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## Genetic and treatment-related risk factors associated with external apical root resorption (EARR) concurrent with orthodontia

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### Structured Abstract

**Objective** – As genetic variation accounts for two-thirds of the variation in external apical root resorption (EARR) concurrent with orthodontic treatment, we analyzed the association of selected genetic and treatment-related factors with EARR concurrent with orthodontic treatment.

**Setting and sample population** – This case–control study of 134 unrelated, orthodontically treated Caucasian individuals was conducted in part at an Indiana Private Practice, Indiana University and the University of Kentucky.

**Methods** – Utilizing a research data bank containing information from ~1450 orthodontically treated patients, pre- and post-treatment radiographs from 460 individuals were evaluated for EARR of the four permanent maxillary incisors. Sixty-seven unrelated Caucasians with moderate to severe EARR were identified and were age-/sex-matched with orthodontically treated Caucasian controls yielding 38 females and 29 males per group. Factors tested for an association with EARR included the following: 1) treatment duration, 2) extraction of maxillary premolars, 3) numerous cephalometric measurements, and 4) DNA polymorphisms within/near candidate genes in a pathway previously implicated in EARR such as the purinergic-receptor-P2X, ligand-gated ion channel 7 (*P2RX7*; rs208294, rs1718119, and rs2230912), caspase-1 (*CASP1*; rs530537, rs580253, and rs554344), interleukin-1 beta (*IL1B*; rs1143634),

interleukin-1 alpha (*IL1A*; rs1800587), and interleukin-1 receptor antagonist (*IL1RA*; rs419598) genes. Stepwise logistic regression was utilized to identify the factors significantly associated (significance taken at or less than the layered Bonferroni correction alpha) with the occurrence of EARR.

**Results** – A long length of treatment and the presence of specific genotypes for *P2RX7* SNP rs208294 were significantly associated with EARR.

**Conclusion** – EARR occurrence was associated with both genetic and treatment-related variables, which together explained 25% of the total variation associated with EARR in the sample tested.

**Key words:** caspase-1; external apical root resorption; interleukin-1; interleukin-1 receptor antagonist; purinergic-receptor-P2X ligand-gated ion channel 7

## Introduction

External apical root resorption (EARR) is a reduction of root structure involving the apices. EARR occurrence has been reported with and without orthodontic treatment (1) and can be diagnosed by orthodontists during routine diagnostic, progress and post-treatment radiographs. It has been reported that 5 mm or more of apical root resorption may occur in 5% of orthodontic patients (1). Any unusual length of the roots should be noted on initial diagnostic radiographs and described to the patient, along with an explanation of how EARR may occur as an inflammatory process resulting in the loss of apical root structure and what factors are contributory. Although root resorption may introduce mobility of involved teeth in severe cases, it usually does not have a significant impact on the longevity of teeth (2). Orthodontic force magnitude, duration of the applied force, force direction, dilacerated, slender or pointed roots, allergy, history of trauma, age at start of treatment, excessive overjet, maxillary premolar extractions, sex, and length of treatment have been reported as risk factors for EARR during the course of orthodontic treatment (3–9).

Although orthodontic tooth movement, or ‘biomechanics’, has been found to account for approximately one-tenth to one-third of the total variation in EARR (10–12), Owman-Moll et al. showed that individual variation overshadowed

the force magnitude and the force type in defining the susceptibility to histologic root resorption associated with orthodontic force (13). Individual variations were considerable regarding both extension and depth of histologic root resorption within individuals, and these were not correlated to the magnitude of tooth movement achieved (14). There was considerable individual variation in the EARR associated with orthodontic treatment, indicating an individual predisposition and multifactorial etiology (15–21). While the severity of EARR could not be fully explained by treatment-related factors, studies were initiated to investigate the association of genetic factors with the variation in EARR seen among individuals. Harris et al. (15) initially pointed out the involvement of genetic variation in EARR concurrent with orthodontia variation through a heritability study. Later, Hartsfield et al. (16) confirmed that genetic variation was associated with as much as 50–66% of the variation observed with EARR concurrent with orthodontia.

As compression of periodontal ligament (PDL) occurs when teeth are being moved, ATP is released from adjacent cells. During the process of binding this released extracellular ATP, the purinergic-receptor-P2X, ligand-gated ion channel 7 (P2RX7) on the cell surface of macrophage and/or monocytes change confirmation creating channels within the cell membrane that facilitate the exchange or changes of potassium (K), sodium

(Na), and/or calcium (Ca) ions from within/outside of the cell. During this ion exchange process, K ions are lost from the cell while intracellular Ca ions increase. This leads to the activation of the inflammasome complex within the cell which contains the caspase-1 enzyme (also known as interleukin-converting enzyme or ICE, which is coded for by the *CASP1* gene). Activated caspase-1 mediates IL1 $\beta$  maturation and the release of active IL1 $\beta$  (22). Based upon literature reports which suggested an association between adult onset chronic periodontitis and the +3953 genetic variant in the *IL1B* gene (23) (also termed +3954 or rs1143634), this variation was investigated and reported to be connected to EARR concurrent with orthodontia by trio genetic association and sib-pair linkage studies (24).

The *IL1B* +3953 genetic variant consists of individuals having either two arginine (AA) nucleic acid bases, two guanine (GG) nucleic acid bases, or a combination of the two (GA) at that particular genetic location. The other polymorphisms investigated for association with orthodontic patient EARR will also have the same possible combination of nucleic acids, or could have a combination of thymine (T), or cytosine (C) as illustrated in Table 2C. Some of the polymorphisms tested were functional, meaning the change in nucleic acids results in a change in protein function. These functional polymorphisms, as well as the non-functional ones, can also be markers for other genetic variations that may affect orthodontic patient EARR. In addition to the *IL1B* gene which codes for IL1 $\beta$  protein, the *IL1* gene locus also contains the *IL1A* and *IL1RN* genes, which code for the IL1 $\alpha$  and IL1 receptor antagonist proteins, respectively (25).

For this study, we hypothesized that by combining the analysis of numerous polymorphisms within and/or near the *P2RX7*, *CASP1*, *IL1B*, *IL1A*, and *IL1RA* genes, together with distinct treatment parameters, specific risk factors could be defined which contribute to in EARR in a population of orthodontic patients. A combined analysis of both multiple genetic and clinical factors potentially associated with EARR with orthodontia was performed in a case-control design format.

## Materials and methods

### Subject population

Approval for the study was obtained from the Institutional Review Boards of both Indiana University-Purdue University Indianapolis (IUPUI, Indianapolis, IN, USA) and the University of Kentucky (Lexington, KY, USA). A research data bank consisting of patient records and DNA was generated within a northern Indiana private orthodontic practice and contained information from 1458 individuals who received standard phase II orthodontic treatment. All patients were treated using conventional fixed edgewise appliances with a general wire sequence of 0.016-inch round nickel-titanium (Ni-Ti) to 0.016-inch round stainless steel to 0.016  $\times$  0.022 inch rectangular Ni-Ti, and were finished with 0.016  $\times$  0.022 inch rectangular wire in 0.018-inch slot brackets. Pre- and post-treatment lateral cephalometric, panoramic, and occlusal radiographs were taken as a part of each patient's standard of care and were available for research purposes. Four hundred and sixty Caucasian individuals were randomly selected from the research data bank and evaluated for EARR the four permanent maxillary incisors after the completion of orthodontic treatment (methodology described below). Individuals diagnosed with a syndrome, cleft lip/palate, showing open apices, and/or with a previous history of trauma were excluded. Individuals with missing treatment records and/or having non-diagnostic quality radiographs were also excluded. Sixty-seven subjects (38 females and 29 males) were identified with moderate to severe EARR (affected group). Among the patients with no EARR or only minimal EARR, 67 age- and gender-matched individuals were selected to serve as the control (unaffected) group.

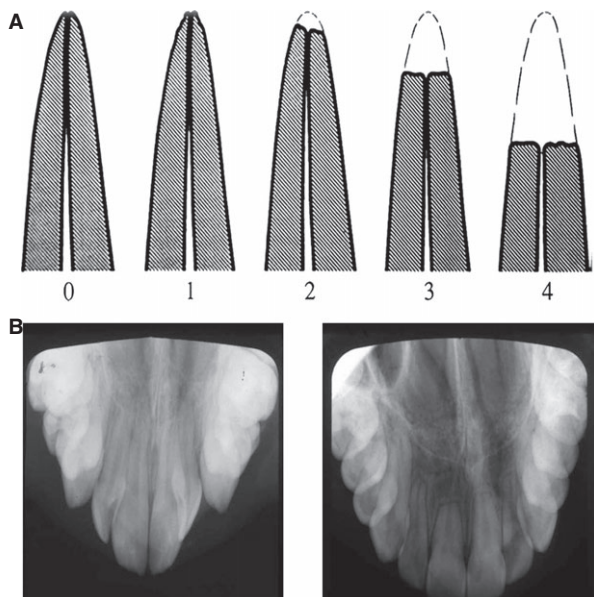
### Determination of EARR

Digitized pre- and post-treatment panoramic and occlusal radiographs for each subject were compared to determine whether EARR had occurred to 1 or more of the four maxillary incisor roots during orthodontic treatment. The

qualitative measures of Malmgren's grading system were used (26), where a rating of 0, 1, or 2 was considered control and a rating of 3 or 4 was graded as moderate or severe EARR (Fig. 1). Ratings of 3 or 4 represented EARR beyond blunting that involved the apical third or more of the tooth root. The evaluation of EARR was performed by three independent investigators in a blinded manner. When at least two of the three examiners agreed that the subject had EARR of 1 or more maxillary incisors, the patient was classified as being affected. All others were classified as being non-affected, even if slight blunting was present.

#### Measurement and/or assessment of clinical and treatment-related factors

The length of treatment in months and type of treatment (extraction vs. non-extraction treatment) were recorded from patient records and confirmed on radiographs. Extraction of maxillary first premolars or maxillary second premolars was analyzed together and as unique variables in the statistical modeling. Digitalized



**Fig. 1.** Evaluation method of EARR. (A). Classification scale of Malmgren (26) for EARR; where 0 = no resorption, 1 = slight irregularities, 2 = resorption of less than apical third, 3 = resorption of apical third, and 4 = resorption beyond apical third. (B). An example of the pre-orthodontic (left) and post-orthodontic treatment (right) occlusal radiographs for a subject diagnosed with EARR.

cephalometric tracings were analyzed with Dolphin Imaging software (version 10.0; Dolphin Software Inc., Lake Oswego, OR, USA). Nine cephalometric variables were chosen as outlined in Table 1 and Fig. 2. The pre-treatment cephalometric values and the change ( $\Delta$ ) in cephalometric values at the end of treatment were considered unique variables in the statistical modeling. A decrease in the cephalometric value for incisor angulation with a negative (–) sign indicates an average retraction of those teeth.

#### Genetic analyses

Buccal swab samples were obtained from each patient by having the subject brush the inside of each cheek 10 times with two sterile nylon bristle

**Table 1.** Definitions of the cephalometric landmarks used

Cephalometric variable	Description
Overjet (OJ)	Horizontal distance between the incisal edges of the maxillary and mandibular central incisors.
Overbite (OB)	Vertical distance between the incisal edges of the maxillary and mandibular central incisors
ANB	Angle formed by hard tissue A-point (A) to Nasion (N) to hard tissue B-point (B)
FMA	Angle formed between Frankfort horizontal (FH) plane and Gonion (Go)–Menton (Me) plane
U1- FH	Angle formed between the long axis of most procumbent maxillary central incisor (Upper1, U1) and FH.
U1-NA	Angle formed between the long axis of U1 and a line connecting hard tissue Nasion and A-point (NA).
U1-NA	Distance between the most anterior labial point of U1 and NA line
U1-ptv	Distance between U1 tip and the pterygoid vertical (ptv) perpendicular to FH
U1r-ptv	Distance between U1 root tip (U1r) and the pterygoid vertical (ptv) perpendicular to FH



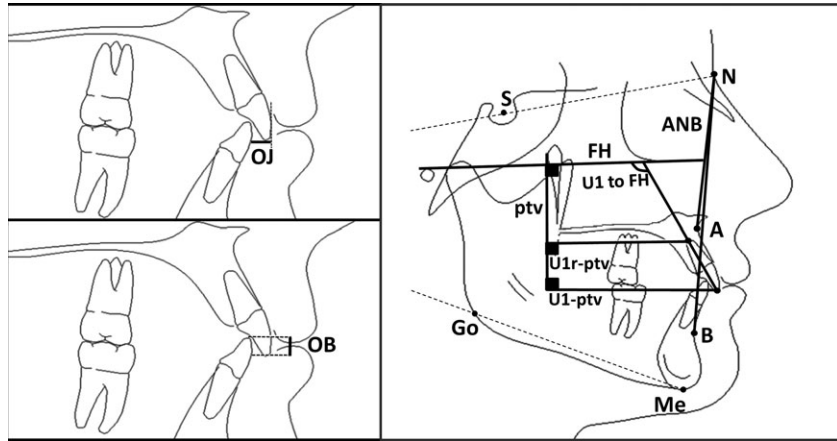


Fig. 2. An illustration showing the selected cephalometric landmarks used in the study.

brushes. Genomic DNA was isolated with the Puregene method (Gentra Systems, Minneapolis, MN, USA), and DNA concentrations were measured on the NanoDrop-1000 spectrophotometer (Thermo Fisher Scientific, Wilmington, DE). Taq-Man<sup>®</sup>-based genotyping was utilized for the allelic discrimination of all polymorphisms tested on the LightCycler480<sup>®</sup> Instrument (Roche, Indianapolis, IN, USA) using standard genotyping reagents (Life Technologies/Applied Biosystems, Grand Island, NY, USA). Single nucleotide polymorphisms (SNPs) were examined within and/or near the purinergic-receptor-P2X, ligand-gated ion channel 7 (*P2RX7*; rs208294, rs1718119, and rs2230912), caspase-1/interleukin-converting enzyme (*CASP1/ICE*; rs580253, rs554344, and rs530537), interleukin-1 beta (*IL1B*; rs1143634), interleukin-1 alpha (*IL1A*; rs1800587), and interleukin-1 receptor antagonist (*IL1RA*, rs419598) genes.

#### Statistical methods

The kappa statistic was utilized to measure the variation associated with three radiographic examiners independently determining a patient's EARR status. Genotyping departures in the unaffected group from Hardy-Weinberg equilibrium (HWE) were assessed using the chi-square test. Stepwise logistic regression analysis applying the *p*-value threshold stopping rule with JMP<sup>®</sup> PRO Statistical Analysis Software (version 10; SAS Institute Inc., Cary, NC, USA) was used to test for variables associated with EARR. In addition to the variables analyzed, the presence of an

interaction between the following pairs of variables was determined: 1) between the length of treatment and *P2RX7* SNP rs208294 genotype; and 2) between extraction of maxillary first premolars and *P2RX7* SNP rs208294 genotype. Statistical significance was when *p* was equal to or <0.05 as the initial alpha value, corrected for multiple comparisons as indicated by the layered Bonferroni technique of the nine variables in the final linear regression analysis of key variables influencing EARR (27).

## Results

The kappa value for the three EARR examiners involved in this study was 0.08, indicating a good agreement between the examiners regarding the EARR status of each subject. There were no age restrictions for study enrollment; subjects' age ranged from 8.4 to 55.4 years. The average age at the start of treatment  $\pm$  std error for the affected group was  $15.78 \pm 1.13$  years (median age = 12.87 years, mode = 12.23 years) and for the unaffected group was  $15.79 \pm 1.14$  years (median age = 12.89 years, mode = 12.15 years) (Table 2A). The average length of treatment  $\pm$  std error for the case and control groups was  $2.49 \pm 0.10$  years and  $1.97 \pm 1.14$  years, respectively. In the final linear regression modeling, the length of treatment was the largest variable influencing the occurrence of EARR in the population studied where longer treatment times increased the probability of EARR (Table 3;

**Table 2. (A) Environmental and treatment variables, (B) cephalometric variables, (C) genetic variables**

	Subjects with moderate to severe EARR (affected)	Subjects with little to no EARR (unaffected)
(A)		
Sex	38 females 29 males	38 females 29 males
Age at start of treatment in years (average + std error)	15.78 ± 1.13	15.79 ± 1.14
Number of years in treatment (average + std error)	2.49 + 0.10	1.97 + 0.08
Number of subjects with upper 1st Premolars extracted or missing (U4's)	20 of 67 subjects	7 of 67 subjects
Number of subjects with upper 2nd Premolars extracted or missing (U5's)	0 of 67 subjects	3 of 67 subjects
Number of subjects who had 1 or more maxillary premolars extracted or missing (U4 or U5)	20 of 67 subjects	10 of 67 subjects
(B)		
Average pre-treatment measures + std error		
OJ (mm)	4.21 + 0.31	4.56 + 0.25
OB (mm)	3.07 + 0.31	3.27 + 0.28
ANB (°)	3.31 + 0.33	3.37 + 0.25
FMA (°)	25.15 + 0.72	23.50 + 0.65
U1-FH (°)	112.17 + 0.94	112.40 + 0.94
U1-NA (°)	21.09 + 0.94	20.85 + 0.97
U1-NA (mm)	3.95 + 0.33	3.66 + 0.35
U1-ptv (mm)	59.68 + 0.74	60.23 + 0.68
U1r-ptv (mm)	51.31 + 0.60	51.38 + 0.70
Average change (Δ) in variable at the end of treatment + std error		
ΔOJ (mm)	1.71 + 0.27	1.87 + 0.27
ΔOB (mm)	1.76 + 0.30	1.92 + 0.25
ΔANB (°)	0.66 + 0.19	0.42 + 0.15
ΔFMA (°)	0.65 + 0.33	-0.30 + 0.37
ΔU1-FH (°)	-3.58 + 1.01	-3.38 + 0.94
ΔU1-NA (°)	-3.67 + 1.01	-3.34 + 1.01
ΔU1-NA (mm)	-0.59 + 0.40	-0.94 + 0.39
ΔU1-ptv (mm)	-1.34 + 0.48	-1.34 + 0.45
ΔU1r-ptv (mm)	0.06 + 0.25	-0.42 + 0.49

**Table 2. (continued)**

	Subjects with moderate to severe EARR (affected)			Subjects with little to no EARR (unaffected)		
(C)						
<i>P2RX7</i> , rs208294	CC	CT	TT	CC	CT	TT
(Functional polymorphism)	28 (41.8%)	33 (49.3%)	6 (9.0%)	15 (22.4%)	34 (50.7%)	18 (26.9%)
<i>P2RX7</i> , rs1718119	GG	GA	AA	GG	GA	AA
(Functional polymorphism)	32 (47.8%)	25 (37.3%)	10 (14.9%)	28 (41.8%)	32 (47.8%)	7 (10.4%)
<i>P2RX7</i> , rs2230912	AA	AG	GG	AA	AG	GG
(Functional polymorphism)	53 (79.1%)	13 (19.4%)	1 (1.5%)	43 (64.2%)	21 (31.3%)	3 (4.5%)
<i>CASP1/ICE</i> , rs530537	TT	TC	CC	TT	TC	CC
	24 (35.8%)	30 (44.8%)	13 (19.4%)	18 (26.9%)	36 (53.7%)	13 (19.4%)
<i>CASP1/ICE</i> , rs580253	GG	GA	AA	GG	GA	AA
(Functional polymorphism)	52 (77.6%)	14 (20.9%)	1 (1.5%)	46 (68.7%)	17 (25.4%)	4 (6.0%)
<i>CASP1/ICE</i> , rs554344	GG	GC	CC	GG	GC	CC
(Functional polymorphism)	52 (77.6%)	14 (20.9%)	1 (1.5%)	45 (67.2%)	18 (26.9%)	4 (6.0%)
<i>IL1β</i> , rs1143634	GG	GA	AA	GG	GA	AA
(Functional polymorphism)	37 (55.2%)	26 (38.8%)	4 (6.0%)	44 (65.7%)	21 (31.3%)	2 (3.0%)
<i>IL1RA</i> , rs419598	TT	TC	CC	TT	TC	CC
	41 (61.2%)	23 (34.3%)	3 (4.5%)	38 (56.7%)	29 (43.3%)	0 (–)
<i>IL1α</i> rs1800587	GG	GA	AA	GG	GA	AA
	28 (41.8%)	28 (41.8%)	11 (16.4%)	32 (47.8%)	29 (43.3%)	6 (9.0%)

Functional polymorphism refers to a polymorphism which either has been shown to alter the expression of the gene, or leads to an amino acid coding change which can influence the function or quantity of protein produced from the gene.

$p < 0.0001$ , significant at a layered Bonferroni correction of  $p = 0.006$ ).

Of the 67 EARR-affected individuals, 20 had maxillary first premolar extractions compared to only seven controls with first premolar extractions and three controls with maxillary second premolar extractions. Although this type of extraction appeared to play a significant role in influencing the occurrence of EARR (Table 3;  $p = 0.0469$ ), this was not significant when compared to the layered Bonferroni correction of  $p = 0.007$  for that variable. Due to the low number of second premolar extractions in this study population, and the fact that second premolar extractions only occurred within the control population, second premolar extractions were not factored into the final modeling analysis.

The average overall change in FMA did not have a statistically significant influence in EARR occurrence during orthodontic treatment (Table 3;  $p = 0.0591$ , compared to being statistically significant if at or less than the layered

Bonferroni correction of  $p = 0.01$ ). Average pre-treatment ANB angles  $\pm$  std error between the case and control groups were comparable at  $3.31^\circ \pm 0.33^\circ$  and  $3.37^\circ \pm 0.25^\circ$ , respectively (Table 2B), with the range of ANB measurements spanning from  $12^\circ$  to  $-4^\circ$ . The average retraction distance for the central incisor(s)  $\pm$  std error, as measured by the  $\Delta U1$ -NA, was  $-0.59 \pm 0.40$  mm for the case group, compared to  $-0.94 \pm 0.39$  mm for the controls (Table 2B). Within the case group, however, individuals who received maxillary first premolar extractions ( $n = 20$ ) had an average  $\Delta U1$ -NA of 1.62 mm compared to an average  $\Delta U1$ -NA of  $-1.54$  mm for individuals in the same group who did not receive maxillary first premolar extractions ( $n = 47$ ). Similarly, individuals in the control group who received maxillary first premolar extractions ( $n = 7$ ) had an average  $\Delta U1$ -NA of 3.31 mm, compared to  $-1.44$  mm for those in the control group who did not have maxillary first premolars removed during orthodontic treatment ( $n = 60$ ).

**Table 3. Final linear regression analysis of key variables influencing EARR**

Variables	p-value	Layered Bonferroni corrected significant alpha
Length of time in treatment (years)	<0.0001*	0.006
<i>P2RX7</i> , rs208294 {CC&CT vs. TT}	0.0028*	0.006
Upper 1st Premolars were extracted or missing	0.0469	0.007
<i>IL1β</i> , rs1143634 {AA&GA vs. GG}	0.0533	0.008
Change in FMA	0.0591	0.01
<i>P2RX7</i> , rs1718119 {AA&GG vs. GA}	0.0813	0.01
Change in Ur-pvt	0.0908	0.02
<i>CASP1/ICE</i> , rs530537 {TT vs. CC&TC}	0.1527	0.03
<i>P2RX7</i> , rs208294 {CC vs. CT}	0.1564	0.05

\*Statistically significant compared to alpha corrected by layered Bonferroni procedure; Extraction of second premolars was not included in the analysis. The factors summarized in the table explain 25.13% of the variation observed in this sample.

The average  $\Delta$ U1-NA angle ( $^{\circ}$ ) for individuals in the case group  $\pm$  std error was  $-3.67^{\circ} \pm 1.01^{\circ}$ , compared to  $-3.34^{\circ} \pm 1.01^{\circ}$  in the control group (Table 2B). Within the case group, individuals who received maxillary first premolar extractions ( $n = 20$ ) had an average  $\Delta$ U1-NA angle of  $1.88^{\circ}$  compared to an average  $\Delta$ U1-NA of  $-6.08^{\circ}$  for individuals in the same group who did not receive maxillary first premolar extractions ( $n = 47$ ). Individuals in the control group who received maxillary first premolar extractions ( $n = 7$ ) had an average  $\Delta$ U1-NA angle of  $8.46^{\circ}$ , compared to  $-4.72^{\circ}$  for those in the control group who did not have maxillary first premolars removed during orthodontic treatment ( $n = 60$ ).

Within the genetic analyses included in this study, no departures from Hardy-Weinberg equilibrium (HWE) were detected within the control population for all of the tested SNPs (data not shown). The complete set of genetic data is summarized in Table 2C. Based on the

final linear regression modeling, variations in the *P2RX7* gene showed the greatest influence on EARR (Table 3). A comparison of the genotypic distributions of the *P2RX7* polymorphisms rs208294 and rs1718119 for the case and control groups is illustrated in Fig. 3A,C, respectively. For the rs208294 polymorphism, patients carrying the CC or CT genotype were more likely to develop EARR than individuals carrying the TT genotype (Table 3;  $p = 0.0028$ ), which is significant at the layered Bonferroni correction of  $p = 0.006$ . The percentage of individuals from each group with a specific *P2RX7* rs208294 genotype is illustrated in Fig. 3B. The percentage of individuals from each group with a specific *P2RX7* rs1718119 genotype is shown in Fig. 3D.

Although findings related to the *IL1B* polymorphism, rs1143634, were not statistically significant in this model ( $p = 0.0533$  compared to the layered Bonferroni corrected significant alpha of  $p = 0.008$ ), *IL1β* likely plays a downstream role in the etiology of EARR given the significance of the *P2RX7* gene for the upstream *P2RX7* receptor on EARR. A comparison of the genotypic distributions of the *IL1B* polymorphism rs1143634 for the case group vs. the control group is illustrated in Fig. 4A. The percentage of individuals from each group with a specific *IL1B* rs1143634 genotype is illustrated in Fig. 4B. Similarly, a comparison of the genotypic distributions of the *CASP1* polymorphism rs580253 is shown by group in Fig. 4C for both the case and control group. The percentage of individuals from each group with a specific *CASP1* rs580253 genotype is shown in Fig. 3D.

In summary, the stepwise logistic regression analysis indicated that the nine modeled variables shown in Table 3 were able to explain ~25.13% of the variation of the EARR among individuals tested. The strongest factor associated with the occurrence of (EARR) was a longer length of treatment, explaining ~10% of the outcome by itself (Table 3). Of the tested genetic variables, the *P2RX7* SNP rs1143634 CC&CT vs. TT genotypes showed a significant difference between affected and control groups. Extraction of the maxillary first premolars also explained a portion of the variation associated with EARR occurrence, although compared to the layered



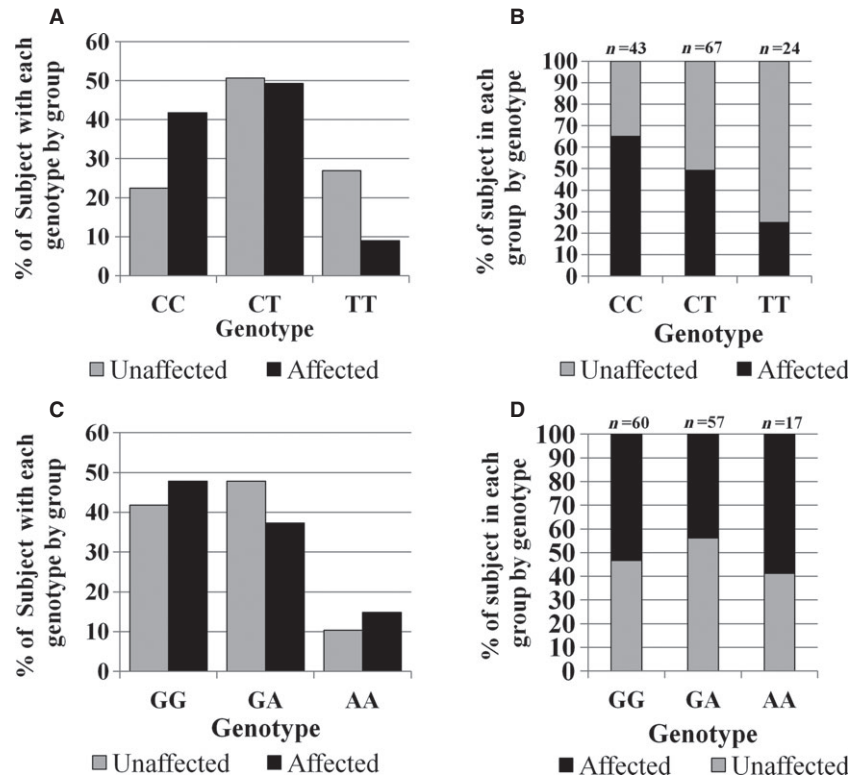


Fig. 3. Genotyping results from *P2RX7* polymorphisms rs208294 and rs1718119 for EARR-affected and unaffected individuals. (A). Genotypic distribution of *P2RX7* SNP rs208294 by group. (B). Percentage of individuals from each group shown by *P2RX7* rs208294 genotype. (C). Genotypic distribution of *P2RX7* SNP rs1718119 by group. (D). Percentage of individuals from each group shown by *P2RX7* rs1718119 genotype.

Bonferroni corrected alpha this variable was not significant by itself (Table 3).

The analysis showed no statistically significant interaction between the length of treatment and the *P2RX7* SNP rs208294 ( $p = 0.8$ ). Similarly, there was no interaction between the extraction of maxillary first premolars and the *P2RX7* SNP rs208294.

## Discussion

Research has aided in identifying many treatment-related factors that influence the occurrence of EARR during orthodontic treatment. It remains a major challenge, however, to resolve why certain subjects are more susceptible to EARR than others. Genetics plays a role in this patient-to-patient difference in susceptibility, with heritability studies indicating genetic variation accounting for approximately two-thirds of the EARR variation seen in orthodontic patients (15, 16).

The novelty of this particular study lays in the multivariate analysis of numerous genetic variables in the IL1B expression pathway, some of which were previously implicated as factors in root resorption, and treatment-related variables

to generate a more comprehensive understanding of their collective influence on the occurrence of EARR concurrent with standard comprehensive 0.018-inch slot edgewise appliance orthodontic treatment. While the influence of age and gender on the etiology of EARR has not consistently been demonstrated in the literature (4, 10, 21, 28), age- and gender-matched case and control groups were utilized in this study to eliminate these potential cofactors from the equation.

Long treatment length and the presence of the CC&CT genotypes for *P2RX7* SNP rs208294 were significantly associated with EARR in this study. Together, the nine cofactors tested in the final linear regression modeling explained ~25.13% of the variation of the EARR observed in the sample tested. Long lengths of treatment proved to be the strongest risk factor for EARR in this study, accounting for ~10% of the variation. This observation is supported by several previously published studies (7, 10, 29). Prolonged stress on the roots increases the chance of root resorption. Therefore, minimizing treatment time when possible or, in some cases, altering the treatment plan to perform a shorter treatment might be the best option.

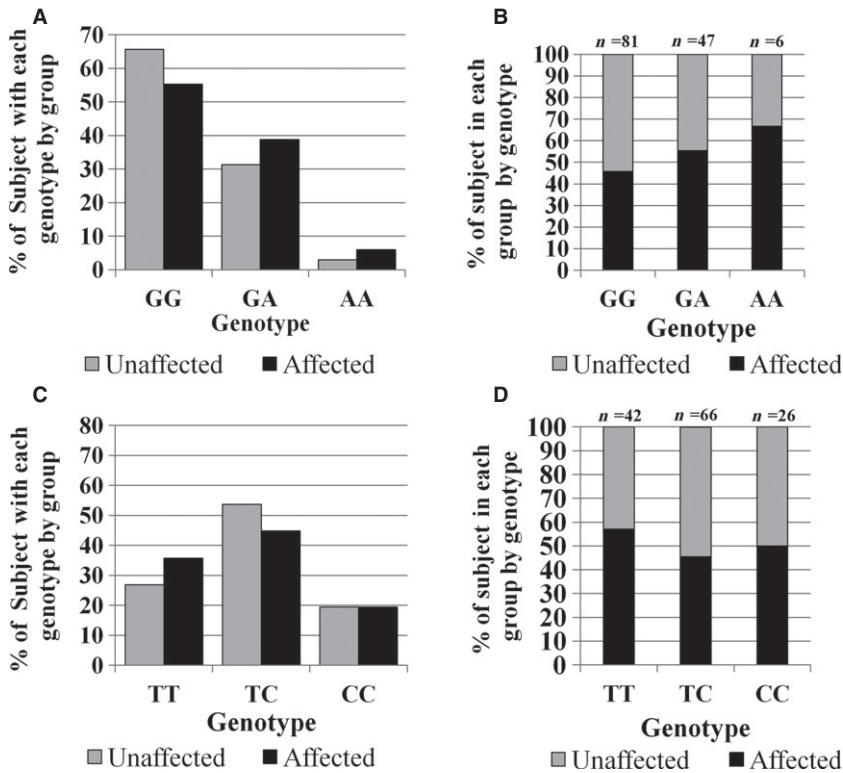


Fig. 4. Genotyping results from *IL1B* SNP rs1143634 and *CASP1* SNP rs580253 for EARR-affected and unaffected individuals. (A). Genotypic distribution of *IL1B* SNP rs1143634 by group. (B). Percentage of individuals from each group shown by *IL1B* rs1143634 genotype. (C). Genotypic distribution of *CASP1* SNP rs530537 by group. (D). Percentage of individuals from each group shown by *CASP1* rs530537 genotype.

Based on mouse gene knockout (KO) experiments, there has been supporting evidence in the literature to suggest that *IL1B* expression pathway has a significant role in the occurrence EARR. In the mouse *Il1b* gene KO model, the absence of *Il1b* was shown to be associated with increased orthodontically induced histologic root resorption of the mouse molars (30, 31). In a separate mouse KO model of the P2x7 receptor (P2rx7), which lies upstream of *Il1b* maturation and release, no difference was observed at baseline between the wild-type ('normal') and KO mice histologic root resorption levels. With the application of force to the teeth of both wild-type and P2rx7 gene KO mice, however, there was a significant increase in histologic root resorption on the KO mouse teeth over the levels of resorption observed in wild-type animals (32, 33).

Pertinent to this study in humans, the second most influential risk factor associated with EARR was the genetic influence of the CC&CT genotypes for the *P2RX7* polymorphism, rs208294. In contrast, the *IL1B* polymorphism rs1143634 A-allele, which had been linked to increased *IL1β* secretion *in vitro* (34), and was thought to be somewhat protective against EARR based on the

initial studies in humans (24, 35, 36), was not significant by itself. It is not clear why this study found that the rs1143634 AA&GA genotypes were associated with EARR, in contrast to previous reports showing that the *IL1B* SNP rs1143634 G-allele was more likely to be associated with orthodontic patients who developed EARR (24, 35, 36).

In a recent published meta-analysis, no association was apparent between EARR and *IL1B* SNP +3954 (37); however, the studies included in the meta-analysis only focused on genetic factors and did not take into account other treatment and environmental factors that could influence the occurrence of EARR in a multivariate analysis. In other studies, the *IL1A* polymorphism rs1800587 was associated with EARR in a German sample, while the rs1143634 *IL1B* SNP was not (38). *IL1B* rs1143634 was reported to be associated with velocity of tooth movement along with *IL1RA*, suggesting a possible connection between velocity of tooth movement, and EARR (39). Taking together the findings of both mouse and human studies, the role of *IL1B* in EARR may be more complicated than first anticipated. Clearly, the role of the *IL1β* expression pathway on influencing EARR warrants further study.

The finding in this study that the genetic factors tested only account for a small amount of the variation associated with EARR when compared to the influence of the length of treatment does not appear to be consistent with the earlier estimation of heritability suggesting that approximately two-thirds of the variation in EARR with orthodontia is due to genetic factors (15, 16). This may be due in part to the inherent limitations of determining heritability in the narrow sense and its application to clinical questions (40), and/or to the limited number of genetic factors that we analyzed for this complex trait. Future studies should look for additional genetic factors that may be involved in orthodontic patient EARR utilizing a non-biased whole-exome or whole-genome sequencing approach when possible instead of the current candidate gene models.

## Conclusion

It can be concluded that increased length of treatment was a strong risk factor of post-orthodontic EARR. The functional SNP, rs208294, located in *P2RX7* gene was also associated with EARR in this model, indicating that the downstream expression of *IL1B* also affects EARR during orthodontic treatment.

## Clinical relevance

A better understanding of the etiology of EARR is needed to improve the ability of predicting

its occurrence. Although EARR may arise in the absence of orthodontic treatment, studies suggest that its incidence may be increased during orthodontic care. When obtaining a patient's consent for orthodontic treatment, clinicians should explain the potential risk factors that lead to EARR and the odds of it occurring. Studies have indicated that two-thirds of the clinical variation in EARR is associated with genetic variation in orthodontic patients. However, determination of these genetic factors is incomplete, requiring studies to include more genetic and clinical factors. By understanding how a combination of both genetic and treatment factors may place a patient at higher risk for EARR during orthodontics, clinicians should be better equipped to explain the risk and minimize treatment-related EARR occurrence.

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## References

1. Killiany DM. Root resorption caused by orthodontic treatment: an evidence-based review of literature. *Semin Orthod* 1999;5:128–33.
2. Remington DN, Joondeph DR, Artun J, Riedel RA, Chapko MK. Long-term evaluation of root resorption occurring during orthodontic treatment. *Am J Orthod Dentofacial Orthop* 1989;96:43–6.
3. Weltman B, Vig KW, Fields HW, Shanker S, Kaizar EE. Root resorption associated with orthodontic tooth movement: a systematic review. *Am J Orthod Dentofacial Orthop* 2010;137:462–76; discussion 12A.
4. Segal GR, Schiffman PH, Tuncay OC. Meta analysis of the treatment-related factors of external apical root resorption. *Orthod Craniofac Res* 2004;7:71–8.
5. Martins DR, Tibola D, Janson G, Maria FR. Effects of intrusion combined with anterior retraction on apical root resorption. *Eur J Orthod* 2012;34:170–5.
6. Parker RJ, Harris EF. Directions of orthodontic tooth movements associated with external apical root resorption of the maxillary central incisor. *Am J Orthod Dentofacial Orthop* 1998;114:677–83.
7. Nanekrungsan K, Patanaporn V, Janhom A, Korwanich N. External apical root resorption in maxillary incisors in orthodontic patients: associated factors and radiographic evaluation. *Imaging Sci Dent* 2012;42:147–54.

8. Acar A, Canyurek U, Kocaaga M, Erverdi N. Continuous vs. discontinuous force application and root resorption. *Angle Orthod* 1999;69:159–63; discussion 63–4.
9. Picanco GV, de Freitas KM, Cancado RH, Valarelli FP, Picanco PR, Feijao CP. Predisposing factors to severe external root resorption associated to orthodontic treatment. *Dental Press J Orthod* 2013;18:110–20.
10. Baumrind S, Korn EL, Boyd RL. Apical root resorption in orthodontically treated adults. *Am J Orthod Dentofacial Orthop* 1996;110:311–20.
11. Horiuchi A, Hotokezaka H, Kobayashi K. Correlation between cortical plate proximity and apical root resorption. *Am J Orthod Dentofacial Orthop* 1998;114:311–8.
12. Linge L, Linge BO. Patient characteristics and treatment variables associated with apical root resorption during orthodontic treatment. *Am J Orthod Dentofacial Orthop* 1991;99:35–43.
13. Owman-Moll P, Kurol J, Lundgren D. Continuous vs. interrupted continuous orthodontic force related to early tooth movement and root resorption. *Angle Orthod* 1995;65:395–401.
14. Kurol J, Owman-Moll P, Lundgren D. Time-related root resorption after application of a controlled continuous orthodontic force. *Am J Orthod Dentofacial Orthop* 1996;110:303–10.
15. Harris EF, Kineret SE, Tolley EA. A heritable component for external apical root resorption in patients treated orthodontically. *Am J Orthod Dentofacial Orthop* 1997;111:301–9.
16. Hartsfield JK, Everett ET, Al-Qawasmī RA. Genetic factors in external apical root resorption and orthodontic treatment. *Crit Rev Oral Biol Med* 2004;15:115–22.
17. Massler M, Malone AJ. Root resorption in human permanent teeth: a roentgenographic study. *Am J Orthod* 1954;40:619–33.
18. Massler M, Perreault J. Root resorption in the permanent teeth of young adults. *J Dent Child* 1954;21:158–64.
19. Newman WG. Possible etiologic factors in external root resorption. *Am J Orthod* 1975;67:522–39.
20. Reitan K. Some factors determining the evaluation of forces in orthodontics. *Am J Orthod* 1957;43:32–45.
21. Sameshima GT, Sinclair PM. Predicting and preventing root resorption: Part I. Diagnostic factors. *Am J Orthod Dentofacial Orthop* 2001;119:505–10.
22. Ferrari D, Pizzirani C, Adinolfi E, Lemoli RM, Curti A, Idzko M et al. The P2X7 receptor: a key player in IL-1 processing and release. *J Immunol* 2006;176:3877–83.
23. Kornman KS, Crane A, Wang HY, Giovine FSD, Newman MG, Pirk FW et al. The interleukin-1 genotype as a severity factor in adult periodontal disease. *J Clin Period* 1997;24:72–7.
24. Al-Qawasmī RA, Hartsfield JK Jr, Everett ET, Flury L, Liu L, Foroud TM et al. Genetic predisposition to external apical root resorption. *Am J Orthod Dentofacial Orthop* 2003;123:242–52.
25. Dinarello CA. Interleukin-1 in the pathogenesis and treatment of inflammatory diseases. *Blood* 2011;117:3720–32.
26. Malmgren O, Goldson L, Hill C, Orwin A, Petrini L, Lundberg M. Root resorption after orthodontic treatment of traumatized teeth. *Am J Orthod* 1982;82:487–91.
27. Darlington RB. *Regression and Linear Models*. New York: McGraw-Hill; 1990.
28. Kjaer I. Morphological characteristics of dentitions developing excessive root resorption during orthodontic treatment. *Eur J Orthod* 1995;17:25–34.
29. DeShields RW. A study of root resorption in treated Class II. Division I malocclusions. *Angle Orthod* 1969;39:231–45.
30. Al-Qawasmī RA, Hartsfield JK, Hartsfield JK Jr, Everett ET, Weaver MR, Foroud TM et al. Root resorption associated with orthodontic force in IL-1Beta knockout mouse. *J Musculoskelet Neuronal Interact* 2004;4:383–5.
31. Hartsfield JK Jr. Pathways in external apical root resorption associated with orthodontia. *Orthod Craniofac Res* 2009;12:236–42.
32. Viecilli RF, Katona TR, Chen J, Hartsfield JK Jr, Roberts WE. Orthodontic mechanotransduction and the role of the P2X7 receptor. *Am J Orthod Dentofacial Orthop* 2009;135:694 e1-16; discussion 94–5.
33. Viecilli R, Katona T, Chen J, Roberts E, Hartsfield J Jr. Comparison of dentoalveolar morphology in WT and P2X7R KO mice for the development of biomechanical orthodontic models. *Anat Rec (Hoboken)* 2009;292:292–8.
34. Pociot F, Molvig J, Wogensens L, Worsaae H, Nerup J. A TaqI polymorphism in the human interleukin-1β (IL-1β) gene correlates with IL-1β secretion *in vitro*. *Eur J Clin Invest* 1992;22:396–402.
35. Bastos Lages EM, Drummond AF, Pretti H, Costa FO, Lages EJ, Gontijo AI et al. Association of functional gene polymorphism IL-1beta in patients with external apical root resorption. *Am J Orthod Dentofacial Orthop* 2009;136:542–6.
36. Iglesias-Linares A. Postorthodontic external root resorption is associated with IL1 receptor antagonist gene variations. *Oral Dis* 2011;18:198–205.
37. Wu FL, Wang LY, Huang YQ, Guo WB, Liu CD, Li SG. Interleukin-1beta +3954 polymorphisms and risk of external apical root resorption in orthodontic treatment: a meta-analysis. *Genet Mol Res* 2013;12:4678–86.
38. Glden N, Eggermann T, Zerres K, Beer M, Meinelt A, Diedrich P. Interleukin-1 polymorphisms in relation to External Apical Root Resorption (EARR). *J Orofacial Orthop* 2009;70:20–38.
39. Iwasaki LR, Gibson CS, Crouch LD, Marx DB, Pandey JP, Nickels JC. Speed of tooth movement is related to stress and IL-1 gene polymorphisms. *Am J Orthod Dentofacial Orthop* 2006;130:e1-698.e9.
40. Harris EF. Interpreting heritability estimates in the orthodontic literature. *Semin Orthod* 2008;14:125–34.

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