res* **96**: 488–500.
- To K, Fotovati A, Reipas KM *et al* (2010). Y-box binding protein-1 induces the expression of CD44 and CD49f leading to enhanced self-renewal, mammosphere growth, and drug resistance. *Cancer Res* **70**: 2840–2851.
- Valentinis B, Reiss K, Baserga R (1998). Insulin-like growth factor-I-mediated survival from anoikis: role of cell aggregation and focal adhesion kinase. J Cell Physiol 176: 648–657.
- Wei L, Yang Y, Zhang X, Yu Q (2004). Altered regulation of Src upon cell detachment protects human lung adenocarcinoma cells from anoikis. *Oncogene* 23: 9052–9061.
- Wendt MK, Drury LJ, Vongsa RA, Dwinell MB (2008). Constitutive CXCL12 expression induces anoikis in colorectal carcinoma cells. *Gastroenterology* **135**: 508–517.
- Zhang Y, Lu H, Dazin P, Kapila Y (2004a). Functional differences between integrin alpha4 and integrins alpha5/ alphav in modulating the motility of human oral squamous carcinoma cells in response to the V region and heparin-binding domain of fibronectin. *Exp Cell Res* **295**: 48–58.
- Zhang Y, Lu H, Dazin P, Kapila Y (2004b). Squamous cell carcinoma cell aggregates escape suspension-induced, p53-mediated anoikis: fibronectin and integrin alphav mediate survival signals through focal adhesion kinase. *J Biol Chem* **279**: 48342–48349.
- Ziober BL, Silverman SS Jr, Kramer RH (2001). Adhesive mechanisms regulating invasion and metastasis in oral cancer. *Crit Rev Oral Biol Med* **12**: 499–510.
- Zouq NK, Keeble JA, Lindsay J *et al* (2009). FAK engages multiple pathways to maintain survival of fibroblasts and epithelia: differential roles for paxillin and p130Cas. *J Cell Sci* **122**: 357–367.

Copyright of Oral Diseases is the property of Wiley-Blackwell and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.