ORIGINAL ARTICLE

N Suda A Bazar O Bold B Jigjid A Garidkhuu G Ganburged K Moriyama

Authors' affiliations:

N. Suda, G. Ganburged, K. Moriyama, Maxillofacial Orthognathics, Division of Maxillofacial/Neck Reconstruction, Department of Maxillofacial Reconstruction and Function, Graduate School of Medical and Dental Sciences, Tokyo Medical and Dental University, Tokyo, Japan N. Suda, G. Ganburged, K. Moriyama, Global Center of Excellence (GCOE) Program, International Research Center for Molecular Science in Tooth and Bone Diseases, Tokyo, Japan

A. Bazar, O. Bold, B. Jigjid, School of Dentistry, Health Sciences University of Mongolia, Ulaanbaatar, Mongolia A. Garidkhuu, Graduate Studies, Health Sciences University of Mongolia, Ulaanbaatar, Mongolia

Correspondence to:

Naoto Suda Maxillofacial Orthognathics Division of Maxillofacial/Neck Reconstruction Department of Maxillofacial Reconstruction and Function Graduate School of Medical and Dental Sciences Tokyo Medical and Dental University 1-5-45 Yushima Bunkyo-ku Tokyo 113-8510 Japan E-mail: n-suda.mort@tmd.ac.jp

Dates:

Accepted 7 January 2010

To cite this article:

Suda N, Bazar A, Bold O, Jigjid B, Garidkhuu A, Ganburged G, Moriyama K: A Mongolian patient with hypohidrotic ectodermal dysplasia with a novel P121S variant in *EDARADD Orthod Craniofac Res* 2010;**13**:114–117

© 2010 John Wiley & Sons A/S

A Mongolian patient with hypohidrotic ectodermal dysplasia with a novel P121S variant in *EDARADD*

Structured Abstract

Authors – Suda N, Bazar A, Bold O, Jigjid B, Garidkhuu A, Ganburged G, Moriyama K

Introduction – Hypohidrotic ectodermal dysplasia is a genetic disorder characterized by diminished or a lack of sweating, congenital missing teeth, and sparse or absent hair. Three genes, *EDA*, *EDAR*, and *EDARADD*, all related to tumor necrosis factor signaling, have been reported as responsible genes for this disorder. Among them, the largest numbers of mutations have been identified in *EDA*, and only two mutations identified in *EDARADD*.

Materials and Methods – DNA analysis of *EDA*, *EDAR*, and *EDARADD* was performed on a Mongolian patient by polymerase chain reaction-direct sequencing. *Results* – The 5-year-old Mongolian individual had no erupted deciduous or permanent teeth. A panoramic radiograph showed only one tooth in the right mandible. His hair and eyebrows were sparse, but he did not have a short stature. He showed diminished sweating. The nails of his fingers and toes were normal. Based on these conditions, he was diagnosed with hypohidrotic ectodermal dysplasia. There was no gene mutation of *EDA* or *EDAR*. A novel heterozygous variant (P121S; c.361C > T) was identified in the death domain of *EDARADD* (NM_080738). No other member of his family was affected, and this variant was not identified in his parents or maternal grandparents.

Conclusion – This study reports an individual affected with hypohidrotic ectodermal dysplasia with a novel heterozygous P121S variant in the death domain of *EDARADD*.

Key words: death domain; ectodermal dysplasia; *EDARADD*; gene mutation; tumor necrosis factor

Introduction

Ectodermal dysplasias are heterogeneous genetic disorders that display different combinations of congenital defects in teeth, nails, sweat glands, and hair (1, 2). The hypohidrotic (or anhidrotic) ectodermal dysplasias (MIM #129490, #224900, #305100) are a subgroup showing abnormalities of the eccrine sweat glands.

It has been reported that the ectodysplasin (Eda) pathway initiates development of the hair, teeth, and other ectodermal derivatives (3, 4). Three genes, *EDA*, *EDAR*, and *EDARADD*, are related to this pathway. *EDA* encodes Eda, which is a ligand of Edar (encoded by *EDAR*) belonging to the tumor necrosis factor (TNF) receptor family (5). The adaptor protein, Edaradd, encoded by *EDARADD*, interacts with the death domain of Edar and transduces the downstream signaling (6–8). Both Edar and Edaradd are known to have a death domain related to the apoptosis signal (6, 8, 9).

The X-linked form of hypohidrotic ectodermal dysplasia (MIM #305100) is mainly caused by *EDA* mutation. The autosomal hypohidrotic ectodermal dysplasias (MIM #129490 and #224900) are caused by the mutations of *EDAR* and *EDARADD*. Among them, only two mutations have been identified in *EDARADD* (6, 8). One is a homozygous E142K mutation in the death domain (6), and the other is a heterozygous L112R mutation (8).

In this report, we describe an individual affected by hypohidrotic ectodermal dysplasia with a novel heterozygous P121S variant (c.361C > T) in the death domain of *EDARADD*.

Materials and methods

Genomic DNA was isolated from nail samples from the affected patient, his parents, and maternal grandparents, after obtaining informed consent. Isolated DNA was amplified by the polymerase chain reaction (GeneAmp PCR System 9700; Applied Biosystems, Foster City, CA, USA) using specific primers for *EDA* (10), *EDAR* (11), and *EDARADD* (8). Amplified products were sequenced using BigDye Terminator v.3.1 cycle sequencing kit (Applied Biosystems).

Results

The 5-year-old Mongolian boy (III-6) was born to healthy parents, and his birth weight was 2980 g (Fig. 1). He was a monozygotic twin, and his counterpart (III-7) had died of cardiac failure at 1 years of age (Fig. 1). Because of his short life, it was not clear whether the counterpart had also been affected with ectodermal dysplasia. His father (II-1) had six younger

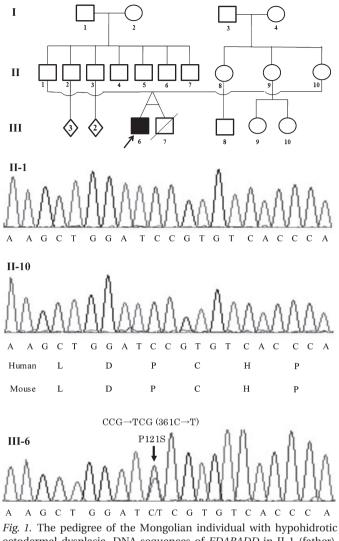


Fig. 1. The pedigree of the Mongolian individual with hyponidrotic ectodermal dysplasia. DNA sequences of *EDARADD* in II-1 (father), II-10 (mother), and III-6 (proband) are shown. III-6 showed a heterozygous P121S (c.361C > T) variant, which was not in II-1 or II-10. N-terminus amino acid sequences of the death domain (start from L119) of human and mouse Edaradd are shown under II-10.

brothers, and his mother (II-10) had two older sisters. No family member was affected with ectodermal dysplasia.

He did not show short stature, and his mental development was normal. His hair and eyebrows were sparse. The nails on his fingers were normal, but he was unable to sweat. He did not show any cardiac abnormality. He had no erupted deciduous or permanent teeth. A lateral cephalogram and panoramic radiograph showed only one mandibular molar on the right side (Fig. 2). The alveolar bone height of the maxilla and mandible was extremely low. Based on these conditions, he was diagnosed with hypohidrotic ectodermal dysplasia.

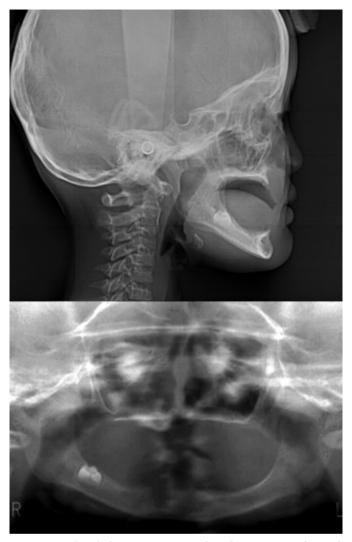


Fig. 2. Lateral cephalogram (upper panel) and panoramic radiograph (lower panel) of the Mongolian individual with hypohidrotic ectodermal dysplasia. Note the extremely short alveolar height of both jaws and only one tooth in the right mandible.

There was no mutation of *EDA* (NG_009809) or *EDAR* (NG_00825). In *EDARADD* gene (NM_080738), there was a heterozygous P121S variant (c.361C > T, starting from ATG) (Fig. 1). P121 is located in the death domain, which starts from the 119th amino acid, in *EDARADD* (Fig. 2). This variant had not been credited in the NCBI database for SNPs. This variant was not found in his parents (father: II-1, mother: II-10) or maternal grandparents (grandfather: I-3, grandmother: I-4).

Discussion

Hypohidrotic ectodermal dysplasia is known to have varying degrees of ectodermal defects such as hyp-

odontia, alopecia, hypohydrosis, and nail abnormalities (1, 2). This patient exhibited sparse hair and eyebrows, an inability to sweat, and severe oligodontia (Fig. 2). Although he had no family history or nail abnormalities, these clinical symptoms were sufficient to diagnose him with a hypohidrotic ectodermal dysplasia.

Three genes, *EDA*, *EDAR*, and *EDARADD*, have so far been associated with hypohidrotic ectodermal dysplasia. Among them, only two mutations have been identified in *EDARADD*: a heterozygous L112R mutation (8) and a homozygous E142K mutation (6). Edar can interact with Edaradd, and it has been reported that this interaction was markedly impaired by L112R (8). The principle function of the death domain is to induce apoptosis (9), and this domain starts from the amino acid position of L119 in Edaradd (Fig. 1), as previously reported (6). E142 is located in this domain, and E142K mutation is reported to diminish the interaction with Edar (6).

The Eda pathway is related to hair and tooth development (12). Mouse models of hypohidrotic ectodermal dysplasia (known as tabby, downless, sleek, and crinkled) are known (13). The cloning of Eda signaling demonstrated that Eda, Edar, and Edaradd were mutated in these mutant mice (4, 6, 14, 15). These mutant mice show abnormalities of tooth, hair, and sweat glands, which are similar to those in hypohidrotic ectodermal dysplasia. As the downstream pathway of Edaradd, another adaptor molecule, TNFRassociated factor 6 (TRAF6), recruits TGF- β activated kinase (TAK1), and finally activates NF κ -B (16). NF κ -B is thought to be a transcriptional regulator, and some genes have been reported as targets of NF κ -B. These genes include *Bmp4* (Bone morphogenic protein-4), Dkk4 (Dickkopf-4), and Shh (Sonic hedgehog), and all of them are related to hair, skin, or tooth development (12).

The death domain of *ERARADD* is highly conserved among species, indicating an important function of this domain (6). The P121S variant has not credited in the NCBI Web site for SNPs. Moreover, this variant was not seen in the parents or grandparents of the affected patient. Considering these points, it is likely that the P121S variant was a *de novo* germ-line mutation of either II-1 or II-10, and it was responsible for hypohidrotic ectodermal dysplasia in III-6. However, the frequency and function of this variant is not known. P121S might be merely a polymorphism in *EDARADD*, which is more than 1% in the general population. Studies are required to clarify the frequency of the P121S variant together with its gene function during hair and tooth development.

Acknowledgement: This study was supported by a Grantin-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan (19406031 and 21390546).

References

- Pinheiro M, Freire-Maia N. Ectodermal dysplasias: a clinical classification and a causal review. *Am J Med Genet* 1994;53:153– 62.
- Zonana J. ED1, EDAR, and EDARADD and the hypohidrotic ectodermal dysplasia and ectodysplasin signaling pathway. In: Epstein CL, Erickson RP, Wynshaw-Boris A, editors. *Inborn Errors* of *Development*. New York, NY, USA: Oxford University Press; 2004. pp. 359–66.
- Headon DJ, Overbeek PA. Involvement of a novel Tnf receptor homologue in hair follicle induction. *Nat Genet* 1999;22:370–4.
- Monreal AW, Ferguson BM, Headon DJ, Street SL, Overbeek PA, Zonana J. Mutations in the human homologue of mouse dl cause autosomal recessive and dominant hypohidrotic ectodermal dysplasia. *Nat Genet* 1999;22:366–9.
- 5. Kumar A, Eby MT, Sinha S, Jasmin A, Chaudhary PM. The ectodermal dysplasia receptor activates the nuclear factor-kappaB, JNK, and cell death pathways and binds to ectodysplasin A. *J Biol Chem* 2001;276:2668–77.
- Headon DJ, Emmal SA, Ferguson BM, Tucker AS, Justice MJ, Sharpe PT et al. Gene defect in ectodermal dysplasia implicates a death domain adapter in development. *Nature* 2001;414:913–6.

- Locksley RM, Killeen N, Lenardo MJ. The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell* 2001;104:487– 501.
- 8. Bal E, Baala L, Cluzeau C, El Kerch F, Ouldim K, Hadj-Rabia S et al. Autosomal dominant anhidrotic ectodermal dysplasias at the *EDARADD* locus. *Hum Mutat* 2007;28:703–9.
- 9. Cleveland JL, Ihle JN. Contenders in FasL/TNF death signaling. *Cell* 1995;81:479–82.
- Zhang XJ, Chen JJ, Song YX, Yang S, Xiong XY, Zhang AP et al. Mutation analysis of the ED1 gene in two Chinese Han families with X-linked hypohidrotic ectodermal dysplasia. *Arch Dermatol Res* 2003;295:38–42.
- 11. Lind LK, Stecksén-Blicks C, Lejon K, Schmitt-Egenolf M. EDAR mutation in autosomal dominant hypohidrotic ectodermal dysplasia in two Swedish families. *BMC Med Genet* 2006;7:80.
- 12. Mikkola ML. Molecular aspects of hypohidrotic ectodermal dysplasia. *Am J Med Genet A* 2009;149:2031–6.
- 13. Mikkola ML, Thesleff I. Ectodysplasin signaling in development. *Cytokine Growth Factor Rev* 2003;14:211–24.
- 14. Srivastava AK, Pispa J, Hartung AJ, Du Y, Ezer S, Jenks T et al. The Tabby phenotype is caused by mutation in a mouse homologue of the EDA gene that reveals novel mouse and human exons and encodes a protein (ectodysplasin-A) with collagenous domains. *Proc Natl Acad Sci USA* 1997;94:13069–74.
- 15. Yan M, Zhang Z, Brady JR, Schilbach S, Fairbrother WJ, Dixit VM. Identification of a novel death domain-containing adaptor molecule for ectodysplasin-A receptor that is mutated in crinkled mice. *Curr Biol* 2002;12:409–13.
- Morlon A, Munnich A, Smahi A. TAB2, TRAF6 and TAK1 are involved in NF-kappaB activation induced by the TNF-receptor, Edar and its adaptator Edaradd. *Hum Mol Genet* 2005;14:3751–7.

Copyright of Orthodontics & Craniofacial Research is the property of Wiley-Blackwell and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.