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### INVITED MEDICAL REVIEW

# Mechanisms involved in the association between peridontal diseases and cardiovascular disease

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It is now well accepted that besides the cholesterol associated mechanisms of atherogenesis, inflammation plays a crucial role in all stages of the development of the atherosclerotic lesion. This 'inflammation hypothesis' raises the possibility that through systemic elevations of pro-inflammatory cytokines, periodontal diseases might also contribute to systemic inflammation and, therefore, to atherogenesis. In fact, there is evidence that periodontal diseases are associated with higher systemic levels of high-sensitivity C-reactive protein and a low grade systemic inflammation. This phenomenon has been explained based on mechanisms associated with either the infectious or the inflammatory nature of periodontal diseases. The purposes of this article were to review (1) the evidence suggesting a role for oral bacterial species, particularly periodontal pathogens, in atherogenesis; (2) the potential mechanisms explaining an etiological role for oral bacteria in atherosclerosis; (3) the evidence suggesting that periodontal infections are accompanied by a heightened state of systemic inflammation; (4) the potential sources of systemic inflammatory biomarkers associated with periodontal diseases; and (5) the effects of periodontal therapy on systemic inflammatory biomarkers and cardiovascular risk. Oral Diseases (2011) 17, 450-461

**Keywords:** cardiovascular disease; atherosclerosis; periodontal diseases; infection; periodontal pathogens; bacteremia; inflammatory response; systemic biomarkers; C-reactive protein

#### Introduction

Since Mattila *et al* (Mattila *et al*, 1989) reported the first study examining the association between cardiovascular disease (CVD) and dental infections in 1989, the literature has been flooded with reports examining the

Correspondence: Ricardo Teles, Department of Periodontology, The Forsyth Institute, 245 First Street, Cambridge, MA 02142. Tel: 617-892-8556, Fax: 617-262-4021, E-mail: rteles@forsyth.org strength of this association and its biological plausibility. Several epidemiological studies have been conducted examining a possible association between periodontal disease and CVD (DeStefano *et al*, 1993; Mattila *et al*, 1995; Beck *et al*, 1996; Joshipura *et al*, 1996; Morrison *et al*, 1999; Hujoel *et al*, 2000; Wu *et al*, 2000; Howell *et al*, 2001; Ajwani *et al*, 2003; Tuominen *et al*, 2003; Hung *et al*, 2004) and three meta-analyses have summarized their findings (Janket *et al*, 2003; Mustapha *et al*, 2007) (Humphrey *et al*, 2008). These meta-analyses have consistently concluded that the available evidence indicates that periodontal diseases confer a moderate risk for atherosclerosis and its consequences (Janket *et al*, 2003; Mustapha *et al*, 2007; Humphrey *et al*, 2008).

Because periodontal diseases are infections/inflammatory diseases, mechanisms mediated by oral microorganisms and the inflammation triggered by them have been proposed to explain their involvement in atherogenesis. Bacteria associated with periodontal diseases can colonize the atheromatous plaques and could cause their damage by inducing local inflammation, resulting in propagation of the inflammatory events that lead to atheroma formation, development and eventual rupture. Alternatively, a low grade systemic inflammation could result from bacteremias, or as a consequence of proinflammatory cytokines generated at the site of the periodontal lesion gaining access to the blood stream. In fact, the potential role of infection in the etiology of CVD is not new and over the years several different microbial agents have been investigated for their role in atherosclerosis. Similarly, diseases associated with a heightened state of systemic inflammation such as rheumatoid arthritis (RA) have also been associated with atherosclerosis and its acute complications. The purpose of this manuscript was to review the infectious and inflammatory mechanisms proposed to explain the link between periodontal diseases and CVD, and to appraise the evidence supporting them.

#### Inflammatory mechanisms of atherogenesis

The normal endothelium is non-adherent to circulating leukocytes. Once exposed to inflammatory stimuli,

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endothelial cells increase the expression of adhesion molecules such as inter-cellular adhesion molecule 1 (ICAM-1), vascular cell adhesion molecule 1 (VCAM-1), and P-selectins and beta2 integrins, allowing for the firm attachment of leukocytes (Libby and Theroux, 2005). Endothelial activation/dysfunction is a first step in the establishment of the incipient atherosclerotic lesion. Under inflammatory conditions, endothelial cells will also secrete chemokines such as monocyte chemotactic protein-1 (MCP-1) which directs the migration of monocytes (Boring et al, 1998). After adhering to the endothelium, leukocytes will migrate into the intima of the blood vessel wall where they multiply. This migration and diapedesis is facilitated by the expression of matrix metalloproteinases (MMPs) such as MMP-9 by the adherent monocytes (Amorino and Hoover, 1998). Once within the intima, monocytes are induced by macrophage colony stimulating factor (M-CSF) to mature into macrophages (Clinton and Libby, 1992). Macrophages will also respond to M-CSF and express scavenger receptors, which engulf through endocytosis lypoproteins modified by inflammation (Packard and Libby, 2008). Accumulation of oxidized low-density lipoprotein (LDL) in the cytoplasma of macrophages will result in the formation of the so-called foam cells, characteristics of fatty streaks. At the same time, macrophages release an array of pro-inflammatory cytokines such as interleukin-1 $\beta$  (IL-1 $\beta$ ) and tumor necrosis factor-a (TNFa) amplifying the local inflammatory response and the events that lead to the earlystages of atherosclerosis. Apoptosis of foam cells will result in the release and accumulation of lipids in the intima (Packard and Libby, 2008).

The cytokines generated in the atherosclerotic lesions induce primarily a T helper 1 cell (Th1) response, which, in turn, amplifies the local inflammation (Robertson and Hansson, 2006). Atheromas grow in shoulder regions of the atherosclerotic lesions where macrophages and T cells infiltrate and deposition of fibrous tissues occurs (Packard and Libby, 2008). Smooth muscle cells (SMCs) migrate from the tunica media into the intima after degradation of the extracellular matrix by MMP-9 and other proteases (Mason et al, 1999). Once in the intima, SMCs proliferate under the influence of various growth factors such as transforming growth factor beta (TGF $\beta$ ) and platelet-derived growth factor (PDGF) and secrete collagen and other extracellular matrix components. This phase of the atheroma formation is characterized by its change from a lipid-rich plaque to a fibrous plaque, leading to stenosis (Packard and Libby, 2008).

The key histological features of the site of the atheroma rupture are the presence of large numbers of macrophages and a paucity of SMCs. There are several inflammatory mechanisms that might lead to fibrous cap rupture. The overall effect of inflammation on extracellular matrix components is the stimulation of their degradation and inhibition of their synthesis. IFN $\gamma$  can inhibit collagen production, and intersticial collagenases including MMP-1, MMP-8 and MMP-13 can be released by macrophages upon stimulation by pro-inflammatory signals such as IL-1 $\beta$  and CD40L. The

shoulder region and areas of foam cells contain MMP-9 and active MMP-9 can be recovered from human atherosclerotic plaques. These effects decrease the collagen content of the fibrous cap rendering it susceptible to rupture. Fibrous cap rupture exposes the atherosclerotic lesion to the liquid phase of blood, releasing a series of pro-coagulant factors, triggering thrombus formation and leading to obstruction of the blood flow (Haraszthy *et al*, 2000; Packard and Libby, 2008).

#### The oral infectious theory of atherogenesis

#### Detection of oral species in atherosclerotic lesions

To explore the possibility that oral bacteria could be associated with atherosclerosis, several investigators have examined if molecular signatures of oral species could be found in atheromatous lesions. These studies have utilized different techniques including polymerase chain reaction (PCR) of 16S rRNA genes (Haraszthy et al, 2000; Fiehn et al, 2005; Pucar et al, 2007; Aimetti et al, 2007; Mahendra et al, 2010), real-time PCR (Kozarov et al, 2006; Gaetti-Jardim et al, 2009), DNA-DNA hybridization (Elkaim et al, 2008), fluorescence in situ hybridization (FISH) (Cavrini et al, 2005) and culture of periodontal pathogens from atheromatous plaques (Kozarov et al, 2005). Although there is some logic in examining periodontal pathogens, as they present several characteristics that make them particularly attractive as causative agents of inflammatory processes, it is also quite conceivable that the properties required by an oral species to induce atherosclerosis differ considerably from those involved in periodontal tissue destruction. For instance, infective endocarditis is caused primarily by viridans group streptococci, staphylococci, and enterococci, none of which are putative periodontal pathogens (Baddour et al, 2005). In fact, an array of other oral species might be recovered from atheromas, including the etiological agent of dental caries, Streptococcus mutans (Nakano et al, 2006). In addition, bacteremia studies have revealed a large diversity in oral species entering the blood stream after different bacteremic stimuli, ranging from *streptococci* species to anaerobic gram-negative bacteria (Forner et al, 2006a).

#### Mechanisms of oral bacteria induced atherosclerosis

*Endothelial cell invasion.* The concept that infection can contribute to atherosclerosis is based primarily on studies examining pathogens that can cause persistent infection with the pathogen residing in cells for prolonged periods of times without proliferating. Agents such as cytomegalovirus (CMV), herpes simplex virus (HSV) and *Chlamydia pneumoniae* that have been implicated in human atherosclerosis by seroepidemiological studies are all intracellular pathogens (Epstein *et al*, 2009). Therefore, when examining potential pathological mechanisms linking periodontal diseases to atherosclerosis, it seems logical to explore host cell invasiveness by oral species. Results from a pioneer study by Deshpande *et al* (Deshpande *et al*, 1998) demonstrated that *P. gingivalis* could invade endothelial

cells in vitro and that the expression of fimbriae was necessary for this process. A study published soon after this initial report (Dorn et al, 1999) indicated that this phenomenon was not only species but also strain specific. A fimbriae expressing strain of P. gingivalis (381) had a higher capacity of invasion than the strain without fimbriae on its surface (W50), a strain of P. intermedia was invasive while another one was not, and Eikenella. corrodens presented a minimal ability to invade. Since these early reports, several other oral species including Porphyromonas endodontalis (Deshpande et al, 1998), S. mutans (Abranches et al, 2009), Streptococcus gordonii, Streptococcus sanguinis, Streptococcus mitis, and Streptococcus oralis (Stinson et al, 2003) have been shown to be able to invade endothelial cells

The importance of invasiveness was highlighted in a study where, using high-density oligonucleotide microarrays, the authors examined the gene expression profile of human aortic endothelial cells (HAEC) after infection with invasive and non-invasive strains of *P. gingivalis*. Infection of HAEC with invasive P. gingivalis strain 381 resulted in the upregulation of 68 genes. Genes coding for the pro-inflammatory cytokines, adhesion molecules, chemokines and cyclooxygenase-2 were among the most highly upregulated genes. Only 4 of these 68 genes were also upregulated in HAEC infected with the noninvasive P. gingivalis fimA mutant. Additional studies using immune techniques confirmed the expression of ICAM-1, VCAM-1, E-/P-selectins, IL-6, and IL-8 in HAEC infected with invasive P. gingivalis (Chou et al, 2005). Invasion of HAEC with P. gingivalis can also induce procoagulant effects including enhanced tissue factor expression and activity, and suppression of tissue factor pathway inhibitor. In addition, infection with invasive P. gingivalis 381 decreased levels and activity of tissue plasminogen activator and enhanced expression and activity of plasminogen activator inhibitor-1. Conversely, infection with a non-invasive strain of P. gingivalis failed to induce these procoagulant effects in HAEC (Roth et al, 2006).

It has also been suggested that certain species can help in the invasiveness of others. It was demonstrated that co-infection with F. nucleatum resulted in 2-20-fold increase in the invasion of endothelial cells by P. gingivalis strains, highlighting the importance of mixed infection in the modulation of the invasiveness of P. gingivalis (Saito et al, 2008). A recent study has suggested that after a prolonged intracellular phase P. gingivalis loses its ability to multiply, but upon co-incubation with fresh vascular host cells, it could be recovered. The data indicated that intercellular transmission could rescue latent intracellular P. gingivalis from a state of dormancy to a viable state (Li et al. 2008). In addition, the use of antibiotics that block the invasiveness of P. gingivalis has been shown in a murine model to decrease the systemic cytokine response after inoculation of P. gingivalis (Amar et al, 2009). These lines of evidence clearly indicate a potential role for endothelial cell invasion by oral species as a pathogenic mechanism in atherogenesis.

Endothelial activation. As described earlier, endothelial activation and the increased expression of adhesion molecules and chemokines are the first steps in the development of atherosclerotic lesions. A. actinomyce*temcomitans* infection of apolipoprotein E-deficient mice with a hyperlipidemic phenotype resulted in increased expression in the aorta of ICAM-1, E-selectin, P-selectin, MCP-1, chemokine (C-C motif) ligand 19 (CCL19), CCL21, and CCR7. Coculture of endothelial cells with P. gingivalis strains also resulted in the expression of ICAM-1, VCAM-1 and P- and E-selectins in a cell invasion-dependant mechanism mediated by fimbriae (Khlgatian et al, 2002; Takahashi et al, 2006). Other studies also reported that *P. gingivalis* can induce endothelial cells to express MCP-1. Dead *P. gingivalis* cells could still induce MCP-1 but at much lower levels. Chemical inhibition of endocytosis blocked MCP-1 upregulation, indicating the need for P. gingivalis internalization for MCP-1 stimulation (Kang and Kuramitsu, 2002).

By contrast, stimulation of endothelial cells with lipopolysacharide (LPS), outer membrane protein and heat shock protein 60 derived from P. gingivalis had only mild effects on the expression of ICAM-1 and VCAM-1 (Honda et al, 2005). Treatment of endothelial cells with P. gingivalis gingipains results in loss of adhesion properties and apoptotic cell death (Sheets et al, 2005). Paradoxically, invasion of endothelial cells by live P. gingivalis resulted in downregulation of IL-8 and MCP-1 through a lysine-specific cysteine proteinase (gingipain K)-mediated mechanism (Nassar et al, 2002). Another report by the same group demonstrated that invasive strains of *P. gingivalis* resulted in upregulation of IL-8 and MCP-1 (Takahashi et al, 2006). Soluble products from Eikenella corrodens can induce ICAM-1, VCAM-1, E-selectin and IL-8. Furthermore, these soluble products were capable of upregulating gene expression and protein production of vascular endothelial growth factor (VEGF) by human endothelial cells and activating endothelial cell proliferation (Yumoto et al, 2007).

Toll-like receptors mediated mechanisms. Exposure of endothelial cells to oral species might also result in increased expression and interactions with Toll-like receptors (TLRs). A. actinomycetemcomitans infection of apolipoprotein E-deficient mice resulted in increased expression in the aorta of TLR2 and TLR4 (Zhang et al 2010). Invasive strains of *P. gingivalis* can also stimulate the expression of TLR2 and TLR 4 on the surface of endothelial cells, while non-invasive, fimbriae deficient mutants, failed to do so (Yumoto et al, 2005). Extended exposure of LPS from P. gingivalis facilitates mononuclear cell adhesion to vascular endothelium via TLR2 mechanisms in vitro (Nakamura et al. 2008). Endothelial cells incubated with P. gingivalis LPS expressed ICAM-1 and VCAM-1 and antibodies against TLR2 and ICAM-1 blocked the enhanced mononuclear cell adhesion (Nakamura et al, 2008). Pro-inflammatory cytokine induction by P. gingivalis fimbriae can also be inhibited by monoclonal antibodies to TLR2, TLR4, CD14 and beta2 inregrins. Similarly, cytokine induction by

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*T. forsythia* protein A requires CD14 and TLR2. Antigen I/II from *S. mutans* can also stimulate cytokines partially through interactions with CD14 and TLR4 (Haijshengallis *et al.* 2002).

Autoimmunity. There is considerable evidence indicating that auto-immune mechanisms are involved in atherosclerosis. Surface expression of heat shock proteins (HSPs) as a consequence of inflammation, infection and oxidizing agents may be perceived as 'cryptic antigens' and recognized as 'foreign' by the immune system. In fact, in human atherosclerotic lesions, endothelial cells, macrophages and SMCs all express human heat shock protein 60 (hHSP60) and serum levels of anti-hHSP60 correlate with the presence and extent of CVDs (Epstein et al, 2009). Crossreaction of the immune response to bacterial GroEL with HSP60 has been suggested as a pathogenic mechanism linking oral species to atherosclerosis. As hHSP60 and the bacterial GroEL are highly conserved, a cross-reaction between antibodies to GroEL might lead to endothelial activation and damage (Ford et al, 2005b). This is an example of how 'molecular mimicry' can contribute to atherogenesis (Epstein et al, 2009). Levels of antibodies to GroEL and P. gingivalis could be reduced in plasma samples from subjects with atherosclerosis using absorption with human HSP60, suggesting cross-reactivity among these antigens (Ford et al, 2005b). In addition, GroEL-, HSP60-, and P. gingivalis-specific T-cell lines from atherosclerotic plaques have also demonstrated cross-reactivity (Ford et al, 2005a). Using immunohistology, the same group demonstrated the expression of hHSP60 in endothelial cells, smooth muscle cells and lymphocytes from carotid specimens, while GroEL was detected in intima cells (Ford et al, 2006).

Metalloproteinase and oxidative stress mediated mechanisms. Oral bacterial species may also induce the upregulation of MMPs by endothelial cells. Increased expression of MMP-9 in the aorta and a proatherogenic lipoprotein profile (smaller particles sizes in VLDL, LDL, and HDL lipoprotein fractions) in Apo-E deficient mice infected with A. actinomycetemcomitans has also been reported (Tuomainen et al, 2008). Oxidation of LDL is essential for its accumulation within macrophages and the formation of foam cells, can upregulate pro-atherogenic chemokines and adhesion molecules (Li et al, 2003) and induce the secretion of IL-6, TNF- $\alpha$ , and C-reactive protein (CRP) (Hulthe and Fagerberg, 2002). P. gingivalis can also stimulate LDL oxidation (Bengtsson et al, 2008) and rupture of atherosclerotic plaque through induction of MMPs (Ding et al. 1995). Co-incubation of a murine macrophage cell line with P. gingivalis, P. gingivalis outer membrane vesicles, and its LPS in the presence of LDL resulted in the formation of foam cells in a dose dependant manner (Qi et al, 2003). Furthermore, fibrous cap material isolated from atheromatous lesions obtained from human autopsy can be degraded by P. gingivalis in vitro (Kuramitsu et al, 2001).

#### Assessment of exposure

Periodontal inflammation results in an increased tendency for gingival bleeding, an ulcerated pocket epithelium and a larger number of oral bacteria accumulated within the gingival crevice. These conditions might increase the frequency and magnitude of bacteremias in subjects with gingivitis and/or periodontitis. The cumulative area of ulcerated epithelium for all periodontal lesions has been calculated to be  $8-20 \text{ cm}^2$  (Ĥujoel *et al*, 2001). There is also evidence that the incidence and magnitude of bacteremias after chewing, tooth brushing and subgingival scaling increased with the severity of periodontal inflammation (Forner et al, 2006a). Forner et al also demonstrated that periodontitis subjects presented a larger variety of species in the positive blood samples, reflecting the increased complexity of the subgingival biofilm in these subjects and, possibly, a higher pathogenic exposure (Forner et al, 2006a). Therefore, if chronic low-grade systemic exposures to oral microorganisms are associated with an increased risk of atherosclerosis, the severity of the periodontal condition of subjects should correlate with increased risk of CVD.

However, recent studies have questioned the correlation between severity of clinical parameters of periodontitis and increased susceptibility to bacteremias. Lockhart et al demonstrated that oral hygiene and gingivitis bleeding were significantly associated with the detection of bacteria in blood samples after tooth brushing, while parameters of periodontitis did not correlate with the incidence of bacteremia (Lockhart et al, 2009). These findings suggest that bleeding tendency was a better parameter to estimate the risk of bacteremia after tooth brushing than parameters of periodontal tissue destruction. A study examining the prevalence of bacteremia after flossing in 30 chronic periodontitis and 30 periodontally healthy subjects could not find a difference in the percentage of bacteremia positive subjects between groups (Crasta et al, 2009). Third molar extractions have also been associated with a high prevalence of bacteremia, independently of the oral health status of the extracted teeth (Tomas et al, 2007). However, a limitation of studies examining only the prevalence of bacteremia is that they do not take into account the intensity of the exposure. This is important because the intensity of the bacteremia is directly correlated with the risk of sequelae. For instance, bacteremia detected in clinical conditions involves levels from 10 to 100 colony forming units (CFU)  $ml^{-1}$ , while the level detected during bacterial endocarditis is approximately 200 CFU  $ml^{-1}$  (Lucas and Roberts, 2000). Therefore, if the intensity and complexity of bacteremias are relevant for the risk of atherogenesis, studies that do not quantify these parameters should be interpreted with caution.

The appropriateness of clinical parameters of periodontal diseases as a measure of exposure to oral infectious agents leading to CVD has also been challenged by others. Beck and Offenbacker (Beck and Offenbacher, 2002) reported that clinical attachment loss had a weaker association with systemic biomarkers of inflammation than probing depth or bleeding on probing. In a subsequent study, the same group examined a subset of participants in the Atherosclerosis Risk in Communities (ARIC) Study of 5,002 individuals. They could not find a correlation between the periodontal status of study subjects and the prevalence of coronary heart disease (Beck *et al*, 2005a). These lines of evidence have led researchers to question what would be the appropriate measure of exposure to investigate the link between periodontal infections and CVD.

There are epidemiological studies that have used direct assessments of colonization by periodontal bacteria demonstrating associations between levels of colonization by specific oral bacterial species and CVD outcomes (Desvarieux et al, 2005; Renvert et al, 2006; Spahr et al, 2006; Nonnenmacher et al, 2007). Studies examining the correlations between oral microorganisms and CVD are particularly relevant if the link between periodontal diseases and CVD is to be supported on the grounds of an infectious mechanisms. In that context, quantifying the exposure to oral species might be more significant than the assessment of clinical manifestations of oral infections, particularly when certain clinical parameters might be poor surrogates for infection exposure. However, direct measurement of levels of oral microorganisms in the mouth might still not be the best measure of exposure. The current understanding is that oral bacteria could lead to atherogenesis once they have entered the blood stream through bacteremias. Direct assessment of the presence of oral species in the blood is also counter productive because bacteremias are short lived and peak between 30 s and 2 min and decreases considerably after 10 min (Forner et al, 2006a; Tomas et al, 2007).

Serum antibodies specific to oral bacteria have also been used to assess exposure to an oral infectious challenge (Pussinen et al, 2004, 2005, 2007; Beck et al, 2005a; Johansson et al, 2005). Although results were not always consistent, a meta-analysis of seroepidemiological studies demonstrated that elevated systemic antibody responses to oral pathogens were strongly associated with an increased risk of coronary heart disease (Mustapha et al, 2007). For instance, Pussinen et al demonstrated in a sample of 893 Finnish subjects free of CVD at baseline and followed for 15 years that the presence of serum antibodies to P. gingivalis increased the risk of stroke. High serum antibody levels to P. gingivalis predicted stroke with ORs of 1.6 and 2.3 for males and females (Pussinen et al, 2007). In their analysis of 5002 subjects enrolled in the ARIC study, Beck et al could not establish a correlation between clinical parameters of periodontitis and coronary heart disease (CHD), but demonstrated an association between systemic antibody responses to several oral species and the prevalence of CHD. They concluded that 'the quality and quantity of the host response to oral bacteria may be an exposure more relevant to systemic atherothrombotic coronary events than clinical measures' (Beck et al, 2005b).

The use of systemic antibody responses to oral microorganisms can be an attractive measure of infec-

tious exposure for several reasons: 1) the presence of systemic antibodies to a given bacterium indicates systemic exposure to the microorganism or its components (Beck et al. 2005a): 2) specific antibodies to a microorganism indicates that the host recognized it as foreign and the level of response can be interpreted as a measure of its pathogenic potential (Haffajee and Socransky, 1994); 3) a vigorous antibody reaction might reflect the level of immune responsiveness of the host, which might also correlate with the increased risk of CVD (Mattila et al, 2005); 4) fluctuations in specific antibody levels correlate to infection and parallel changes in the burden of a specific microorganism in the subgingival plaque (Ebersole et al, 1992); 5) systemic antibody levels decrease after periodontal therapy, in response to a decrease in the bacterial challenge but remain elevated compared with periodontally healthy subjects (Aukhil et al, 1988; Ebersole et al, 1992; Papapanou et al, 2004); 6) IgG antibodies indicate cumulative bacterial exposure while IgA response indicates more recent exposure (Pussinen et al, 2007); 7) systemic antibodies can be used to examine past exposure in edentulous subjects (Pussinen et al, 2003; Papapanou et al, 2004); 8) despite the requirement of an invasive procedure to be obtained, serum is very convenient sample that can be aliquoted and stored frozen for several years and can be used to examine immune reactivity to a vast array of oral and non-oral microorganisms; 9) because immunoglobulins are present in large amounts, minute quantities of serum can be used for their measurement; and 10) high-throughput techniques exist for the quantification of antibody response to a multitude of microorganisms at the same time (Sakellari et al, 1997). In fact, most of the evidence in the early studies examining the infectious hypothesis of atherogenesis used seroepidemiology to determine a correlation between exposure to cytomegalovirus, herpes simplex virus, Chlamidia pneumoniae and Helicobacter pylori and CVD (Epstein et al, 2009).

#### Infectious/pathogen burden

The large clinical trials testing if macrolid antibiotic therapy targeting C. pneumoniae would lead to a decrease in cardiovascular events all resulted in negative outcomes (Andraws et al, 2005). These studies clearly indicated that the routine use of antibiotic therapy for this pathogen was not indicated in secondary prevention of coronary events. However, these studies were often erroneously interpreted as negating the participation of infectious agents in atherosclerosis. The most important limitation of these antibiotic trials is the concept that if microorganisms contribute to atherogenesis and its complications, many such microorganisms might contribute to these biological processes (Epstein et al, 2009). Therefore, the infection-associated risk of atherosclerosis should correlate best with the aggregate pathogen load, what was termed 'pathogen burden' (Zhu et al, 2000). Prospective studies performed in coronary artery disease subjects with several years of follow-up demonstrated that the number of viral and bacterial pathogens with which an individual had been infected

predicted the incidence of acute myocardial infarction and death (Rupprecht *et al*, 2001; Zhu *et al*, 2001).

These concepts have important implications to the interpretation of the findings of studies examining the association between periodontal infections and atherosclerosis. The data discussed above outlying putative mechanisms linking oral species to atherogenesis indicated that several oral species are capable of mechanisms that might trigger the formation and progression of atherosclerotic lesions. However, most mechanistic studies have focused primarily on a limited number of periodontal pathogens. As the oral cavity can be colonized by over 500 different species, the task of examining the potential involvement of all such species is overwhelming. In addition, studies examining the exposure to oral pathogens should also take into account the potential contribution of non-oral bacterial pathogens and viruses. Several studies recognize the relevance of accounting for the exposure to non-oral pathogens and have examined the presence of molecular signatures of C. pneumoniae and human cytomegalovirus (HCMV) in atherosclerotic plaques, in addition to oral species (Haraszthy et al, 2000; Ford et al, 2005b, 2006; Pucar et al, 2007). The results from these studies varied considerably and were based on limited numbers of samples, however, a few general trends could be observed. P. gingivalis was the most commonly found periodontal pathogen. The prevalence of several periodontal pathogens and, particularly P. gingivalis, was higher than C. pneumoniae (Haraszthy et al, 2000; Ford et al, 2005b, 2006; Pucar et al, 2007), while the prevalence of HCMV was higher than any periodontal pathogen tested (Haraszthy et al, 2000; Pucar et al, 2007). Universal probes for bacteria revealed the presence of bacterial DNA that could not be accounted for by the specific probes, suggesting the presence of unidentified bacteria (Haraszthy et al, 2000; Pucar et al, 2007). These preliminary findings indicate that if an association between periodontal diseases and CVD is to be explained by the exposure to oral microorganisms, a measure of the total pathogenic burden must be determined. Hitherto, there is no consensus regarding which microorganisms should be examined and a method to assess the infectious burden (Elkind, 2010). Considering the complexity of the oral microbiota and the contribution of non-oral bacterial species and viruses to the microbial burden, obtaining a measure of the cumulative microbial burden will not be an easy task.

#### The oral inflammatory theory of atherogenesis

#### Periodontal diseases and systemic biomarkers of inflammation

The role for inflammation in the pathogenesis of atherosclerosis was established in part by examining associations between systemic biomarkers of inflammation and measurements of CVD and its consequences. Originally, it was thought that such systemic biomarkers would reflect a heightened inflammatory state at the atheromatous lesions. However, in population studies, systemic elevations of CRP and cytokine levels are not

explained by the magnitude of atherosclerotic lesions detected per subject (Libby, 2002). Systemic elevations in inflammatory parameters are more associated with age, smoking and adiposity than with measures of plaque thickness. Adiposity, in particular, has been shown to explain up to 30% of the systemic inflammatory burden in population studies (Libby, 2002; Pearson et al, 2003). This implies that elevations in systemic cytokines can be derived not necessarily from the site of the disease (the atherosclerotic lesion), but also from other tissues such as the periodontium. These studies led to the exploration of the correlation between oral parameters of inflammation and the levels of systemic biomarkers. A recent meta-analysis of cross-sectional studies examining the association between periodontal diseases and CRP concluded that 'There is strong evidence from cross-sectional studies that plasma CRP in periodontitis is elevated compared with controls' (Paraskevas et al, 2008). The authors included only reports using high-sensitivity CRP measurements. The mean levels of CRP reported in 10 studies reviewed in their article were  $3.41 \text{ mg l}^{-1}$  for subjects with periodontal diseases and  $1.72 \text{ mg l}^{-1}$  for periodontally healthy individuals (Paraskevas et al, 2008). These data are quite relevant as an elevated risk for atherosclerosis is indicated by CRP levels  $\geq 3 \text{ mg l}^{-1}$ . Since then, recent investigations have added to the evidence that periodontal diseases are associated with elevated systemic levels of CRP (Buhlin et al, 2009; Gani et al, 2009; Nakajima et al, 2010). In a longitudinal study involving 11 162 Japanese men, the authors reported that for men with normal levels of CRP at baseline, the presence of periodontal disease correlated with increases in CRP levels 1 year later (OR: 1.34; 95% CI: 1.12-1.67), suggesting that periodontal diseases preceded the systemic increase in CRP (Yoshii et al, 2009). In addition to CRP, other studies have reported elevated systemic levels of IL-6, a major inducer of the acute phase reaction and CRP, in periodontitis subjects (Loos, 2005; Nakajima et al, 2010). Higher levels of fibrinogen (Kweider et al, 1993) and IL-18 (Buhlin et al, 2009) have also been reported in periodontal disease subjects. Therefore, periodontal diseases seem to be associated with a low-grade systemic inflammation, with levels of serum pro-inflammatory biomarkers being elevated in periodontitis subjects.

#### Sources of systemic inflammatory mediators in periodontal diseases

The increase in systemic biomarkers of inflammation associated with periodontal diseases has been interpreted as a mechanistic link between these infections and CVD. However, details of the biological processes that lead to the systemic inflammation associated with periodontal diseases are poorly understood. As discussed above, it is possible that these elevations in systemic markers of inflammation occur in response to systemic exposure to oral bacteria during bacteremias. Alternatively, it has been proposed that cytokines secreted within the periodontal tissues would find their way into the blood stream and cause a low-grade

systemic inflammation. A similar mechanism is at play in RA, where the synovial tissues are the primary site of inflammation and act as a source of pro-inflammatory cvtokines such as TNF- $\alpha$ . IL-1 $\beta$ , and IL-6 which contributes to the 'high-grade' systemic inflammation. In support of mechanism, in RA the magnitude of the systemic inflammation is associated with an increase in several risk factors for atherosclerosis (Sattar et al, 2003). Furthermore, CVD and premature mortality in RA subjects correlates with the number of inflamed joints, indicating that these affected sites might release mediators of atherogenesis (Pincus et al, 1984). Several publications have demonstrated local elevations of proinflammatory biomarkers within periodontal tissues affected by gingivitis and periodontitis (Gorska et al. 2003; Hou et al, 2003; Lester et al, 2009). These include mediators that have been implicated in systemic inflammation such as IL-1 $\beta$ , IL-6, and TNF- $\alpha$ . However, different from RA, the contribution of these local mediators to the systemic inflammation lacks supporting evidence. In fact, periodontal inflammation has not been shown to be associated with systemic elevations of IL-1 $\beta$ (Mengel *et al*, 2002) or TNF- $\alpha$  (Meyle, 1993), although both biomarkers are abundantly present in tissue samples from periodontal disease sites both at the mRNA and protein levels (Roberts et al, 1997a,b; Hou et al, 2003), and in proximal samples of gingival crevicular fluid (Gamonal et al, 2000, 2003; Teles et al, 2010a,b). Studies examining the correlation between local (in GCF samples) and systemic (serum or plasma) levels of biomarkers in periodontitis subjects have failed to demonstrate a strong association (Gorska et al, 2003). Several hypotheses could explain this observation. Pro-inflammatory cytokines released in the periodontal sites might be partially consumed locally within the periodontal tissues and the excess be preferentially 'drained' by the GCF, minimizing the systemic exposure. In any case, it seems that the periodontal lesion is well circumscribed and partially isolated from the rest of the body.

An alternative hypothesis posits that subjects susceptible to periodontal diseases would present a propensity to low-grade systemic inflammations. This inflammatory phenotype would expose them to a higher risk for inflammatory diseases such as periodontal diseases and atherosclerosis, explaining their co-expression. Rheumatoid arthritis exemplifies how a pro-inflammatory phenotype might predispose subjects to CVD and possibly other inflammatory conditions. In these individuals atherosclerosis can be considered as an 'extraarticular manifestation of the syndrome' and the term 'rheumatoid vasculopathy' has been proposed (Fietta and Delsante, 2009). Similarly, a few reports have described an association between the RA and periodontal diseases (Kasser et al, 1997; Gleissner et al, 1998; Mercado et al, 2001), implying common inflammatory mechanisms. However, as discussed above, there is evidence that periodontal disease might precede systemic CRP elevations, suggesting that the higher levels of CRP in periodontal diseases would occur as a consequence of the disease process (Yoshii et al, 2009).

#### Oral Diseases

## Effects of periodontal therapy on systemic biomarkers of inflammation

The immediate effect of tooth debridement seems to be an increase in plasma levels of TNFa. CRP. and IL-6. suggesting an acute response to the bacteremia resultant from this procedure (D'Aiuto et al, 2004, 2005; Ide et al, 2004; Tonetti et al, 2007). However, not all studies on the impact of bacteremias on systemic levels of cytokines have reported elevations as a consequence of bacteremic events. Forner et al measured the levels of six cytokines in serum samples in 20 severe periodontitis subjects 8 h after scaling (Forner et al, 2006b). Bacteremia was detected in 15 such subjects but only IL-6 was statistically significantly elevated, IL-8 was statistically significantly reduced, while the levels of IL-1 $\beta$ , TNF $\alpha$ , IL-10, and IL-12p70 were not affected. Furthermore, changes in serum levels of biomarkers were not associated with magnitude, duration or composition of the bacteremias. A recently published systematic review on the effects of different periodontal therapies on the systemic levels of CRP concluded that there was modest evidence that periodontal treatment was associated with reductions in CRP levels (Paraskevas et al, 2008). Interestingly, of the studies reviewed, baseline CRP levels were all below 2.0 mg  $l^{-1}$ , comparing to levels above 3.0 mg  $l^{-1}$  for periodontitis subjects in cross-sectional studies as previously discussed. Furthermore, reductions were rather modest: 0.4-0.6 mg l<sup>-1</sup> and most studies were of shortterm follow-up (maximum of 6 months). Since this review was published, a few other studies have addressed the effects of periodontal treatment on systemic biomarkers of inflammation and reported inconsistent findings. Lopez et al reported statistically significant increases in CRP in a group of 73 patients 6 weeks after periodontal therapy with adjunctive antibiotics from  $3.6 \pm 3.7 \text{ mg l}^{-1}$  to  $5.4 \pm 5.7 \text{ mg l}^{-1}$ (Lopez et al, 2009). By contrast, other studies reported statistically significant reductions in plasma levels of CRP, IL-6, and fibrinogen 1–3 months after periodontal therapy (Blum et al, 2007; Marcaccini et al, 2009; Vidal et al, 2009). Although reported decreases in CRP associated with periodontal treatment were statistically significant, they were rather modest and would not result in changes in the risk stratification for CVD. Ushida et al found no changes in serum CRP after three different types of periodontal therapy (Ushida et al, 2008).

Interestingly, a recent study examining the effects of periodontal therapy on a panel of 19 systemic biomarkers demonstrated that treatment resulted in significant mean reductions in levels of PAI-1, sE-selectin, sVCAM-1, MMP-9 and myeloperoxidase (Behle *et al*, 2009). However, only reductions in sE-selectin and sICAM and serum amyloid P were maintained 4 weeks after the end of therapy. Importantly, they found a great variability in the systemic response to periodontal treatment among studied subjects with <sup>1</sup>/<sub>3</sub> showing marked reductions and <sup>1</sup>/<sub>4</sub> demonstrating significant increases in systemic biomarkers, with the remainder unchanged. The authors also reported a poor correlation between clinical, microbiological and serological

parameters of periodontal disease and changes in the inflammation biomarkers. This study illustrates how the effects of periodontal therapy on systemic biomarkers of inflammation may vary greatly among study subjects. This variability might partially explain the inconsistency in the reports discussed above.

Full-mouth tooth extraction can be interpreted as a definitive (although highly undesirable) form of treatment of periodontal diseases. Therefore, examining the effects of full-mouth extraction on systemic biomarkers associated with CVD could be quite instructive. So far, only a few publications have reported on this type of intervention to address the effects of elimination of periodontal disease on atherogenesis risk. The extraction of all remaining teeth in 10 subjects with 'end-stage periodontitis' resulted in statistically significant decreases in the mean plasma levels of CRP from 3.5 to 1.6 mg  $l^{-1}$ , 12 months after the extractions. Interestingly, this reduction occurred over several months, with most subjects only reaching values below  $1 \text{ mg l}^{-1}$ , 9 months after the intervention. Furthermore, in two subjects with very high baseline values (>6 mg  $l^{-1}$ ), the extractions resulted in either no change or only a reduction to levels still above 4 mg  $l^{-1}$ , demonstrating a certain degree of variability in the response to treatment (Rahman et al, 2005). Another study involving 67 subjects with advanced periodontal disease, full-mouth extraction resulted in a significant decrease in plasma levels of CRP, PAI-1 and fibrinogen, which lasted up to 12 weeks after the procedure (Taylor et al, 2006). However, the impact of the extraction on the mean values of CRP was modest, with a mean reduction of  $0.7 \text{ mg } l^{-1}$ .

To put the magnitude of these effects in perspective, the JUPITER study, which resulted in a statistically significant reduction in the occurrence of a first major cardiovascular event (P < 0.00001) and death from any cause (P < 0.02), reported median levels of CRP of approximately 2.0 mg  $l^{-1}$  for the statin group compared to 3.5 mg  $l^{-1}$  for the placebo group during 48 months of follow-up (Ridker et al, 2008). These data suggest that the clinical effects associated with the use of statins on cardiovascular events required a long-term consistent reduction in CRP to levels below the high risk threshold of  $3.0 \text{ mg } l^{-1}$ . So far, no intervention study on periodontal disease subjects has been able to report this magnitude of change in systemic CRP. Periodontal therapy seems to reduce the mean levels of systemic CRP by approximately 0.5 mg  $l^{-1}$ , 1/3 of the reduction obtained with the use of rosuvastatin. Furthermore, there is little to no information on the long-term effects of periodontal therapy on the plasma levels of CRP.

Hitherto, only one multi-center study has examined the effects of therapy on the secondary prevention of cardiac events, the Periodontitis and Vascular Events (PAVE) study (Beck *et al*, 2008; Offenbacher *et al*, 2009). The study demonstrated no significant effect of periodontal therapy on serum CRP levels 6 months after treatment when data were analyzed using intent-to-treat analyses. However, the study lacked a proper control group as subjects were assigned either to 'protocol therapy' provided by the study team or to 'community care'. When subjects receiving any form of treatment were compared with subjects with no treatment in a secondary analysis, periodontal therapy resulted in a significant reduction in the proportion of subjects with CRP levels  $> 3 \text{ mg l}^{-1}$ . Nevertheless, this effect was only present when non-obese people were included in the analyses. The study was also compromised by the inclusion of subjects with mild periodontitis with mean ( $\pm$  s.d.) PD values of 2.72 mm  $\pm$  0.05 and 2.69 mm  $\pm$  0.06 for the community and protocol treatment groups, respectively.

Therefore, the intervention studies testing the effects of periodontal treatment on surrogate biomarkers of cardiovascular risk can only be considered inconclusive at this point. Even when a beneficial effect in reducing systemic CRP was reported, the magnitude of the effect might not have been sufficient to alter the cardiovascular risk of the study subjects. This observation coupled with evidence of a high degree of inter-individual variability in the systemic effects of periodontal therapy, and a lack of major changes in systemic inflammation biomarkers in response to a 'definitive therapy' such as full-mouth extraction, indicate that the impact of periodontal treatment on cardiovascular risk can only be expected to be marginal. These data are in accord with evidence that there are more important sources of systemic inflammation burden, such as adiposity and smoking and reinforce the notion that periodontal diseases confer a moderate risk to atherosclerosis and its consequences. In addition, the reduction in the risk can only be sustained for as long as reduction in systemic inflammation lasts.

#### Conclusion and future direction

Several periodontal pathogens and other oral species have been indisputably identified in bacteremias and in atherosclerotic lesions and several mechanisms that implicate them in the inflammatory events that lead to all stages of atherosclerosis have been proposed. Evidence also supports the notion that periodontal diseases are accompanied by low-grade systemic inflammation, as documented by an increase in plasma CRP levels. However, additional evidence linking oral bacterial exposure to this state of heightened systemic inflammation is still needed. The study of the association between the two phenomena, i.e. entry of oral bacteria into the blood stream and systemic inflammation, is hampered by the complexity of the oral microbiota and difficulties in determining a proper measure of exposure to the cumulative microbial burden. The understanding of the biological mechanisms underlying this potential association between periodontal diseases and atherosclerosis is also compromised by the existence of a series of confounding factors that might contribute to systemic inflammation and possibly affect both diseases such as age, adiposity, smoking and insulin resistance. Intervention studies have contributed little to clarify the mechanisms involved in a possible increased risk for CVDs conferred by periodontal infections. Future studies might benefit from the lessons learned from

both the PAVE and the JUPITER studies. Recruitment of a large enough sample size to compensate for covariates; inclusion of appropriate treatment groups to test the impact of the periodontal infection to the outcome measures (e.g. adjunct systemic antibiotic vs. mechanical therapy); longitudinal monitoring to examine the duration of the therapeutic effect on systemic biomarkers; and selection of subjects that present concomitantly moderate to severe periodontal disease and high systemic levels (> 3.0 mg l<sup>-1</sup>) of CRP might be required to clarify if periodontal therapy has a real impact on cardiovascular risk. Evidence that control of periodontal infections leads to a decrease risk for cardiovascular events, would greatly substantiate a link between both conditions.

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#### References

- Abranches J, Zeng L, Belanger M *et al* (2009). Invasion of human coronary artery endothelial cells by Streptococcus mutans OMZ175. *Oral Microbiol Immunol* **24**: 141–145.
- Aimetti M, Romano F, Nessi F (2007). Microbiologic analysis of periodontal pockets and carotid atheromatous plaques in advanced chronic periodontitis patients. J Periodontol 78: 1718–1723.
- Ajwani S, Mattila KJ, Tilvis RS, Ainamo A (2003). Periodontal disease and mortality in an aged population. *Spec Care Dentist* 23: 125–130.
- Amar S, Wu SC, Madan M (2009). Is Porphyromonas gingivalis cell invasion required for atherogenesis? Pharmacotherapeutic implications. J Immunol 182: 1584–1592.
- Amorino GP, Hoover RL (1998). Interactions of monocytic cells with human endothelial cells stimulate monocytic metalloproteinase production. *Am J Pathol* **152**: 199–207.
- Andraws R, Berger JS, Brown DL (2005). Effects of antibiotic therapy on outcomes of patients with coronary artery disease: a meta-analysis of randomized controlled trials. *JAMA* 293: 2641–2647.
- Aukhil I, Lopatin DE, Syed SA, Morrison EC, Kowalski CJ (1988). The effects of periodontal therapy on serum antibody (IgG) levels to plaque microorganisms. J Clin Periodontol 15: 544–550.
- Baddour LM, Wilson WR, Bayer AS *et al* (2005). Infective endocarditis: diagnosis, antimicrobial therapy, and management of complications: a statement for healthcare professionals from the Committee on Rheumatic Fever, Endocarditis, and Kawasaki Disease, Council on Cardiovascular Disease in the Young, and the Councils on Clinical Cardiology, Stroke, and Cardiovascular Surgery and Anesthesia, American Heart Association: endorsed by the Infectious Diseases Society of America. *Circulation* **111**: e394–e434.
- Beck JD, Offenbacher S (2002). Relationships among clinical measures of periodontal disease and their associations with systemic markers. *Ann Periodontol* **7:** 79–89.
- Beck J, Garcia R, Heiss G, Vokonas PS, Offenbacher S (1996). Periodontal disease and cardiovascular disease. *J Periodontol* **67:** 1123–1137.

- Beck JD, Eke P, Heiss G *et al* (2005a). Periodontal disease and coronary heart disease: a reappraisal of the exposure. *Circulation* **112**: 19–24.
- Beck JD, Eke P, Lin D *et al* (2005b). Associations between IgG antibody to oral organisms and carotid intima-medial thickness in community-dwelling adults. *Atherosclerosis* **183**: 342–348.
- Beck JD, Couper DJ, Falkner KL *et al* (2008). The Periodontitis and Vascular Events (PAVE) pilot study: adverse events. *J Periodontol* **79:** 90–96.
- Behle JH, Sedaghatfar MH, Demmer RT *et al* (2009). Heterogeneity of systemic inflammatory responses to periodontal therapy. *J Clin Periodontol* **36**: 287–294.
- Bengtsson T, Karlsson H, Gunnarsson P *et al* (2008). The periodontal pathogen Porphyromonas gingivalis cleaves apoB-100 and increases the expression of apoM in LDL in whole blood leading to cell proliferation. *J Intern Med* **263**: 558–571.
- Blum A, Front E, Peleg A (2007). Periodontal care may improve systemic inflammation. *Clin Invest Med* **30:** E114–E117.
- Boring L, Gosling J, Cleary M, Charo IF (1998). Decreased lesion formation in CCR2-/- mice reveals a role for chemokines in the initiation of atherosclerosis. *Nature* **394**: 894–897.
- Buhlin K, Hultin M, Norderyd O et al (2009). Risk factors for atherosclerosis in cases with severe periodontitis. J Clin Periodontol 36: 541–549.
- Cavrini F, Sambri V, Moter A *et al* (2005). Molecular detection of Treponema denticola and Porphyromonas gingivalis in carotid and aortic atheromatous plaques by FISH: report of two cases. *J Med Microbiol* **54**: 93–96.
- Chou HH, Yumoto H, Davey M *et al* (2005). Porphyromonas gingivalis fimbria-dependent activation of inflammatory genes in human aortic endothelial cells. *Infect Immun* **73**: 5367–5378.
- Clinton SK, Libby P (1992). Cytokines and growth factors in atherogenesis. *Arch Pathol Lab Med* **116**: 1292–1300.
- Crasta K, Daly CG, Mitchell D, Curtis B, Stewart D, Heitz-Mayfield LJ (2009). Bacteraemia due to dental flossing. *J Clin Periodontol* **36**: 323–332.
- D'Aiuto F, Ready D, Tonetti MS (2004). Periodontal disease and C-reactive protein-associated cardiovascular risk. *J Periodontal Res* **39:** 236–241.
- D'Aiuto F, Casas JP, Shah T, Humphries SE, Hingorani AD, Tonetti MS (2005). C-reactive protein (+1444C>T) polymorphism influences CRP response following a moderate inflammatory stimulus. *Atherosclerosis* **179**: 413–417.
- Deshpande RG, Khan M, Genco CA (1998). Invasion strategies of the oral pathogen porphyromonas gingivalis: implications for cardiovascular disease. *Invasion Metastasis* **18**: 57–69.
- DeStefano F, Anda RF, Kahn HS, Williamson DF, Russell CM (1993). Dental disease and risk of coronary heart disease and mortality. *BMJ* **306**: 688–691.
- Desvarieux M, Demmer RT, Rundek T *et al* (2005). Periodontal microbiota and carotid intima-media thickness: the Oral Infections and Vascular Disease Epidemiology Study (INVEST). *Circulation* **111**: 576–582.
- Ding Y, Uitto VJ, Firth J *et al* (1995). Modulation of host matrix metalloproteinases by bacterial virulence factors relevant in human periodontal diseases. *Oral Dis* 1: 279–286.
- Dorn BR, Dunn WA Jr, Progulske-Fox A (1999). Invasion of human coronary artery cells by periodontal pathogens. *Infect Immun* 67: 5792–5798.

- Ebersole JL, Cappelli D, Steffen MJ (1992). Characteristics and utilization of antibody measurements in clinical studies of periodontal disease. *J Periodontol* **63**: 1110–1116.
- Elkaim R, Dahan M, Kocgozlu L *et al* (2008). Prevalence of periodontal pathogens in subgingival lesions, atherosclerotic plaques and healthy blood vessels: a preliminary study. *J Periodontal Res* **43**: 224–231.
- Elkind MS (2010). Infectious burden: a new risk factor and treatment target for atherosclerosis. *Infect Disord Drug Targets* **10**: 84–90.
- Epstein SE, Zhu J, Najafi AH, Burnett MS (2009). Insights into the role of infection in atherogenesis and in plaque rupture. *Circulation* **119**: 3133–3141.
- Fiehn NE, Larsen T, Christiansen N, Holmstrup P, Schroeder TV (2005). Identification of periodontal pathogens in atherosclerotic vessels. J Periodontol 76: 731–736.
- Fietta P, Delsante G (2009). Atherogenesis in rheumatoid arthritis: the "rheumatoid vasculopathy"? *Acta Biomed* 80: 177–186.
- Ford P, Gemmell E, Walker P, West M, Cullinan M, Seymour G (2005a). Characterization of heat shock protein-specific T cells in atherosclerosis. *Clin Diagn Lab Immunol* **12:** 259–267.
- Ford PJ, Gemmell E, Hamlet SM *et al* (2005b). Crossreactivity of GroEL antibodies with human heat shock protein 60 and quantification of pathogens in atherosclerosis. *Oral Microbiol Immunol* **20**: 296–302.
- Ford PJ, Gemmell E, Chan A *et al* (2006). Inflammation, heat shock proteins and periodontal pathogens in atherosclerosis: an immunohistologic study. *Oral Microbiol Immunol* **21**: 206–211.
- Forner L, Larsen T, Kilian M, Holmstrup P (2006a). Incidence of bacteremia after chewing, tooth brushing and scaling in individuals with periodontal inflammation. J Clin Periodontol 33: 401–407.
- Forner L, Nielsen CH, Bendtzen K, Larsen T, Holmstrup P (2006b). Increased plasma levels of IL-6 in bacteremic periodontis patients after scaling. *J Clin Periodontol* **33**: 724–729.
- Gaetti-Jardim E Jr, Marcelino SL, Feitosa AC, Romito GA, Avila-Campos MJ (2009). Quantitative detection of periodontopathic bacteria in atherosclerotic plaques from coronary arteries. J Med Microbiol 58: 1568–1575.
- Gamonal J, Acevedo A, Bascones A, Jorge O, Silva A (2000). Levels of interleukin-1 beta, -8, and -10 and RANTES in gingival crevicular fluid and cell populations in adult periodontitis patients and the effect of periodontal treatment. *J Periodontol* **71:** 1535–1545.
- Gamonal J, Sanz M, O'Connor A *et al* (2003). Delayed neutrophil apoptosis in chronic periodontitis patients. *J Clin Periodontol* **30**: 616–623.
- Gani DK, Lakshmi D, Krishnan R, Emmadi P (2009). Evaluation of C-reactive protein and interleukin-6 in the peripheral blood of patients with chronic periodontitis. *J Indian Soc Periodontol* **13:** 69–74.
- Gleissner C, Willershausen B, Kaesser U, Bolten WW (1998). The role of risk factors for periodontal disease in patients with rheumatoid arthritis. *Eur J Med Res* **3**: 387– 392.
- Gorska R, Gregorek H, Kowalski J, Laskus-Perendyk A, Syczewska M, Madalinski K (2003). Relationship between clinical parameters and cytokine profiles in inflamed gingival tissue and serum samples from patients with chronic periodontitis. *J Clin Periodontol* **30**: 1046–1052.
- Haffajee AD, Socransky SS (1994). Microbial etiological agents of destructive periodontal diseases. *Periodontol* **2000**: 78–111.

- Hajishengallis G, Sharma A, Russell MW, Genco RJ (2002). Interactions of oral pathogens with toll-like receptors: possible role in atherosclerosis. *Ann Periodontol* **7:** 72–78.
- Haraszthy VI, Zambon JJ, Trevisan M, Zeid M, Genco RJ (2000). Identification of periodontal pathogens in atheromatous plaques. J Periodontol 71: 1554–1560.
- Honda T, Oda T, Yoshie H, Yamazaki K (2005). Effects of Porphyromonas gingivalis antigens and proinflammatory cytokines on human coronary artery endothelial cells. *Oral Microbiol Immunol* 20: 82–88.
- Hou LT, Liu CM, Liu BY, Lin SJ, Liao CS, Rossomando EF (2003). Interleukin-1beta, clinical parameters and matched cellular-histopathologic changes of biopsied gingival tissue from periodontitis patients. *J Periodontal Res* **38**: 247–254.
- Howell TH, Ridker PM, Ajani UA, Hennekens CH, Christen WG (2001). Periodontal disease and risk of subsequent cardiovascular disease in U.S. male physicians. J Am Coll Cardiol 37: 445–450.
- Hujoel PP, Drangsholt M, Spiekerman C, DeRouen TA (2000). Periodontal disease and coronary heart disease risk. *JAMA* **284**: 1406–1410.
- Hujoel PP, White BA, Garcia RI, Listgarten MA (2001). The dentogingival epithelial surface area revisited. *J Periodontal Res* **36**: 48–55.
- Hulthe J, Fagerberg B (2002). Circulating oxidized LDL is associated with increased levels of cell-adhesion molecules in clinically healthy 58-year old men (AIR study). *Med Sci Monit* 8: CR148–CR152.
- Humphrey LL, Fu R, Buckley DI, Freeman M, Helfand M (2008). Periodontal disease and coronary heart disease incidence: a systematic review and meta-analysis. J Gen Intern Med 23: 2079–2086.
- Hung HC, Joshipura KJ, Colditz G *et al* (2004). The association between tooth loss and coronary heart disease in men and women. *J Public Health Dent* **64**: 209–215.
- Ide M, Jagdev D, Coward PY, Crook M, Barclay GR, Wilson RF (2004). The short-term effects of treatment of chronic periodontitis on circulating levels of endotoxin, C-reactive protein, tumor necrosis factor-alpha, and interleukin-6. *J Periodontol* **75:** 420–428.
- Janket SJ, Baird AE, Chuang SK, Jones JA (2003). Metaanalysis of periodontal disease and risk of coronary heart disease and stroke. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 95: 559–569.
- Johansson A, Buhlin K, Koski R, Gustafsson A (2005). The immunoreactivity of systemic antibodies to Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in adult periodontitis. *Eur J Oral Sci* **113**: 197–202.
- Joshipura KJ, Rimm EB, Douglass CW, Trichopoulos D, Ascherio A, Willett WC (1996). Poor oral health and coronary heart disease. *J Dent Res* **75**: 1631–1636.
- Kang IC, Kuramitsu HK (2002). Induction of monocyte chemoattractant protein-1 by Porphyromonas gingivalis in human endothelial cells. *FEMS Immunol Med Microbiol* **34**: 311–317.
- Kasser UR, Gleissner C, Dehne F, Michel A, Willershausen-Zonnchen B, Bolten WW (1997). Risk for periodontal disease in patients with longstanding rheumatoid arthritis. *Arthritis Rheum* **40**: 2248–2251.
- Khlgatian M, Nassar H, Chou HH, Gibson FC III, Genco CA (2002). Fimbria-dependent activation of cell adhesion molecule expression in Porphyromonas gingivalis-infected endo-thelial cells. *Infect Immun* **70**: 257–267.
- Kozarov EV, Dorn BR, Shelburne CE, Dunn WA Jr, Progulske-Fox A (2005). Human atherosclerotic plaque contains viable invasive Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis. *Arterioscler Thromb Vasc Biol* **25**: e17–e18.

- Kozarov E, Sweier D, Shelburne C, Progulske-Fox A, Lopatin D (2006). Detection of bacterial DNA in atheromatous plaques by quantitative PCR. *Microbes Infect* **8:** 687–693.
- Kuramitsu HK, Qi M, Kang IC, Chen W (2001). Role for periodontal bacteria in cardiovascular diseases. Ann Periodontol 6: 41–47.
- Kweider M, Lowe GD, Murray GD, Kinane DF, McGowan DA (1993). Dental disease, fibrinogen and white cell count; links with myocardial infarction? *Scott Med J* 38: 73–74.
- Lester SR, Bain JL, Serio FG, Harrelson BD, Johnson RB (2009). Relationship between gingival angiopoietin-1 concentrations and depth of the adjacent gingival sulcus. *J Periodontol* **80:** 1447–1453.
- Li D, Liu L, Chen H, Sawamura T, Mehta JL (2003). LOX-1, an oxidized LDL endothelial receptor, induces CD40/CD40L signaling in human coronary artery endothelial cells. *Arterioscler Thromb Vasc Biol* **23**: 816–821.
- Li L, Michel R, Cohen J, Decarlo A, Kozarov E (2008). Intracellular survival and vascular cell-to-cell transmission of Porphyromonas gingivalis. *BMC Microbiol* **8**: 26.
- Libby P (2002). Inflammation in atherosclerosis. *Nature* **420**: 868–874.
- Libby P, Theroux P (2005). Pathophysiology of coronary artery disease. *Circulation* **111**: 3481–3488.
- Lockhart PB, Brennan MT, Thornhill M *et al* (2009). Poor oral hygiene as a risk factor for infective endocarditis-related bacteremia. *J Am Dent Assoc* **140**: 1238–1244.
- Loos BG (2005). Systemic markers of inflammation in periodontitis. *J Periodontol* **76:** 2106–2115.
- Lopez NJ, Quintero A, Llancaqueo M, Jara L (2009). [Effects of periodontal therapy on markers of systemic inflammation in patients with coronary heart disease risk]. *Rev Med Chil* 137: 1315–1322.
- Lucas V, Roberts GJ (2000). Odontogenic bacteremia following tooth cleaning procedures in children. *Pediatr Dent* 22: 96–100.
- Mahendra J, Mahendra L, Kurian VM, Jaishankar K, Mythilli R (2010). 16S rRNA-based detection of oral pathogens in coronary atherosclerotic plaque. *Indian J Dent Res* **21**: 248–252.
- Marcaccini AM, Novaes AB Jr, Meschiari CA *et al* (2009). Circulating matrix metalloproteinase-8 (MMP-8) and MMP-9 are increased in chronic periodontal disease and decrease after non-surgical periodontal therapy. *Clin Chim Acta* **409**: 117–122.
- Mason DP, Kenagy RD, Hasenstab D *et al* (1999). Matrix metalloproteinase-9 overexpression enhances vascular smooth muscle cell migration and alters remodeling in the injured rat carotid artery. *Circ Res* **85**: 1179–1185.
- Mattila KJ, Nieminen MS, Valtonen VV et al (1989). Association between dental health and acute myocardial infarction. BMJ 298: 779–781.
- Mattila KJ, Valtonen VV, Nieminen M, Huttunen JK (1995). Dental infection and the risk of new coronary events: prospective study of patients with documented coronary artery disease. *Clin Infect Dis* **20**: 588–592.
- Mattila KJ, Pussinen PJ, Paju S (2005). Dental infections and cardiovascular diseases: a review. *J Periodontol* **76**: 2085–2088.
- Mengel R, Bacher M, Flores-De-Jacoby L (2002). Interactions between stress, interleukin-1beta, interleukin-6 and cortisol in periodontally diseased patients. J Clin Periodontol 29: 1012–1022.
- Mercado FB, Marshall RI, Klestov AC, Bartold PM (2001). Relationship between rheumatoid arthritis and periodontitis. *J Periodontol* **72:** 779–787.

- Meyle J (1993). Neutrophil chemotaxis and serum concentration of tumor-necrosis-factor-alpha (TNFA). *J Periodontal Res* **28**: 491–493.
- Morrison HI, Ellison LF, Taylor GW (1999). Periodontal disease and risk of fatal coronary heart and cerebrovascular diseases. *J Cardiovasc Risk* **6**: 7–11.
- Mustapha IZ, Debrey S, Oladubu M, Ugarte R (2007). Markers of systemic bacterial exposure in periodontal disease and cardiovascular disease risk: a systematic review and meta-analysis. *J Periodontol* **78**: 2289–2302.
- Nakajima T, Honda T, Domon H *et al* (2010). Periodontitisassociated up-regulation of systemic inflammatory mediator level may increase the risk of coronary heart disease. *J Periodontal Res* **45:** 116–122.
- Nakamura N, Yoshida M, Umeda M et al (2008). Extended exposure of lipopolysaccharide fraction from Porphyromonas gingivalis facilitates mononuclear cell adhesion to vascular endothelium via Toll-like receptor-2 dependent mechanism. *Atherosclerosis* **196**: 59–67.
- Nakano K, Inaba H, Nomura R *et al* (2006). Detection of cariogenic Streptococcus mutans in extirpated heart valve and atheromatous plaque specimens. *J Clin Microbiol* **44**: 3313–3317.
- Nassar H, Chou HH, Khlgatian M, Gibson FC III, Van Dyke TE, Genco CA (2002). Role for fimbriae and lysine-specific cysteine proteinase gingipain K in expression of interleukin-8 and monocyte chemoattractant protein in Porphyromonas gingivalis-infected endothelial cells. *Infect Immun* **70**: 268–276.
- Nonnenmacher C, Stelzel M, Susin C *et al* (2007). Periodontal microbiota in patients with coronary artery disease measured by real-time polymerase chain reaction: a case-control study. *J Periodontol* **78**: 1724–1730.
- Offenbacher S, Beck JD, Moss K *et al* (2009). Results from the Periodontitis and Vascular Events (PAVE) Study: a pilot multicentered, randomized, controlled trial to study effects of periodontal therapy in a secondary prevention model of cardiovascular disease. *J Periodontol* **80**: 190–201.
- Packard RR, Libby P (2008). Inflammation in atherosclerosis: from vascular biology to biomarker discovery and risk prediction. *Clin Chem* **54:** 24–38.
- Papapanou PN, Neiderud AM, Disick E, Lalla E, Miller GC, Dahlen G (2004). Longitudinal stability of serum immunoglobulin G responses to periodontal bacteria. J Clin Periodontol 31: 985–990.
- Paraskevas S, Huizinga JD, Loos BG (2008). A systematic review and meta-analyses on C-reactive protein in relation to periodontitis. *J Clin Periodontol* **35:** 277–290.
- Pearson TA, Mensah GA, Alexander RW *et al* (2003). Markers of inflammation and cardiovascular disease: application to clinical and public health practice: A statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. *Circulation* **107**: 499–511.
- Pincus T, Callahan LF, Sale WG, Brooks AL, Payne LE, Vaughn WK (1984). Severe functional declines, work disability, and increased mortality in seventy-five rheumatoid arthritis patients studied over nine years. *Arthritis Rheum* 27: 864–872.
- Pucar A, Milasin J, Lekovic V *et al* (2007). Correlation between atherosclerosis and periodontal putative pathogenic bacterial infections in coronary and internal mammary arteries. *J Periodontol* **78:** 677–682.
- Pussinen PJ, Jousilahti P, Alfthan G, Palosuo T, Asikainen S, Salomaa V (2003). Antibodies to periodontal pathogens are associated with coronary heart disease. *Arterioscler Thromb Vasc Biol* **23**: 1250–1254.

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- Pussinen PJ, Alfthan G, Tuomilehto J, Asikainen S, Jousilahti P (2004). High serum antibody levels to Porphyromonas gingivalis predict myocardial infarction. *Eur J Cardiovasc Prev Rehabil* 11: 408–411.
- Pussinen PJ, Nyyssonen K, Alfthan G, Salonen R, Laukkanen JA, Salonen JT (2005). Serum antibody levels to Actinobacillus actinomycetemcomitans predict the risk for coronary heart disease. *Arterioscler Thromb Vasc Biol* 25: 833–838.
- Pussinen PJ, Alfthan G, Jousilahti P, Paju S, Tuomilehto J (2007). Systemic exposure to Porphyromonas gingivalis predicts incident stroke. *Atherosclerosis* **193**: 222–228.
- Qi M, Miyakawa H, Kuramitsu HK (2003). Porphyromonas gingivalis induces murine macrophage foam cell formation. *Microb Pathog* 35: 259–267.
- Rahman AU, Rashid S, Noon R *et al* (2005). Prospective evaluation of the systemic inflammatory marker C-reactive protein in patients with end-stage periodontitis getting teeth replaced with dental implants: a pilot investigation. *Clin Oral Implants Res* **16**: 128–131.
- Renvert S, Pettersson T, Ohlsson O, Persson GR (2006). Bacterial profile and burden of periodontal infection in subjects with a diagnosis of acute coronary syndrome. *J Periodontol* 77: 1110–1119.
- Ridker PM, Danielson E, Fonseca FA *et al* (2008). Rosuvastatin to prevent vascular events in men and women with elevated C-reactive protein. *N Engl J Med* **359**: 2195–2207.
- Roberts FA, Hockett RD Jr, Bucy RP, Michalek SM (1997a). Quantitative assessment of inflammatory cytokine gene expression in chronic adult periodontitis. *Oral Microbiol Immunol* **12:** 336–344.
- Roberts FA, McCaffery KA, Michalek SM (1997b). Profile of cytokine mRNA expression in chronic adult periodontitis. *J Dent Res* 76: 1833–1839.
- Robertson AK, Hansson GK (2006). T cells in atherogenesis: for better or for worse? *Arterioscler Thromb Vasc Biol* 26: 2421–2432.
- Roth GA, Moser B, Huang SJ *et al* (2006). Infection with a periodontal pathogen induces procoagulant effects in human aortic endothelial cells. *J Thromb Haemost* **4**: 2256–2261.
- Rupprecht HJ, Blankenberg S, Bickel C *et al* (2001). Impact of viral and bacterial infectious burden on long-term prognosis in patients with coronary artery disease. *Circulation* **104**: 25–31.
- Saito Y, Fujii R, Nakagawa KI, Kuramitsu HK, Okuda K, Ishihara K (2008). Stimulation of Fusobacterium nucleatum biofilm formation by Porphyromonas gingivalis. Oral Microbiol Immunol 23: 1–6.
- Sakellari D, Socransky SS, Dibart S, Eftimiadi C, Taubman MA (1997). Estimation of serum antibody to subgingival species using checkerboard immunoblotting. *Oral Microbiol Immunol* 12: 303–310.
- Sattar N, McCarey DW, Capell H, McInnes IB (2003). Explaining how "high-grade" systemic inflammation accelerates vascular risk in rheumatoid arthritis. *Circulation* **108**: 2957–2963.
- Sheets SM, Potempa J, Travis J, Casiano CA, Fletcher HM (2005). Gingipains from Porphyromonas gingivalis W83 induce cell adhesion molecule cleavage and apoptosis in endothelial cells. *Infect Immun* **73**: 1543–1552.
- Spahr A, Klein E, Khuseyinova N *et al* (2006). Periodontal infections and coronary heart disease: role of periodontal bacteria and importance of total pathogen burden in the Coronary Event and Periodontal Disease (CORODONT) study. *Arch Intern Med* **166**: 554–559.
- Stinson MW, Alder S, Kumar S (2003). Invasion and killing of human endothelial cells by viridans group streptococci. *Infect Immun* **71**: 2365–2372.

- Takahashi Y, Davey M, Yumoto H, Gibson FC III, Genco CA (2006). Fimbria-dependent activation of pro-inflammatory molecules in Porphyromonas gingivalis infected human aortic endothelial cells. *Cell Microbiol* **8**: 738–757.
- Taylor BA, Tofler GH, Carey HM *et al* (2006). Full-mouth tooth extraction lowers systemic inflammatory and thrombotic markers of cardiovascular risk. *J Dent Res* 85: 74–78.
- Teles R, Sakellari D, Teles F *et al* (2010a). Relationships among gingival crevicular fluid biomarkers, clinical parameters of periodontal disease, and the subgingival microbiota. *J Periodontol* **81:** 89–98.
- Teles RP, Gursky LC, Faveri M *et al* (2010b). Relationships between subgingival microbiota and GCF biomarkers in generalized aggressive periodontitis. *J Clin Periodontol* **37**: 313–323.
- Tomas I, Alvarez M, Limeres J, Potel C, Medina J, Diz P (2007). Prevalence, duration and aetiology of bacteraemia following dental extractions. *Oral Dis* 13: 56–62.
- Tonetti MS, D'Aiuto F, Nibali L *et al* (2007). Treatment of periodontitis and endothelial function. *N Engl J Med* **356**: 911–920.
- Tuomainen AM, Jauhiainen M, Kovanen PT, Metso J, Paju S, Pussinen PJ (2008). Aggregatibacter actinomycetemcomitans induces MMP-9 expression and proatherogenic lipoprotein profile in apoE-deficient mice. *Microb Pathog* 44: 111–117.
- Tuominen R, Reunanen A, Paunio M, Paunio I, Aromaa A (2003). Oral health indicators poorly predict coronary heart disease deaths. *J Dent Res* 82: 713–718.
- Ushida Y, Koshy G, Kawashima Y *et al* (2008). Changes in serum interleukin-6, C-reactive protein and thrombomodulin levels under periodontal ultrasonic debridement. *J Clin Periodontol* **35**: 969–975.
- Vidal F, Figueredo CM, Cordovil I, Fischer RG (2009). Periodontal therapy reduces plasma levels of interleukin-6, C-reactive protein, and fibrinogen in patients with severe periodontitis and refractory arterial hypertension. J Periodontol 80: 786–791.
- Wu T, Trevisan M, Genco RJ, Falkner KL, Dorn JP, Sempos CT (2000). Examination of the relation between periodontal health status and cardiovascular risk factors: serum total and high density lipoprotein cholesterol, C-reactive protein, and plasma fibrinogen. *Am J Epidemiol* **151**: 273–282.
- Yoshii S, Tsuboi S, Morita I *et al* (2009). Temporal association of elevated C-reactive protein and periodontal disease in men. *J Periodontol* **80**: 734–739.
- Yumoto H, Chou HH, Takahashi Y, Davey M, Gibson FC III, Genco CA (2005). Sensitization of human aortic endothelial cells to lipopolysaccharide via regulation of Toll-like receptor 4 by bacterial fimbria-dependent invasion. *Infect Immun* 73: 8050–8059.
- Yumoto H, Yamada M, Shinohara C *et al* (2007). Soluble products from Eikenella corrodens induce cell proliferation and expression of interleukin-8 and adhesion molecules in endothelial cells via mitogen-activated protein kinase pathways. *Oral Microbiol Immunol* **22**: 36–45.
- Zhang T, Kurita-Ochiai T, Hashizume T, Du Y, Oguchi S, Yamamoto M (2010). Aggregatibacter actinomycetemcomitans accelerates atherosclerosis with an increase in atherogenic factors in spontaneously hyperlipidemic mice. *FEMS Immunol Med Microbiol* **59**: 143–51.
- Zhu J, Quyyumi AA, Norman JE *et al* (2000). Effects of total pathogen burden on coronary artery disease risk and C-reactive protein levels. *Am J Cardiol* **85:** 140–146.
- Zhu J, Nieto FJ, Horne BD, Anderson JL, Muhlestein JB, Epstein SE (2001). Prospective study of pathogen burden and risk of myocardial infarction or death. *Circulation* **103**: 45–51.

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